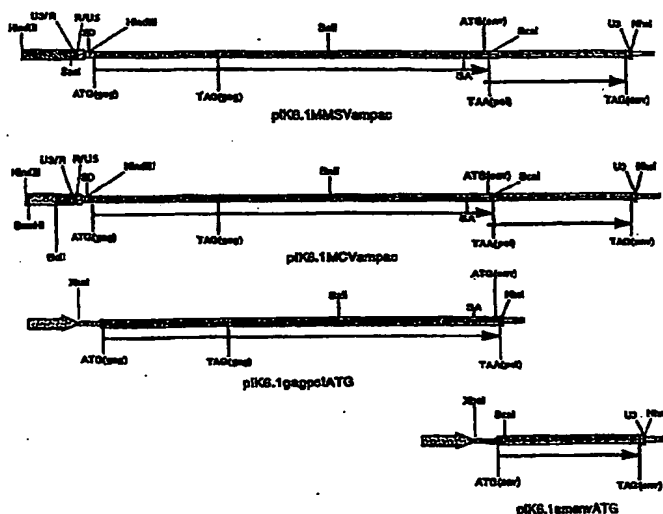




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<p>(21) International Application Number: PCT/US96/13737</p> <p>(22) International Filing Date: 21 August 1996 (21.08.96)</p> <p>(30) Priority Data: 517,488 21 August 1995 (21.08.95) US</p> <p>(71) Applicant: CELL GENESYS, INC. [US/US]; 322 Lakeside Drive, Foster City, CA 94404 (US).</p> <p>(72) Inventors: FINER, Mitchell, H.; 54 Madera, San Carlos, CA 94070 (US). DULL, Thomas, J.; 1850 Great Highway, San Francisco, CA 94122 (US). ZSEBO, Krisztina, M.; 200 Allen Road, Woodside, CA 94062 (US). COOKE, Keegan; 430 Olive Avenue, Palo Alto, CA 94306 (US). FARSON, Deborah, A.; 521 62nd Street, Oakland, CA 94609 (US).</p> <p>(74) Agents: HALLUIN, Albert, P. et al.; Pennie & Edmonds, 1155 Avenue of the Americas, New York, NY 10036 (US).</p>		<p>(81) Designated States: AL, AM, AU, AZ, BB, BG, BR, BY, CA, CN, CU, CZ, EE, FI, GE, HU, IL, IS, JP, KG, KP, KR, KZ, LK, LR, LS, LT, LV, MD, MG, MK, MN, MX, NO, NZ, PL, RO, RU, SG, SI, SK, TJ, TM, TR, TT, UA, UZ, VN, ARIPO patent (KE, LS, MW, SD, SZ, UG), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG).</p> <p>Published <i>Without international search report and to be republished upon receipt of that report.</i></p>

(54) Title: HIGH EFFICIENCY RETROVIRAL PACKAGING SYSTEM



(57) Abstract

The invention provides a novel retroviral packaging system, in which retroviral packaging plasmids and packageable vector transcripts are produced from high expression plasmids after stable or transient transfection in mammalian cells. High titers of recombinant retrovirus are produced in these transfected mammalian cells and can then transduce a mammalian target cell by cocultivation or supernatant infection. The methods of the invention include the use of the novel retroviral packaging plasmids and vectors to transduce primary human cells, including T cells and human hematopoietic stem cells, with foreign genes by cocultivation or supernatant infection at high efficiencies. The invention is useful for the rapid production of high titer viral supernatants, and to transduce with high efficiency cells that are refractory to transduction by conventional means.

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HIGH EFFICIENCY RETROVIRAL PACKAGING SYSTEM

5 Mitchell H. Finer, Thomas J. Dull,
Krisztina M. Zsebo, Keegan Cooke and Deborah A. Farson

CROSS-REFERENCE TO RELATED APPLICATIONS

10 This application is a continuation-in-part of
Application Serial No. 08/258,152 filed June 10, 1994 which
is a continuation-in-part of Application Serial No.
08/076,299, filed June 11, 1993, the disclosures of both
application are incorporated by reference in their entirety
15 herein.

FIELD OF THE INVENTION

This invention relates to novel retrovirus
20 packaging plasmids and vectors, to their use in the
production of recombinant retrovirus in mammalian cells, and
to methods of using such constructs to transduce mammalian
target cells with high efficiency. The invention also
relates to the construction of stable cell lines in which
25 novel retroviral packaging plasmids and/or vectors are stably
expressed in viral packaging cell lines.

BACKGROUND OF THE INVENTION

30 Retrovirus vectors have become the primary tool for
gene delivery in human gene therapy applications (Miller,
Nature 357:455-460 (1992)). The ability of retrovirus
vectors to deliver an unarranged, single copy gene into a
broad range of rodent, primate and human somatic cells in
35 primary culture makes them well suited for this purpose.
Identification and subsequent deletion of the sequences
present within retroviral transcripts encoding the packaging

signals for avian (E) and murine (ψ) retroviruses, has enabled development of packaging cell lines to supply in trans the proteins necessary for production of infectious virions, but render the packaging cell lines unable to

5 package their own viral genomic mRNA (Watanabe and Temin, Molec. Cell. Biol. 3(12):2241-2249 (1983); Mann et al., Cell 33:153-159 (1983); and Embretson and Temin, J. Virol. 61(9):2675-2683 (1987)). The most important consideration in the construction of retroviral packaging lines has been both

10 the production of high titer vector supernatants free of recombinant replication competent retrovirus, which has been shown to produce T cell lymphomas in rodents (Cloyd et al., J. Exp. Med. 151, 542-552 (1980)) and primates (Donahue et al., J. Exp. Med. 176, 1125-1135 (1992)). Although early murine

15 retroviral packaging lines were highly prone to generation of replication competent retrovirus (RCR) (Cone and Mulligan, Proc. Nat'l. Acad. Sci. USA 81:6349-6353 (1984)) or prone to co-package the ψ -genome (Mann et al., supra, 1983; Buttimore and Miller, Mol. Cell. Biol. 6(8):2895-2902 (1986)), two

20 strategies have evolved for the construction of second generation packaging lines with significantly reduced ability for the generation of RCR. One strategy, embodied by PA317, uses a single genome packaging construct from which the initiation site for second strand synthesis, the 3' LTR, and

25 the ψ site have been deleted (Miller and Buttimore, Molec. Cell. Biol. 6(8): 2895-2902 (1986)). These modifications eliminate as much as possible homology between the packaging genome and the viral vector to reduce the ability to form recombinants, and have resulted in production of high titer,

30 helper-free virus with many vector systems (Miller and Rosman, BioTechniques 7(9):980-990 (1989)). The second approach has been to divide the packaging functions into two genomes: one that expresses the gag and pol gene products, and the other that expresses the env gene product (Bosselman

35 et al., Molec. Cell. Biol. 7(5):1797-1806 (1987); Markowitz et al., J. Virol. 62(4): 1120-1124 (1988); Danos and Mulligan, Proc. Nat'l. Acad. Sci. (USA) 85:6460-6464 (1988)).

This approach eliminated the ability for co-packaging and subsequent transfer of the ψ -genome, as well as significantly decreased the frequency of recombination due to the presence of three retroviral genomes in the packaging cell that must undergo recombination to produce RCR. In the event recombinants arise, mutations (Danos and Mulligan, supra) or deletions (Bosselman et al., supra; and Markowitz et al., supra) within the undesired gene products render recombinants non-functional. In addition, deletion of the 3' LTR on both packaging function constructs further reduces the ability to form functional recombinants. Although early attempts at the generation of two genome packaging lines yielded low titer producer clones (Bosselman et al., supra) producer lines are now available that yield high titer producer clones (Danos and Mulligan, supra; and Markowitz et al., supra).

Packaging lines currently available yield producer clones of sufficient titer to transduce human cells for gene therapy applications and have led to the initiation of human clinical trials (Miller, supra). However, there are two areas in which these lines are deficient. First, design of the appropriate retroviral vectors for particular applications requires the construction and testing of several vector configurations. For example, Belmont et al., Molec. and Cell. Biol. 8(12):5116-5125 (1988), constructed stable producer lines from 16 retroviral vectors in order to identify the vector capable of producing both the highest titer producer and giving optimal expression. Some of the configurations examined included: (1) LTR driven expression vs. an internal promoter; (2) selection of an internal promoter derived from a viral or a cellular gene; and (3) whether a selectable marker was incorporated in the construct. A packaging system that would enable rapid, high-titer virus production without the need to generate stable producer lines would be highly advantageous in that it would save approximately two months required for the identification

of high titer producer clones derived from several constructs.

Second, compared to NIH 3T3 cells, the infection efficiency of primary cultures of mammalian somatic cells with a high titer amphotropic retrovirus producer varies considerably. The transduction efficiency of mouse myoblasts (Dhawan et al., Science 254:1509-1512(1991) or rat capillary endothelial cells (Yao et. al., Proc. Natl. Acad. Sci. USA 88:8101-8105 (1991)) was shown to be approximately equal to that of NIH 3T3 cells, whereas the transduction efficiency of canine hepatocytes (Armentano et. al., Proc. Natl. Acad. Sci. USA 87:6141-6145 (1990)) was only 25% of that found in NIH 3T3 cells. Primary human tumor-infiltrating lymphocytes ("TILs"), human CD4+ and CD8+ T cells isolated from peripheral blood lymphocytes, and primate long-term reconstituting hematopoietic stem cells, represent an extreme example of low transduction efficiency compared to NIH 3T3 cells. Purified human CD4+ and CD8+ T Cells have been reported on one occasion to be infected to levels of 6%-9% with supernatants from stable producer clones (Morecki et al., Cancer Immunol. Immunother. 32:342-352 (1991)), and primate or human long-term reconstituting hematopoietic stem cells have only been infected to $\leq 1\%$ with a producer of titer of 10^6 per ml on NIH 3T3 cells (van Beusechem et al., Proc. Natl. Acad. Sci. USA 89:7640-7644 (1992); and Donahue et al., supra). If the retrovirus vector contains the neo^r gene, populations that are highly enriched for transduced cells can be obtained by selection in G418. However, selectable marker expression has been shown to have deleterious effects on long-term gene expression in vivo in hematopoietic stem cells (Apperly et.al. Blood 78:310-317(1991)).

An approach that yields significantly increased transduction of mammalian cells in primary culture would be highly advantageous, and this need is currently unmet.

SUMMARY OF THE INVENTION

Accordingly, the present invention provides novel plasmid based expression vectors that direct the synthesis of both packagable retroviral vector transcripts and retroviral gene products required for rapid production of high titer recombinant retrovirus in human cells by transient transfection, thereby eliminating the need to generate stable producer lines. In addition, the invention provides a method for highly efficient transduction of mammalian cells that have previously been described as difficult to transduce with retroviral constructs. The invention also describes the construction of cell lines in which the plasmid-based expression vectors of the invention that direct the synthesis of retroviral gene products required in trans for virus production have been stably integrated into the genome of the producing cells. This invention also describes the construction of retroviral vector plasmids with sequences enabling the episomal persistence retroviral vectors of the invention without the need for stable integration of the vector plasmid. All of these stably transfected lines can be used to generate stable cell lines that continuously produce recombinant retrovirus at high titer.

The retroviral constructs are packaging plasmids consisting of at least one retroviral helper DNA sequence derived from a replication-incompetent retroviral genome encoding in trans all virion proteins required to package a replication incompetent retroviral vector, and for producing virion proteins capable of packaging the replication-incompetent retroviral vector at high titer, without the production of replication-competent helper virus. The retroviral DNA sequence lacks the region encoding the native enhancer and/or promoter of the viral 5'LTR of the virus, and lacks both the psi function sequence responsible for packaging helper genome and the 3' LTR, but encodes a foreign polyadenylation site, for example the SV40 polyadenylation

site, and a foreign enhancer and/or promoter which directs efficient transcription in a cell type where virus production is desired. The retrovirus is a leukemia virus such as a Moloney Murine Leukemia Virus (MMLV), the Human

5 Immunodeficiency Virus (HIV), or the Gibbon Ape Leukemia virus (GALV). The foreign enhancer and promoter may be the human cytomegalovirus (HCMV) immediate early (IE) enhancer and promoter, the enhancer and promoter (U3 region) of the Moloney Murine Sarcoma Virus (MMSV), the U3 region of Rous

10 Sarcoma Virus (RSV), the U3 region of Spleen Focus Forming Virus (SFFV), or the HCMV IE enhancer joined to the native Moloney Murine Leukemia Virus (MMLV) promoter. The retroviral packaging plasmid may consist of two retroviral helper DNA sequences encoded by plasmid based expression

15 vectors, for example where a first helper sequence contains a cDNA encoding the gag and pol proteins of ecotropic MMLV or GALV and a second helper sequence contains a cDNA encoding the env protein. The Env gene, which determines the host range, may be derived from the genes encoding xenotropic,

20 amphotropic, ecotropic, polytropic (mink focus forming) or 10A1 murine leukemia virus env proteins, or the Gibbon Ape Leukemia Virus (GALV env protein, the Human Immunodeficiency Virus env (gp160) protein, the Vesicular Stomatitis Virus (VSV) G protein, the Human T cell leukemia (HTLV) type I and

25 II env gene products, chimeric envelope gene derived from combinations of one or more of the aforementioned env genes or chimeric envelope genes encoding the cytoplasmic and transmembrane of the aforementioned env gene products and a monoclonal antibody directed against a specific surface

30 molecule on a desired target cell.

Specific embodiments of the retroviral packaging plasmids of the invention include: pIK6.1MMSVampac, pIK6.1MCVampac, pIK6.1gagpolATG and pIK6.1amenvATG.

35

The invention includes retroviral vectors that contain a modified 5' LTR, which enables efficient

transcription of packagable vector transcripts in the desired cell type. In addition, the invention includes retroviral constructs encoding foreign genes.

5 In one method of the invention, the packaging plasmids and retroviral vectors are transiently cotransfected into a first population of mammalian cells that are capable of producing virus, such as human embryonic kidney cells, for example 293 cells (ATCC, Rockville, MD) to produce high titer
10 recombinant retrovirus-containing supernatants. In another method of the invention this transiently transfected first population of cells is then cocultivated with mammalian target cells, for example human lymphocytes, to transduce the target cells with the foreign gene at high efficiencies. In
15 yet another method of the invention the supernatants from the above described transiently transfected first population of cells are incubated with mammalian target cells, for example human lymphocytes or hematopoietic stem cells, to transduce the target cells with the foreign gene at high efficiencies.
20

 In yet another method of the invention, the packaging plasmids (either single or double genome) are transiently cotransfected with a retroviral vector plasmid into a first
25 population of mammalian cells, for example 293 cells, to produce high titer recombinant retrovirus containing supernatants.

 In still yet another method of the invention, the
30 packaging plasmids are stably expressed in a first population of mammalian cells that are capable of producing virus, such as human embryonic kidney cells, for example 293 cells. Retroviral vectors are introduced into cells by either cotransfection with a selectable marker or infection with
35 pseudotyped virus. In both cases, the vectors integrate. Alternatively, vectors can be introduced in an episomally

maintained plasmid. High titer recombinant retrovirus-containing supernatants are produced.

The invention further includes mammalian target
5 cells expressing a foreign gene produced by any of the above methods of the invention. The foreign gene may be a chimeric T cell receptor such as a CD4/zeta or single-antibody chain/zeta T cell receptor, for example.

10

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1A, B and C is a diagrammatic representation of the retroviral packaging plasmids of the invention used to produce the proteins necessary to package retroviral vector
15 transcripts: pIK6.1MMSVampac, pIK6.1MCVampac, pIK6.1gagpolATG, and pIK6.1envATG.

Figure 2 shows the FACS profile of 293 cells transfected with retroviral constructs, as described in
20 Example I, infra.

Figure 3 shows the transduction efficiency determined by Southern blot analysis of infected 3T3 DNA, as described in Example I, infra.

25

Figure 4 is a bar graph of the data from experiments in which CD8+ T cells were transduced by, first, transient transfection of 293 cells with either pRTD2.2F3 or pRTD2.2F15 and pIK6.1MCVampac, followed by cocultivation of
30 the 293 cells with the CD8+ T cells and analysis of transduction efficiency by FACS, as described in Example II, infra.

Figure 5 shows the results of FACS analysis of
35 hematopoietic stem cells transduced with the KAT packaging constructs and cocultivation with 293 cells, as described in Example III, infra.

Figure 6 examines whether the cocultivation of CD34+ cells with KAT transfected 293 cells leads to high efficiency transduction as analyzed by Southern blotting, as described in Example III, infra.

5

Figure 7 compares the transduction efficiency of CD34+ cells transduced by the KAT system to that of cocultivation with a stable PA317 producer by Southern blotting, as described in Example III, infra.

10

Figure 8 shows the results of FACS analysis of human CD34+ hematopoietic progenitors transduced with the KAT pIKT retrovirus vector constructs following transfection of 293 cells with pIKT vectors and cocultivation, as described in Example IV, infra.

DETAILED DESCRIPTION OF THE INVENTION

20 In order that the invention herein described may be more fully understood, the following description is set forth.

The present invention provides novel optimized transient expression plasmids (designated "KAT") for production of retroviral virions in which high steady state levels of retroviral packaging functions and packagable vector transcripts are produced following introduction of KAT plasmids into mammalian cells capable of efficient transient transfection and expression, in the absence of plasmid replication of viral vector and packaging function plasmids. The absence of plasmid replication enables production of high titer virions while minimizing the potential for production of replication competent retrovirus by recombination. Use of the KAT system yields 10-30 fold higher viral titers compared to cotransfection of packaging functions and vector plasmids into COS cells, as described by Landau and Litman, J. Virol.

66(8):5110-5113 (1992)). Alternatively, because the KAT packaging function and viral vector plasmids contain the SV40 origin of replication, they can be transfected into cell lines that enable replication of SV40 origin-containing
5 plasmids due to expression of the SV40 T antigen, such as tsa201 (Heinzel et al., J. Virol. 62(10):3738-3746 (1988)). Using the KAT system, viral titers in the presence of plasmid replication are 3 to 10-fold higher than in the absence of replication. Whether replicating or nonreplicating plasmids
10 are used, the KAT system permits the rapid production of high titer recombinant retrovirus supernatants without the need for generating stable producer lines.

The retroviral constructs of the invention also
15 find use in the method of the invention to transduce by cocultivation or by supernatant infection, with high efficiency, mammalian cells, such as primary human cells, that are typically refractory to transduction by conventional means.

20

The plasmids of the invention also find use in the construction of stable cell lines that constitutively produce the retroviral proteins required in trans for the production of retrovirus particles: gag, pol and env. These stable
25 packaging constructs can be introduced into human cell lines by calcium phosphate transfection or electroporation, together with a dominant selectable marker, such as neo, DHFR*, Gln Synthetase, ADA, followed by selection in the appropriate drug and isolation of clones. This enables the
30 production of high titer stable producer clones following introduction of a retroviral construct into these cells. These cell lines have all of the same properties of the transiently transfected producer cells. However, due to stable integration of both packaging function and virus
35 vector, they continue to produce high titer retrovirus indefinitely in the absence of drug selection.

Plasmids containing the packaging functions can be split with one encoding the gag and pol genes and a second encoding the env gene product. Packaging lines containing two viral genomes have been described (Bosselman et al., Molec. Cell. Biol. 7(5):1797-1806 (1987); Markowitz et al., J. Virol. 62(4): 1120-1124 (1988); Danos and Mulligan, Proc. Natl. Acad. Sci. (USA) 85:6460-6464 (1988)) and are desirable due to the significantly reduced chance for the generation of replication competent retrovirus (RCR) following

10 recombination between a retroviral vector and packaging construct. Use of the plasmids of the invention results in a packaging line yielding the high efficiency transduction of the transient system. The novel plasmids of the invention enable a significant advance over previously described two

15 genome packaging lines. The KAT plasmids encoding gagpol and env genes have been constructed so that only protein coding sequence from the retroviral genome is present. Using the retroviral vectors described in the invention, no overlap exists between the retrovirus vector and packaging genomes at

20 their 3' ends. This structure in combination with replacement of the gag start codon (ATG) in the vector with a stop codon absolutely precludes the generation of replication competent retrovirus in contrast to previously described packaging lines where complete viral genomes containing

25 mutations (Danos and Mulligan, Proc. Nat'l. Acad. Sci. (USA) 85:6460-6464 (1988)) or deletions (Bosselman et al., Molec. Cell. Biol. 7(5):1797-1806 (1987); Markowitz et al., J. Virol. 62(4): 1120-1124 (1988)). These prior known packaging lines contain overlap at the 3' end of the virus vector with

30 the packaging line and can potentially generate RCR.

Two genome packaging lines are constructed by sequential introduction of the gagpol plasmid followed by the env- containing plasmid. The env genes are responsible for

35 recognition of cell surface receptors. Five functionally and structurally different env genes have been identified in murine leukemia viruses and have been shown to have

genetically distinct receptors (Battini et al., J. of Virol. 66:1468-1475 (1992)). Human host range with murine leukemia virus vectors is possible by the introduction of the amphotropic env gene into a cell line that expresses the
5 ecotropic MLV gagpol (Danos and Mulligan, Proc. Nat'l. Acad. Sci. USA 85:6460-6464 (1988)). The xenotropic and 10A1 MLV viruses have human host range, as well as the gibbon ape leukemia and feline leukemia viruses. Using cDNA clones of these env genes, one or more can be stably introduced into a
10 gagpol line to create a packaging line where the retrovirus produced following introduction of a retroviral vector can enter the target through multiple genetically distinct receptors. This leads to substantial increases in apparent viral titer. The vectors of the invention provide for the
15 ability to create these types of novel packaging lines.

The expression plasmids for gag/pol and env contain a functional poly A addition signal (poly A site) which is essential for transcription termination by RNA polymerase II
20 (Connelly and Manley Genes Dev. 2:440-452 (1988)). The poly A site may be derived from a viral transcription unit, a cellular gene, or a synthetic oligonucleotide. Examples of viral poly adenylation sites include the SV40 early region poly A site (Fitzgerald and Shenk, Cell 24:251-260 (1981))
25 or the hepatitis B surface antigen poly A site (Simonsen and Levinson, Mol. Cell., Bio. 3:2250-2258 (1983)). Examples of polyadenylation signals derived from cellular genes include human pro alpha 2(1) collagen (Myers et. al., J. Biol. Chem. 258:10128-10135 (1983)), bovine growth hormone
30 (Woychik et al., Proc. Natl. Acad. Sci., USA 81:3944-3988 (1984)) and the human alpha globin gene (Orkin et. al., 4:453-456 (1985)). An example of an efficient synthetic polyadenylation site is the sequence AATAAA(N)22-23(GT)n(T)N (Levitt et. al., Genes Dev. 3:1019-1025 (1989)). One skilled
35 in the art may substitute any of the above polyadenylation signals for the SV40 poly A signal used in the instant plasmids by using conventional techniques.

The techniques used to construct vectors, and transfect and infect cells, are widely practiced in the art, and most practitioners are familiar with the standard resource materials which describe specific conditions and procedures. However, for convenience, the following paragraphs may serve as a guideline.

Construction of the vectors of the invention employs standard ligation and restriction techniques which are well understood in the art (see Maniatis et al., in Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory, N.Y. (1982)). Isolated plasmids, DNA sequences, or synthesized oligonucleotides are cleaved, tailored, and religated in the form desired.

Site-specific DNA cleavage is performed by treating with the suitable restriction enzyme (or enzymes) under conditions which are generally understood in the art, and the particulars of which are specified by the manufacturer of these commercially available restriction enzymes. (See, e.g. New England Biolabs, Product Catalog.) In general, about 1 μ g of plasmid or DNA sequences is cleaved by one unit of enzyme in about 20 μ l of buffer solution. Typically, an excess of restriction enzyme is used to insure complete digestion of the DNA substrate. Incubation times of about one hour to two hours at about 37°C are workable, although variations can be tolerated. After each incubation, protein is removed by extraction with phenol/chloroform, and may be followed by ether extraction, and the nucleic acid recovered from aqueous fractions by precipitation with ethanol. If desired, size separation of the cleaved fragments may be performed by polyacrylamide gel or agarose gel electrophoresis using standard techniques. A general description of size separations is found in Methods of Enzymology 65:499-560 (1980).

Restriction cleaved fragments may be blunt ended by treating with the large fragment of *E. coli* DNA polymerase I (Klenow) in the presence of the four deoxynucleotide triphosphates (dNTPs) using incubation times of about 15 to 5 25 minutes at 20°C in 50 mM Tris (pH 7.6) 50 mM NaCl, 6 mM MgCl₂, 6 mM DTT and 5-10 µM dNTPs. The Klenow fragment fills in at 5' sticky ends but chews back protruding 3' single strands, even though the four dNTPs are present. If desired, selective repair can be performed by supplying only one of 10 the dNTPs, or with selected dNTPs, within the limitations dictated by the nature of the sticky ends. After treatment with Klenow, the mixture is extracted with phenol/chloroform and ethanol precipitated. Treatment under appropriate conditions with S1 nuclease or Bal-31 results in hydrolysis 15 of any single-stranded portion.

Ligations are performed in 15-50 µl volumes under the following standard conditions and temperatures: 20 mM Tris-Cl pH 7.5, 10 mM MgCl₂, 10 mM DTT, 33 mg/ml BSA, 10 mM-50 20 mM NaCl, and either 40 µM ATP, 0.01-0.02 (Weiss) units T4 DNA ligase at 0°C (for "sticky end" ligation) or 1 mM ATP, 0.3-0.6 (Weiss) units T4 DNA ligase at 14°C (for "blunt end" ligation). Intermolecular "sticky end" ligations are usually performed at 33-100 µg/ml total DNA concentrations (5-100 mM 25 total end concentration). Intermolecular blunt end ligations (usually employing a 10-30 fold molar excess of linkers) are performed at 1 µM total ends concentration.

The retroviral vectors and packaging plasmids of 30 the KAT system are prepared as follows:

Production of novel retroviral vectors and packaging plasmids

The KAT constructs include DNA packaging plasmids 35 consisting of at least one retroviral helper DNA sequence derived from a replication-incompetent retroviral genome, e.g. a leukemia virus genome, encoding in trans all virion

proteins required to package a replication incompetent retroviral vector, and for producing virion proteins capable of packaging the replication-incompetent retroviral vector at high titer, without the production of replication-competent 5 helper virus. In one embodiment the retroviral packaging DNA sequence lacks the region encoding the native enhancer and/or promoter of the viral 5'LTR of the virus, and lacks the psi function sequence responsible for packaging helper genome as well as the 3' LTR, but encodes a foreign enhancer and/or 10 promoter which directs efficient transcription in a cell type where virus production is desired, and includes an SV40 polyadenylation site. The transcription initiation site of the foreign enhancer and promoter is joined to the leukemia virus genome at the 5' end of the "R" region of the 5' LTR.

15

The retrovirus may be a Moloney Murine Leukemia Virus (MMLV), the Human Immunodeficiency Virus (HIV) or the Gibbon Ape Leukemia virus (GALV). The foreign enhancer and promoter joined to the R region of the 5' LTR may be the 20 human cytomegalovirus (HCMV) immediate early (IE) enhancer and promoter (the U3 region) of the Moloney Murine Sarcoma Virus (MMSV), the U3 region of the Rous Sarcoma Virus (RSV), the U3 region of the Spleen Focus Forming Virus (SFFV), or the HCMV IE enhancer joined to the native Moloney Murine 25 Leukemia Virus (MMLV) promoter.

All psi (ψ)-packaging plasmids are derivatives of the plasmid pIK1.1. pIK1.1 is a mammalian expression vector constructed by four successive insertions into pMF2, which is 30 created by inserting the synthetic polylinker 5'-HindIII-SphI-EcoRI-AatII-BglI-XhoI-3' into KpnI and SacI sites of pSKII (Stratagene, San Diego, CA), with loss of the Kpn I and Sac I sites. First, a BamHI-XbaI fragment containing the SV40 T antigen polyadenylation site (nucleotides 2770 to 2533 35 of SV40, Reddy *et al.*, Science 200:494-502 (1978)) and an NheI-SalI fragment containing the SV40 origin of replication (nucleotides 5725 to 5578 of SV40) are inserted by three-part

ligation between the BglII and XhoI sites, with the loss of the BglII, BamHI, XbaI, NheI, SalI and XhoI sites. These BamHI-XbaI and NheI-SalI fragments are synthesized by PCR with pSV2neo (Southern and Berg, J. Mol. Appl. Gen. 1:327-341 5 (1982)) as the template using oligonucleotide primer pairs 3 and 4, and 5 and 6, respectively, which incorporated BamHI, XbaI, NheI and SalI sites at their respective ends. Second, an SphI-EcoRI fragment containing the splice acceptor of the human $\alpha 1$ globin gene second exon (nucleotides +143 to +251) 10 is inserted between the SphI and EcoRI sites. This SphI-EcoRI fragment is synthesized by PCR with p π SV α HP (Treisman et al., Proc. Natl. Acad. Sci. USA 80:7428-7432 (1983)) as the template using oligonucleotide primers 7 and 8, which incorporate SphI and EcoRI sites at their respective ends. 15 Third, the synthetic polylinker 5'-EcoRI-BglII-NcoI-ApaI-AatII-3' is inserted between the EcoRI and the AatII sites. Fourth, a HindIII-SacI fragment containing the CMV IE enhancer/promoter (nucleotides -674 to -19, Boshart et al., Cell 41:521-530 (1985)) and a chemically synthesized SacI- 20 SphI fragment containing the CMV IE first exon/splice donor (nucleotides -19 to +170) are inserted by three-part ligation between the HindIII and SphI sites. The HindIII-SacI fragment is prepared by PCR with pCDM8 (Seed, Nature 329:840-842 (1987); Seed and Aruffo, Proc. Natl. Acad. Sci. USA 25 84:3365-3369 (1987)) as the template using oligonucleotide primers 9 and 10, which incorporated HindIII and SacI sites at their respective ends.

Primer 3: 5'- GGTCGACCTGGATCCGCCATACCACATTTGTAG -3' (SEQ ID 30 NO. 1)

Primer 4: 5'- GCCGCGGCTCTAGAGCCAGACATGATAAGATAC -3' (SEQ ID NO. 2)

Primer 5: 5'- AAGCTTGTGCTAGCTATCCCGCCCCTAACTCCG-3' (SEQ ID NO. 3)

35 Primer 6: 5'- CGAAATCGGTCGACCGCAAAGCCTAGGCCTCC-3' (SEQ ID NO. 4)

Primer 7: 5'- GTCTATAGCATGCTCCCCTGCTCCGACCCG-3'

(SEQ ID NO. 5)

Primer 8: 5'- GGTACCGAATTCTCCTGCGGGGAGAAGCAG-3'

(SEQ ID NO. 6)

Primer 9: 5'- CGCCAAGCTTGGCCATTGCATACGGT-3'

5 (SEQ ID NO. 7)

Primer 10: 5'- GAGGTCTAGACGGTTCATAACGAGCTCT-3'

(SEQ ID NO. 8)

An Xba I site is introduced at the transcription
10 initiation site of the HCMV IE promoter in pIK1.1 by
replacement of the chemically synthesized Sac I / Sph I
oligonucleotide encoding -19 to +170, described above, with a
chemically synthesized Sac I/Sph I oligonucleotide where an
Xba I site at nucleotides +1 to +6 had been introduced to
15 generate pIK6.1. This allows insertion of any
enhancer/promoter as a Hind III to Xba I cassette so as to
insert the appropriate enhancer and promoter that will direct
the highest possible expression level of the desired
sequences in the desired cell type. In order to obtain the
20 highest expression levels in mouse fibroblast NIH 3T3 (ATCC
CRL 1658) or M. dunni (ATCC CRL2017), the complete MMSV U3
region was synthesized by PCR using the plasmid pN7 (Miller
et al., Mol. Cell. Biol. 5:431-437 (1985)) as a template and
two primers: one which encoded a HindIII site and the 5' 21
25 nucleotides of the U3, and a second which encoded the 3' 21
nucleotides of the MMLV U3 region and an Xba I site. This PCR
fragment was cloned between the HindIII and Xba I sites of
pIK6.1 to generate pIK6.1MMSV. In order to direct high level
expression in human cells, pIK6.1MCV was constructed by
30 isolation of the Nco I/Spe I fragment of the HCMV IE enhancer
(Boshart et al., supra), addition of synthetic
oligonucleotide Bcl I linkers, and insertion in the Bam HI
site of the plasmid pΔHB (Dr. P. Robbins, University of
Pittsburgh, Pittsburgh, PA). This plasmid was designated
35 pMCV. pΔHB is a plasmid in which the ClaI to EcoRI fragment
of pZIPneoSVX (Cepko et.al, supra), containing viral
sequences including the 3' LTR, has been cloned into the ClaI

and Eco RI sites of pBR322 and where the Sau 3AI to Hpa II enhancer fragment of MMLV U3 has been removed. Due to the homology between the MMLV U3 and the MMSV U3, the PCR primers described above were used to generate a Hind III/Xba I linker
5 fragment encoding the U3 fragment of pMCV, which was cloned into pIK6.1 to generate pIK6.1MCV. These plasmids, as well as pIK6.1, were further modified by deletion of 112 nucleotides of the SV40 polyadenylation site between the ApaI site at the 3' end of the pIK polylinker and the Hpa I site in the SV40
10 polyadenylation site and replacement with an Nhe I linker to create pIK6.1.Nhe, pIK6.1.MMSV.Nhe and pIK.6.1MCV.Nhe.

pIK6.1MMSVampac and pIK6.1MCVampac were constructed by insertion of 3813 base Sac I/Sal fragment encoding a
15 portion of the U3 region, the R, and U 5 regions, the gag gene and a portion of the pol gene of pMOV psi- (Mann et al., supra), and the 4140 base pair Sal I-Nhe I fragment encoding pol/env, derived from pCRIPamgag-2 (Danos and Mulligan, Proc. Natl. Acad. Sci. USA, 85:6460-6464 (1988) between the Sac I
20 and Nhe I sites of pIK6.1MMSV.Nhe or pKI6.1MCV.Nhe, respectively. pCRIPamgag-2 is a derivative of pCRIPamgag where the pBR322 plasmid backbone has been replaced by the plasmid pUC19. The resulting plasmids encode the gag and pol genes from ecotropic MMLV and the envelope gene from the
25 4070A amphotropic MLV (Chattopadhyay et al., J. Virol. 39(3):777-791 (1981)) and are diagramed in Figure 1A.

To delete untranslated sequences 3' from the envelope gene of pIK6.1MCVampac a PCR reaction was performed
30 using pIK6.1MCVampac as the template with synthetic oligonucleotides 5' CTGATCTTACTCTTTGGACC3' (SEQ ID NO. 31) and 5'GAATTCGCTAGCCTATGGCTCGTACTCTATAG 3' (SEQ ID NO. 32). The resulting 142 basepair PCR product was cut with ClaI and NheI. This 100 base pair fragment was excised and used to
35 replace the corresponding 172 base pair ClaI to NheI fragment of pIK6.1MCVampac to give pIK6.1MCVampac UTA.

pIK6.1amenvATGUTΔ was constructed by replacing the 172 base pair ClaI to NheI fragment in pIK6.1amenvATG with the 100 base pair ClaI to NheI fragment from pIK6.1MCVampacUTΔ. pIK6.1MCVamenvATGUTΔ was constructed by replacing the 961 base pair Hind III to Eco RI fragment containing the CMV promotor and alpha globin splice acceptor in pIK6.1amenvATGUTΔ with the corresponding 896 base pair Hind III to Eco RI fragment from pIK6.1MCV.

10

pIK6.1MCVgagpolATG was constructed by replacing the 961 base pair Hind III to Eco RI fragment containing the CMV promotor and alpha globin splice acceptor in pIK6.1gagpolATG with the corresponding 896 base pair Hind III to Eco RI fragment from pIK6.1MCV.

The retroviral packaging plasmids of the invention, designated pIK6.1MMSVampac and pIK6.1MCVampac, have been deposited with the American Type Culture Collection (ATCC), 12301 Parklawn Drive, Rockville, Maryland, under the Budapest Treaty, and have there been identified as follows:

<u>Plasmid</u>	<u>ATCC Accession No.</u>	<u>Deposit Date</u>
pIK6.1MMSVampac	75484	June 11, 1993
25 pIK6.1MCVampac	75483	June 11, 1993

In another embodiment, the packaging functions may be encoded by two plasmid based expression vectors, for example two helper sequences, where a first helper sequence contains a cDNA encoding the gag and pol proteins of ecotropic MMLV and a second helper sequence contains a cDNA encoding a retroviral env protein. The Env gene, which determines the host range, may be derived from the genes encoding the xenotropic, amphotropic, ecotropic, polytropic (mink focus-forming) or 10A1 murine leukemia virus, Gibbon Ape Leukemia Virus (GALV), the Human Immunodeficiency Virus (gp160) env proteins; the Vesicular Stomatitis Virus (VSV) G

protein; the Human T cell leukemia (HTLV) type I and II env gene products; a chimeric envelope gene derived from combinations of one or more of the aforementioned env genes; or chimeric envelope genes encoding the cytoplasmic and transmembrane of the aforementioned env gene products and a monoclonal antibody directed against a specific surface molecule on a desired target cell.

Construction of plasmids which reflect this embodiment is described as follows: pIK6.1gagpolATG, encoding the MMLV gag and pol genes, was constructed first by digestion of pMOVpsi- with Sca I, addition of a Nhe I synthetic linker, redigestion with Afl II and isolation of the 5.2 kb Afl II/Nhe I fragment (nucleotides 644 to 5869 of MMLV). A synthetic oligonucleotide encoding nucleotides 621 to 644 of MMLV (ATG of the gag gene to Afl II), in which the ATG at nucleotide 621 was converted to a Nco I site, was ligated together with the Afl II/Nhe I fragment between the Nco I site polylinker and the Nhe I site at the 5' end of the SV40 poly adenylation site of pIK6.1Nhe.

pIK6.1amenvATG, encoding the MLV 4070A Env gene, was constructed by digestion of pCRIPAMGAG-2 (Danos and Mulligan, supra) with Afl 111 and redigestion with either Nhe1 or HinP1 and isolation of the .325 kb HinP 1/Afl 111 fragment (nucleotides 37 to 365 of the MLV 4070A Env gene; (Ott et.al., J. Virol. 64(2):757-766(1990)) and the 1.7 kb Afl 111/Nhe 1 fragment (from nucleotide 365 of the MLV 4070A Env gene; (Ott et.al., supra) to the Nhe 1 site in the MMLV 3' LTR of pCRIPAMGAG-2 (Danos and Mulligan, supra) respectively. A synthetic oligonucleotide encoding nucleotides 37 to 43 of the MLV 4070A Env gene (ATG of the env gene to HinP 1), in which the ATG at nucleotide 37 was converted to a Nco I site, was ligated together with the HinP 1/Afl 111 fragment and the Afl 111/Nhe 1 fragment between the Nco I site in the polylinker and the Nhe I site at the 5' end of the SV40 poly

adenylation site of pIK6.1Nhe. These plasmids are diagrammed in Figure 1A.

The two genome retroviral packaging plasmids of the invention, designated pIK6.1gagpolATG and pIK6.1amenvATG, have been deposited with the American Type Culture Collection (ATCC), 12301 Parklawn Drive, Rockville, Maryland, under the Budapest Treaty, and have there been identified as follows:

<u>10 Plasmid</u>	<u>ATCC Accession No.</u>	<u>Deposit Date</u>
pIK6.1gagpolATG	75486	June 11, 1993
pIK6.1amenvATG	75485	June 11, 1993

15 Retroviral Vectors

Both single genome and two genome packaging constructs utilize retroviral vectors that contain modified 5' LTRs that direct efficient transcription in the cell type where retrovirus is to be produced. The retroviral vectors of the invention are modeled after pZen (Johnson *et al.*, The EMBO Journal 8(2):441-448 (1989)), a neo- version of pZIPneoSVX (Cepko *et al.*, Cell 37:1053-1062(1985)), in which the gene product to be expressed is cloned downstream of the splice acceptor in the position normally occupied by the neo cassette (Cepko *et al.*, *supra*). In addition, viral gag sequences up to the Nar I site of MMLV (nucleotide 1038) were added for improved packaging (Armentano *et al.*, J. Virol. 61:11647-1650 (1987)) and the Xho I to Cla I fragment of pZIPneoSVX was deleted (Cepko *et al.*, *supra*). The Eco RI to Apa I polylinker from pIK1.1 was inserted downstream of the splice acceptor to enable transfer of inserts from pIK plasmids into retroviral constructs. The resulting plasmid is designated pRTD1.2 and contains both 5' and 3' MMLV LTRs. The 5' LTR U3 region of pZIPneoSVX was replaced with the MMSV U3, derived from the HindIII/Sac I fragment of pIKMMSV, to generate pRTD4.2. In pRTD2.2, the U3 region of the 5' LTR of

pZIPneoSVX was replaced with the Hind III/Sac I fragment from pIK1.1 encoding the CMV immediate early enhancer/promoter, which was fused to the MMLV R region by an oligonucleotide that encodes nucleotides 19 (Sac I) to +1 of the HCMV promoter linked to nucleotides +1 to +32(KpnI) of MMLV (Schinnick et al., Nature 293:543-548 (1980)). pRTD2.2SVG was constructed by replacement of the (750 base pair) Sac I to Bst EII fragment of pRTD2.2 with the (736 base pair) Sac I to Bst EII fragment of LXSX (Miller and Rosman, BioTechniques 7:980-990(1989)). pRTD2.2SSA was constructed by replacement of the (1441 base pair) Sac I to Eco RI fragment of pRTD2.2 with the (1053 base pair) Sac I to Eco RI fragment of LXSX (Miller and Rosman, supra). pRTD2.2SVGE- was constructed by synthesis of an oligonucleotide encoding nucleotides 2878-2955 of pLXSX (GenBank Accession Bank, M28248) which had been appended by addition of an Apa I site on it's 5' end. This was used to replace the Apa I to Nhe I fragment of pRTD2.2SVG, which contains the DNA sequence 3' of the of the polylinker and 5' of the Nhe I site in the 3' LTR. These retroviral vector constructs of the invention have a pBR322 backbone and include pRTD2.2, pRTD4.2, pRTD2.2SVG, pRTD2.2SVGE- and pRTD2.2SSA.

In order to permit plasmid replication in cells which express the SV40 T antigen, the sequences between the 5' and 3' LTRs of pRTD2.2 were cloned between the SacI and Eco RI sites of pIK1.1, described above, which contains the SV40 origin of replication to form vector pIKT2.2. pIKT2.2SVG was constructed by insertion of the fragment defined at its 5' end by the Sac I site in the HCMV promoter of pRTD2.2SVG and defined at its 3' end by an Eco RI site located 750 base pair downstream of the 3' LTR of pRTD2.2SVG, between the SacI and Eco RI sites of pIK1.1. pIKT2.2SVGE-F3 was constructed by replacing the 182 base pair ApaI to NheI fragment of pIKT2.2SVGF3 with the 80 base pair ApaI to NheI fragment from pRTD2.2SVGE-F3 as described above.

pRT43.2F3 was derived from pIKT2.2SVGE-F3 by replacing the Eco RI to ApaI polylinker located approximately 750 base pairs downstream from the 3' LTR with a synthetic oligonucleotide containing an AscI recognition site. In addition, the Nde I site at the 3' end of the viral gag sequences has been converted to an XhoI site by oligonucleotide insertion. pRT43.3PGKF3 was derived from pRT43.2F3 first by removal of the 3' LTR in pRT43.2F3 and insertion of a 3' LTR in which the sequences from PvuII to XbaI were deleted (MMLV, GenBank session #J02255 nucleotide numbers 7938-8115). In addition the MMLV splice acceptor region has been replaced with the human phosphoglycerate kinase gene promoter (GenBank Accession #M11958 nucleotides 2-516) which was cloned into a polylinker with a XhoI site at its 5' end and an Eco RI at its 3' end.

In one embodiment of the retroviral vectors of the invention, DNA encoding genes to be transduced into mammalian target cells using the method of the invention, for expression of chimeric receptor constructs are prepared. The construction of the chimeric receptor constructs is described below.

CD4/CD3 zeta and Anti-HIV/CD3 zeta Retroviral vectors

25

KAT retroviral vectors pRTD2.2F3, pRTD2.2SVGF3, pRTD2.2SSAF3, pRTD2.2SVGF3E-, pIKT2.2SVGF3 were constructed by Eco RI/Apa I digestion of pIKF3 (described below), isolation of the 1.9 kb fragment, followed by ligation of this fragment between the Eco RI and Apa I sites in the pIK polylinker of the vectors pRTD2.2, pRTD2.2SVG, pRTD2.2SSAF3, pRTD2.2SVGE-, pIKT2.2SVG. KAT retroviral vector pRTD2.2F15 was constructed by Eco RI/Apa I digestion of pIKF15neo (described below), isolation of the 2.2 kb fragment, followed by ligation of this fragment between the Eco RI and Apa I sites in the pIK polylinker of the vector pRTD2.2. These vectors encode a chimeric molecule containing the

extracellular domain of human CD4 (F3 derivatives) or a single chain antibody against gp41 of HIV (F15 derivatives), respectively, fused to the cytoplasmic domain of the CD4 receptor (amino acids 372-395 of the mature CD4 chain) and
5 the transmembrane domain of the CD3-complex associated-gene zeta (Δ) (amino acids 372-395 of the mature zeta chain). Chimeric receptor cassettes encoding either the extracellular domains (residues 1-371 of the mature CD4 protein) of the human CD4 receptor (designated F3) or a single chain antibody
10 to HIV gp41 derived from a human antibody (98.6) specific for the gp41 moiety of the HIV envelope protein (designated F15) were fused to the CD3 Δ chain and cloned between the Eco RI and Apa I sites of pIK1.1 described above. In the single-chain antibody, the variable domains of both the heavy and
15 light chain genes were covalently linked via a peptide tether, to create an antigen binding site on a single molecule.

A more detailed description of the construction of
20 the chimeric receptors follows.

Construction of CD4-zeta Chimeras

Plasmid pGEM3zeta bears the human zeta cDNA
25 (Weissman et al., Proc. Natl. Acad. Sci. USA 85:9709-9713 (1988)). The plasmid pBS.L3T4 bears the human CD4 cDNA (Littman and Gettner, Nature 325:453-455 (1987)). A BamHI-ApaI restriction fragment (approximately 0.64 kb) encompassing the entire human zeta chain coding sequence from
30 residue 7 of the extracellular (EXT) domain, was excised from pGEM3zeta, and subcloned into the BamHI and ApaI restriction sites of the polylinker of pBluescript II SK (+) 9pSK is a phagemid based cloning vector from Stratagene (San Diego, CA), generating pSK.zeta. Subsequently, a BamHI restriction
35 fragment encompassing the entire CD4 coding sequence (approximately 1.8 kb) was excised from pBS.L3T4, and

subcloned into the BamHI site of pSK.zeta, generating pSK.CD4.zeta.

Single-stranded DNA was prepared from pSK.CD4.zeta
5 (Stratagene pBluescript II protocol), and used as a template for oligonucleotide-mediated directional mutagenesis (Zoller and Smith, Nucleic Acids Res. 10:6487-6500 (1982)) in order to generate CD4-zeta chimeras with the desired junctions described below. CD4-zeta fusions 1, 2, and 3 were
10 subsequently sequenced via the Sanger dideoxynucleotide technique (Sanger et al., Proc. Natl. Acad. Sci. 74:5463-5467 (1977)), excised as EcoRI-ApaI restriction fragments, and cloned into the polylinker of expression vector pIK.1.1 or pIK.1.1.Neo at identical sites.

15 An EcoRI-BamHI restriction fragment (approximately 1.8 kb) encompassing the entire coding region of CD4 was excised from pSK.CD4.zeta, and subcloned between the EcoRI and BglII sites of the pIK.1.1 or pIK.1.1.Neo polylinker.

20 The plasmid pUCRNeoG (Hudziak, et al., Cell (1982) 31:137-146) carries the neomycin gene under the transcriptional control of the Rous Sarcoma virus (RSV) 3' LTR. The RSV-neo cassette was excised from PURCNeoG as a
25 HincII restriction fragment (approximately 2.3 kb), and subcloned between the two SspI sites of pIK.1.1, generating pIK.1.1.Neo.

The CD4-zeta chimeric receptor F3 was constructed
30 from the extracellular (EC) and cytoplasmic (CYT) domains of CD4 and zeta respectively. The transmembrane (TM) domain of this receptor was derived from CD4. F3 retains the CD4 EXT domain comprising all four V domains (residues 1-371 of the mature CD4 protein), the TM domain of CD4 (residues 372-395
35 of the mature CD4 chain), and the CYT domain of zeta (residues 31-142 of the mature zeta chain).

Preparation of Single Chain Antibody-Zeta Chimeric Receptor

Construction of expression vector encoding the heavy chain of human monoclonal antibody (mAb) 98.6:

5

To direct the expression of the heavy chain of human mAb 98.6 (S. Zolla-Pazner, Proc. Natl. Acad. Sci. (1989) 86:1624-1628), the plasmid pIK.98.6- γ FL was constructed. A full length IgG1 heavy chain cDNA was
10 generated by reverse transcription of 5 μ g of total RNA from the cell line SP-1/98.6 (Zolla-Pazner, supra) using oligo-dT as the primer, followed by PCR using oligonucleotide primers 17 and 2 (see below). The 1.5 kb Eco RI to Bgl II fragment was cloned between the Eco RI and Bgl II sites of pIK1.1. To
15 ensure that the heavy chain would be of the desired allotype, the Kas I-Bgl II fragment of the cDNA was replaced with a 0.94 kb Kas I - Bgl II fragment from pIK.C γ 1. pIK.C γ 1 was constructed by the insertion of a cDNA coding for the constant region of IgG1 heavy chain obtained by PCR using DNA
20 from a human spleen cDNA library (Clontech, Inc., Palo Alto, CA) as substrate and oligonucleotide primers 2 and 18 (see below), between the Eco RI and Bgl II sites of pIK1.1.

Construction of expression vector encoding the light chain of
25 human monoclonal antibody (mAb) 98.6:

To direct the expression of the light chain of mAb 98.6, the plasmid pIK.98.6 κ FL was constructed. A full length IgG1 light chain cDNA was generated by reverse transcription
30 of 5 μ g of total RNA from the cell line SP-1/98.6 using pdN $_6$ (Pharmacia/LKB) as the primer, followed by PCR with primers 19 and 20 (see below). The 0.78 fragment was then cut with Eco RI and Bgl II and cloned between the Eco RI and Bgl II sites of pIK1.1.

35

Construction of expression vector encoding SAb derived from the heavy and light chains of mAb 98.6:

a) Construction of pIK98.6-K/L/H:

To direct the expression of a single-chain antibody (SAb) form of mAb 98.6, pIK.98.6-K/L/H was constructed. The SAb expressed consists of the secretion leader sequence and amino acids 1-107 of the mature 98.6 mAb light chain variable (V_L) region fused to a 14 amino acid linker of the sequence GSTSGSGSSEGGK (SEQ ID NO. 9) (L212, Betzyk *et al.*, *J. Biol. Chem.* (1990) 265:18615-18620), which in turn was fused to amino acid 1 of the mature 98.6 mAb heavy chain V_H region. This was then fused at amino acid 113 to amino acid 234 of the IgG1 heavy chain constant region, in order to delete the CH1 domain of the IgG1 heavy chain constant region for improved secretion. pIK.98.6-K/L/H was constructed in three steps.

First, deletion mutagenesis was performed to fuse amino acid 113 of the V_H region of mAb 98.6 to amino acid 234 of the IgG1 heavy chain, using the single stranded template form of pIK.98.6- γ FL as the template and oligonucleotide 21 as primer (see below). Correctly deleted plasmids were found using oligonucleotide 22 as a probe (see below). This plasmid is referred to as pIK.H/Fc-int. To fuse amino acid 107 to the amino terminus of the linker peptide, the V_L region of the mAb 98.6 light chain was generated by PCR using pIK.98.6- κ FL as substrate and oligonucleotides 23 and 24 as primers (see below). This was done to place a Sal I site at the 3' end of the V_L sequence, without altering the amino acid sequence of the resulting protein. This fragment, together with oligonucleotides 25 and 26 (see below) was ligated between the EcoRI and Bgl II sites of pIK1.1, generating the plasmid pIK.K/L-int.

In the final step, the 0.45 kb fragment of pIK.K/L-int was cloned between the Eco RI and Kpn I sites of pIK.H/Fc-int., generating plasmid pIK.K/L/H-int. Single-stranded DNA from this plasmid was used as template and

oligonucleotide 27 was used as primer (see below) to fuse the carboxy-terminal amino acid of the linker to amino acid 1 of the V_H region of mAb 98.6 by deletion mutagenesis. Correctly deleted plasmids were found using oligonucleotide 28 as a 5 probe (see below). The resulting plasmid is pIK.98.6K/L/H.

b) Construction of pIK.CD4 γ 2:

The plasmid pIK.CD4 γ 2 was constructed to direct the
10 expression of a fusion protein composed of the secretion leader and the first 180 amino acids of the mature CD4 antigen fused to amino acid 234 of the human IgG2 heavy chain constant region and thus containing part of the hinge and all of the CH2 and CH3 domains. This deletes the CH1 domain of
15 the IgG2 heavy chain for improved secretion. pIK.CD4 γ 2 was constructed by generating a fragment containing the Fc portion of the human IgG2 heavy chain by PCR using DNA from a human spleen cDNA library (Clontech) as substrate and oligonucleotides 3 and 4 as the primers. The 0.75 kb Nhe I
20 to Bgl II fragment generated was ligated together with the 0.6 kb Eco RI to Nhe I fragment from pSKCD4 Δ between the Eco RI and Bgl II sites of pIK1.1.

c) Construction of pIK.F5:

25

The plasmids pIK.F7 was constructed to direct expression of several versions of CD4/IgG/zeta (Δ) fusion proteins which all contain a human membrane-bound IgG membrane hinge domain (Tyler et al. (1982) Proc. Natl. Acad. Sci. 79:2008-2012). Each protein to be expressed contained
30 amino acids 1-180 of CD4 receptor, followed by amino acids 234-445 of human IgG2 heavy chain constant region, followed by the 18 amino acid M1 membrane hinge domain of human IgG3 (Bensmana and Lefranc, (1990) Immunogenetics 32:321-330),
35 followed by a transmembrane domain, followed by amino acids 31-142 of the human Δ chain. pIK.F7 contains the transmembrane domain (amino acids 372-395) of CD4.

To construct this plasmid, the first step was cloning the human IgG3 M1 exon (Bensmana and Lefranc, supra). This was done by generating a 0.13 kb Bam HI to Bgl II fragment containing the M1 exon by PCR using DNA from the 5 human cell line W138 as substrate and oligonucleotides 7 and 8, and cloning it into the Bgl II site of pIK.CD4 γ 2. The resulting plasmid is referred to as pIK.CH3/M1-int. Single stranded DNA from this plasmid was used as template and oligonucleotide 9 was used as the primer to fuse amino acid 10 445 of human IgG2 to the first amino acid of the IgG3 membrane hinge domain by deletion mutagenesis. The fusion is designed to generate the sequence found at the natural junction between CH3 and M1 in membrane-bound IgG molecules. Correctly deleted clones were found using oligonucleotide 10 15 as a probe. The resulting plasmid is referred to as pIK.CD4 γ 2/M1.

pIK.CD4 γ 2/M1 was cut with Bgl II and blunted with T4 polymerase, then cut with Nhe I. The resulting 0.83 kb 20 fragment was ligated together with the 1.3 kb Pvu II to Apa I fragment from pIK.F3 between the Nhe I and Apa I sites of pIK.CD4 γ 2 to generate the plasmid pIK.F7-int. Single stranded DNA from this plasmid was used as template and oligonucleotide 15 was used as the primer to fuse the last 25 amino acid of the IgG3 M1 membrane hinge domain to amino acid 372 of CD4 by deletion mutagenesis. Correctly deleted clones were found by using oligonucleotide 16 as a probe. The resulting plasmid is pIK.F7.

30 The oligonucleotides used as primers and probes as described above were as follows:

Oligonucleotides

- 35 2. CGGAGATCTCGTGCGACCGCGAGAGCC (SEQ ID NO. 10)
3. GGAATTCGCTAGCTTTCCAGGAGCGCAAATGTTGTGTC (SEQ ID NO. 11)
4. CGGAGATCTC (A/G) CGCGACCCCGAGAGCC (SEQ ID NO. 12)

7. CGGGATCCAGAGCTGCAACTGGAG (SEQ ID NO. 13)
8. GAAGATCTGACCTTGAAGAAGGTGAC (SEQ ID NO. 14)
9. TCTCCTCCAGTTGCAGCTCCGGAGACAGGGAGAGGC (SEQ ID NO. 15)
10. TTGCAGCTCCGGAGAC (SEQ ID NO. 16)
- 5 15. CAGCACAATCAGGGCCATGTCCAGCTCCCCGTCCTG (SEQ ID NO. 17)
16. AGGGCCATGTCCAGCT (SEQ ID NO. 18)
17. CGGAATTCGGTACCTCCTGTGCAAGAAC (SEQ ID NO. 19)
18. CGGAATTCGCCTCCACCAAGGGCCCA (SEQ ID NO. 20)
19. CGGAATTCACGCGTCCCAGTCAGGACACAGC (SEQ ID NO. 21)
- 10 20. GAGAGAGATCTGCTAGCGGTGAGGCTGGAACTGAG (SEQ ID NO. 22)
21. GCATGTGTGAGTTTGTCTGAGGAGACGGTGACCAG (SEQ ID NO. 23)
22. GTTTTGTCTGAGGAGA (SEQ ID NO. 24)
23. GTGACAGTCGACCCCTTGAAGTCCACTTTGGT (SEQ ID NO. 25)
24. CCACCCCTCACTCTGCTTCTC (SEQ ID NO. 26)
- 15 25. TCGACCAGCGGCAGCGGCAAGAGCAGCGAGGGTAAGGGTACCA (SEQ ID NO. 27)
26. GATCTGGTACCCTTACCCTCGCTGCTCTTGCCGCTGCCGCTGG (SEQ ID NO. 28)
27. CTCCTGTAGTAGCACCTGACCCTTACCCTCGCTGCT (SEQ ID NO. 29)
- 20 28. AGCACCTGACCCTTAC (SEQ ID NO. 30)

Construction of pIK.F15neo:

To direct the expression of a fusion protein consisting of the K/L/H SAb form of mAb 98.6 linked at amino acid 445 of the IgG1 heavy chain to the 18 amino acid IgG3 M1 membrane hinge, which was in turn fused to the CD4 transmembrane domain (amino acids 372-395) and Δ cytoplasmic domain (amino acids 31-142), pIK.F15neo was constructed by inserting the 1.5 kb Nsi I fragment of pIK.98.6-K/L/H between the Nsi I sites of pIK.F7neo and a clone of the correct orientation was selected.

Production of Retrovirus in mammalian cells

35

Single or double genome KAT packaging plasmids, for example pIK6.1MMSVampac, pIK6.1MCVampac, or pIK6.1amenvATG and

pIK6.1gagpolATG (all described above), together with KAT retroviral constructs, for example, but not limited to pRTD2.2F3, pRTD2.2SVGF3, pRTD2.2SSAF3, pRTD2.2SVGF3E-, pIKT2.2SVGF3, pRTD2.2F15 (as described above), prepared as 5 described above, are introduced into mammalian cells that can produce virus by standard means such as calcium phosphate cotransfection (Wigler et al., Proc. Natl. Acad. Sci. USA 76:1373-1377 (1979)). Mammalian cells that can produce virus and that may be transfected by the KAT constructs of the 10 invention include, but are not limited to, human embryonic kidney cells such as 293 cells, tsA201 cells, mouse 3T3 mouse fibroblasts, M. dunni fibroblasts, and African green monkey kidney (COS) cells. Transfected cells are assayed for surface expression of the chimeric receptor by FACS to verify that 15 DNA constructs have been successfully introduced.

Viral supernatants are harvested using standard techniques such as filtration of supernatants 48 hours post 20 transfection. The viral titer is determined by infection of 10^6 NIH 3T3 cells with an appropriate amount of viral supernatant, in the presence of 8 μ g/ml polybrene (Sigma Chemical Co., St. Louis, MO). 48 hours later, the transduction efficiency of the 3T3 cells is assayed by both 25 FACS analysis and Southern blotting.

High Efficiency Transduction of target cells

In the method of the invention the KAT constructs 30 of the invention are further used to transduce mammalian target cells with a foreign gene at high efficiency by cocultivation of KAT transfected cells with the mammalian target cells. In a preferred embodiment, desired virus producing cells, such as 293 cells, are transfected with the 35 appropriate KAT constructs, then 24 hours post transfection, the transfected 293 cells are cocultivated for 48 hours with the purified mammalian target cells, such as CD8+ T cells.

Alternatively, fresh media is added 24 hours post-transfection. Forty-eight hours post-transfection, virus supernatants are harvested, filtered through a 0.45 μ filter and used to infect target cells. The target cells are
5 harvested using standard procedures, expanded and tested for transduction efficiency, by well-known techniques such as flow cytometry or Fluorescence-activated Cell Sorter (FACS) analysis and Southern blot DNA analyses. Transduction efficiency is defined as the percentage of positive
10 transduced cells as measured by FACS or Southern blot analysis compared to controls.

Using the KAT constructs transfected into human 293 cells to produce virus, a from 5 to 50-fold increase in viral
15 titer as determined by supernatant infection of established cell lines, such as 3T3, is obtained, when compared to virus produced by the previously described COS transient virus production system (Landau and Litman, supra). In addition, primary human cells such as hematopoietic stem cells and
20 human T cells, are transduced at levels 3 to 20 fold greater by cocultivation with KAT plasmid transfected 293 cells, than traditional packaging lines such as PA317 (Miller and Buttimore, supra).

25 While not wishing to be bound by any particular theory of the invention, it is believed that the high efficiency transduction of human target cells obtained using the cocultivation transduction method of the invention is mediated by cell-cell contact of the retrovirally infected
30 human 293 cells with the target cells. The component of human 293 cells which effects high efficiency transduction of various target cells is expected to be a protein or lipid synthesized by the 293 cells. To determine the active component of this system, the membrane proteins and lipids of
35 293 cells are purified using known procedures and the ability of various purified components is tested for its ability to effect the transduction efficiency of the target cells. Once

the active component is identified it can be synthesized by recombinant DNA or chemical technique. These synthesized components may be incorporated into virus particles to enhance the transduction efficiency of supernatants.

5

Suitable target cells are any mammalian cells of interest, and include, but are not limited to lymphocytes, particularly cytotoxic T cells, human hematopoietic stem cells, fibroblasts, epithelial cells, endothelial cells, myoblasts, retinal epithelial cells, islets of Langerhans, adrenal medulla cells, osteoblasts, osteoclasts, neurons, glial cells, ganglion cells, embryonic stem cells, and hepatocytes.

15 The genes which may be introduced into the target cells include, but are not limited to genes encoding chimeric receptors for signal transduction in lymphocytes, such as those described in copending U.S. patent application Serial No. 988,194, filed December 9, 1992, the disclosure of which is incorporated in its entirety herein by reference; growth factors, such as

G-, M- and GM-colony stimulating factor (CSF), epidermal growth factor, platelet derived growth factor, transforming growth factor (TGF) and stem cell growth factor (SCF); lymphokines such as the interleukins; hormones such as ACTH, somatomedin, insulin, angiotensin; and coagulation factors, such as Factor VIII and Factor IX; the Multidrug Resistance Drug (MDR) gene; human adenosine deaminase (ADA); glucose cerebrosidase; the normal β -globin gene and erythropoietin (EPO).

The following examples are presented to illustrate the present invention and to assist one of ordinary skill in making and using the same. The examples are not intended in any way to otherwise limit the scope of the disclosure or the protection granted by Letters Patent hereon.

EXAMPLE ITransient Production of High Titer Recombinant RetrovirusCell growth, transfection and infection of5 established cell lines

Human embryonic kidney cells, designated 293 cells (ATCC, Rockville, MD) cells were grown in DMEM (JHR Biosciences, Lenexa, Kansas), 1 g/l glucose, 10% Donor calf serum (Tissue Culture Biologics, Tulare, CA) and split 1:10 every 3 days. 3T3 (ATCC, Rockville, MD) cells were grown in DMEM (JHR Biosciences), 4.5 g/l glucose, 10% Donor calf serum (Tissue Culture Biologics) and split 1:10 every 3 days. COS (ATCC, Rockville, MD) cells were grown in DME/F12 (GIBCO, Grand Island, NY), 10% fetal bovine serum (Tissue Culture Biologics, Tulare, CA) and split 1:10 every 3 days. tsa201 cells, a derivative of 293s which contain the temperature sensitive mutant of the SV40 T antigen co-transfected with the neomycin resistance gene (Heinzel *et al.*, *J. Virol.* 62(10):3738-3746 (1988)), were grown in DME/F12 (GIBCO), 10% fetal bovine serum (Tissue Culture Biologics) and split 1:10 every 3 days. 293 cells and tsa201 cells were plated 1×10^6 and 0.5×10^6 cells per 10 cm plate, respectively, 48 hours prior to transfection. COS and 3T3 cells were plated at 0.5×10^6 cells per 10 cm plate 24 hours prior to transfection. 10 μ g of each plasmid, alone or in various combinations, was transfected by calcium phosphate coprecipitation (Wigler *et al.*, *supra*) for all cell types. 24 hours following transfection, the media was changed. 24 hours later, viral supernatants were harvested and filtered through a 0.45 μ m filter and flash frozen on dry ice. 3T3 cells were plated at 0.5×10^6 cells per 10 cm plate 24 hours prior to infection. Infections were carried out in 5 ml of media containing viral supernatant and 8 μ g/ml polybrene (Sigma Chemical Co., St. Louis, MO). 24 hours following infection, the media was changed to polybrene-free media and the cells were grown for an additional 24 hours.

293 cells produced high titer retrovirus following transient transfection

293 cells were assayed for their ability to
5 transiently produce recombinant retrovirus upon
cotransfection with the either the KAT packaging plasmid(s)
pIK6.1MCVampac or pIK6.1amenvATG and pIK6.1gagpolATG, and the
retroviral vectors pRTD2.2F3, pRTD2.2SVGF3, pRTD2.2SSAF3,
pRTD2.2SVGF3E-, pIKT2.2SVGF3, and pRTD2.2F15, encoding the F3
10 or F15 chimeric receptors, by harvesting viral supernatants
48 hours post transfection, followed by infection of mouse
3T3 cells, and FACS analysis 48 hours later.

For FACS analysis, infected 3T3 cells are removed
15 from the culture dish in the absence of trypsin and are
processed for FACS analysis after incubation in 40 mM Tris,
pH 7.5, 150 mM NaCl, 1 mM EDTA. Cells are washed 1X with
phosphate buffered saline (PBS) plus 2% (FCS) fetal calf
serum (Hyclone), followed by incubation with the appropriate
20 FITC-conjugated detection antibody in the presence of PBS
plus 2% FCS at a density of 1×10^6 /ml for 30 minutes at 40°C.
The cells are washed 3X with PBS plus 2% FCS, and finally
resuspended in 0.5 ml PBS and analyzed by flow cytometry.

25 The results of FACS analysis are shown in Figure 2.
293 cells cotransfected pIK6.1ampac and pRTD2.2F3 express
high levels of F3 on their surface (Figure 2B), compared to
mock (control) transfected cells (Figure 2A). 3T3 cells
infected with viral supernatants harvested from transfected
30 293 cells revealed two well separated peaks corresponding to
uninfected and infected 3T3 cells (Figure 2D), which was
significantly different compared to the FACS profile of
transfected 293 cells (Figure 2B) or mock infected 3T3 cells
(Figure 2C).

35

Table 1 demonstrates that cotransfection of KAT
packaging plasmids and KAT retroviral constructs results in

the production of high titer viral supernatants 48 hours following transfection, as assayed by 3T3 infection and FACS analysis. Cotransfection of pIK6.1ampac and pRTD2.2F3 yields viral supernatants that transduce 50% of the 10^6 3T3 cells initially present at the time of infection. In contrast, virus produced by transient cotransfection in COS cells, as described by Landau and Litman (Landau and Litman, supra) was 10-fold less than the titers described by cotransfection of KAT plasmids into 293 cells. Virus production is highly reproducible in four transfection experiments, where duplicate 3T3 infections were carried out. In contrast, no detectable 3T3 infection is observed following transfection of the retroviral construct pRTD2.2F3 alone, demonstrating that viral production is dependant upon the presence of the packaging construct and the retroviral vector. High titer virus production is also dependant upon the presence of the retroviral construct. Transfection of pIKF3 expression vector alone, or cotransfection of pIKF3 expression vector and pIK6.1MMSVampac yields supernatants that fail to transduce 3T3 cells.

Table 1

Construct	Packaging Function	% Transfection	% 3T3 Transduction
pRTD2.2F3	--	52	0/0
pRTD2.2F3	--	55	0/0
pRTD2.2F3	pIK6.1MCVampac	80	49/50
pRTD2.2F3	pIK6.1MCVampac	85	50/49
pRTD2.2F3	pIK6.1MCVampac	83	47/43
pRTD2.2F3	pIK6.1MCVampac	85	49/48
pRTD2.2F3	pIK6.1gagpolATG, pIK6.1amenvATG	78	27/77
pRTD2.2F3	pIK6.1gagpolATG,pIK6. 1amenvATG	78	25/26
pIKF3	-	67	0/0

	pIKF3	-	59	0/0
	pIKF3	pIK6.1MCVampac	90	0/0
	pIKF3	pIK6.1MCVampac	90	0/0
5	pRTD2.2ssaF3	pIK6.1MCVampac	78	33/35
	pRTD2.2svgF3	pIK6.1MCVampac	84	44/39
	pRTD2.2svge-F3	pIK6.1MCVampac	81	42/43
	pRTD2.2F15	pIK6.1MCVampac	93	70/70
10	pRTD2.2F15	pIK6.1MCVampac	91	69/70

High titer virus can also be produced by cotransfection of pIK6.1amenvATG, pIK6.1gagpolATG and pRTD2.2F3 (Table 1). Although the transfection efficiency of the later plasmids was approximately equal to the transfection efficiency of pIK6.1MCVampac and pRTD2.2F3, virus production was reduced by a factor of 2 to 27%. Similar results have been described by Landau and Litman (Landau and Litman, supra), where they observed a 5-fold decrease. The overall efficiency of the KAT system, using one or two genome packaging plasmids, is still 10 to 20-fold greater than that described for the COS cell system.

The high 3T3 cell transduction efficiency observed by FACS analysis of viral supernatants produced following KAT plasmid transfection of 293 cells was confirmed by Southern blotting of integrated proviral DNA from infected 3T3 cells. High molecular weight DNA was prepared 48 hours post infection and digestion of 10 μ g of DNA with Eco RV. The samples were electrophoresed on a 0.8% agarose gel, transferred to Zetabind and probed with a 605 base pair fragment encoding the zeta transmembrane and cytoplasmic domains. Eco RV digestion of the transfected plasmid pRTD2.2F3 yielded a 4.2 kb band. Eco RV digestion of pRTD1.2F3, which contains MMLV 5' and 3' LTRs, yielded a 3.6 kb fragment. Following virus infection, integration and duplication of the 3' LTR, Eco RV digestion should yield a

3.6 kb fragment. This allows determination of the presence of integrated proviral DNA in the target cells. Table 2 gives the sizes of the expected bands from transfected plasmid DNA and integrated provirus following Eco RV digestion and hybridization to the zeta probe.

Table 2

10	EcoRV Fragment Size (in Kb) Hybridizing to Δ Probe		
	Retroviral Construct	Transfected Plasmid	Integrated Provirus
	pRT.D 2.2F3	4.20	3.60
	pRT.D 2.2SSAF3	3.80	3.20
15	pRT.D 2.2SVGF3	4.17	3.57
	pRT.D 2.2SVGE-F3	4.22	3.61
	pRT.D 2.2F15	4.47	3.87

20 Genomic DNA prepared from infected 3T3s was digested with Eco RV and 10 μg of digested DNA from infected and control cells were electrophoresed on a 0.8% agarose gel, transferred to Zetabind and probed with a 605 base pair fragment encoding the Δ transmembrane and cytoplasmic domains. Only the DNA derived from 3T3 cells infected with 25 supernatants obtained following cotransfection of 293 cells with pRTD2.2F3 and pIKMCVampac yielded a 3.6 kb fragment (Figure 3A, lanes 4 and 5), identical to the fragment seen in the Eco RV digested pRTD1.2F3 plasmid control lanes (Figure 30 3A, lanes 11-14), indicative of integrated provirus. Quantitation of southern blots by scanning densitometry and comparison to plasmid standards representing 0.1 to 3.0 copies, in 3-fold increasing increments (Figure 3A, lanes 11-14), was consistent with a transduction efficiency of with a 35 transduction efficiency of 0.5 copies/cell/ml of viral supernatant. The transduction efficiency was identical to the efficiency observed by FACS analysis. The probe did not

detect a band in DNA from 3T3 cells infected with supernatants derived from mock transfected 293 cells (lane 1), 293 cells transfected with pRTD2.2F3 alone (Figure 3A, lanes 2 and 3), transfected with the expression vector pIKF3 5 alone (Figure 3A, lanes 6 and 7) or cotransfected with pIK6.1MCVampac and pIKF3 (Figure 3A, lanes 8 and 9), which is also consistent with the FACS analysis.

Three additional retroviral constructs, two which 10 differed in the viral backbone, pRTD2.2SSAF3 (Figure 3B, lane 4), pRTD2.2SVGF3 (Figure 3B, lane 5), pRTD2.2SVGE-F3 (Figure 3B, lane 6), and one which differed in the chimeric receptor insert, pRTD2.2F15 (Figure 3B, lanes 7 and 8), were cotransfected into 293 cells with pIK6.1MCVampac, the 15 supernatant used to infect 3T3 cells, followed by both FACS analysis (Table 1) and southern blotting (Figure 3B). All of the F3 constructs showed similar titer by both FACS analysis (Table 1) and hybridized to the zeta probe with similar intensities, as expected. The F15 retrovirus had 20 approximately 50% greater titer as determined by FACS analysis (Table 1), as well as by densitometric analysis of the Southern blots. Retrovirus as produced in 293 with each of the vectors, upon infection, yielded the correct size for the integrated provirus. Therefore, the FACS and Southern 25 blotting results from 5 KAT retroviral constructs demonstrate that high titer retrovirus can be produced in 293 cells, that production was dependent upon cotransfection of the retroviral construct and packaging functions, and production of high titer retroviral supernatants in 293 cells does not 30 lead to any unusual rearrangements of the retroviral constructs.

Virus Production in Mammalian Cell Lines:

35 Seven additional cell lines were screened for their ability to produce retrovirus by cotransfection

with KAT plasmids, followed by virus harvest and 3T3 infection (Table 3).

Table 3

5	Cell Type	Packaging Construct	Surface CD4%	3T3 inf%	Retro-F3 Constr.
	293	Mock	1	0/0	Mock
	293	pIK6.1MCVampac	88	39/38	pRTD2.2-F3
	293	pIK6.1MCVampac	88	41/38	pRTD2.2-F3
10	COS	Mock	0	ND	Mock
	COS	pIK6.1MCVampac	58	12/14.	pRTD2.2-F3
	COS	pIK6.1MCVampac	58	14/15	pRTD2.2-F3
	143B	Mock	0	ND	Mock
	143B	pIK6.1MCVampac	54	1/1.	pRTD2.2-F3
15	143B	pIK6.1MCVampac	50	1/1.	pRTD2.2-F3
	HELA	Mock	0	ND	Mock
	HELA	pIK6.1MCVampac	48	0/0	pRTD2.2-F3
	HELA	pIK6.1MCVampac	54	0/0	pRTD2.2-F3
20	L929	Mock	0	ND	Mock
	L929	pIK6.1MCVampac	1	0/0	pRTD2.2-F3
	L929	pIK6.1MCVampac	1	0/0	pRTD2.2-F3
	3T3	Mock	0	0/0	Mock
	3T3	pIK6.1MCVampac	39	2/3.	pRTD2.2-F3
25	3T3	pIK6.1MCVampac	44	4/3.	pRTD2.2-F3
	CHO D-	pIK6.1MCVampac	0	0/0	pRTD2.2-F3
	CHO D-	pIK6.1MCVampac	0	0/0	pRTD2.2-F3

30 CD4 surface expression and virus production was absent from L929 and CHO D- following cotransfection of pIK6.1MCVampac with pRTD2.2F3. However, these cell lines were highly transfectable under conditions with a plasmid encoding the lac z gene was used. FACS analysis of
 35 transfected HELA, 143B, 3T3 and COS demonstrated high surface CD4 expression, with a transfection efficiency of approximately 50% for all four cell types. However, virus

production among these cells was substantially different. HELA and 143B cells produced no virus at all, whereas 3T3 cells produced virus capable of 3% 3T3 transduction/ml of frozen supernatant. Cotransfection of COS cells with KAT
5 plasmids, even in the absence of DNA replication of the retroviral construct, produced virus with titers of 4.5-fold greater than that produced by 3T3 cells. These titers, without plasmid replication of the viral vector construct, are 200 fold greater than those described by Landau and
10 Litman (Landau and Litman, supra). This demonstrates that the KAT constructs are unique in their ability to produce retrovirus upon transfection of a wide variety of cells, without plasmid replication. Given the 100 fold increase that Landau and Litman observed with plasmid replication of the
15 viral vector construct, transfection of KAT packaging function and retroviral vector plasmids that support plasmid replication, into hosts that support plasmid replication, could potentially further increase titer 10 to 100 fold and further increase the utility of KAT transfected cells to
20 infect cell types that are currently difficult to infect.

EXAMPLE II

High Efficiency Transduction of Human T Cells

25 This example demonstrates the method of the invention in which 293 cells transfected with the KAT constructs are able to transduce primary, human target CD8+ T cells by cocultivation with high efficiency.

30 Construction of retroviral vectors and packaging plasmids

KAT constructs were prepared as described above in Example I.

35 Isolation and activation of human CD8+ T cells from peripheral blood

Primary human CD8+ T cells were purified from the peripheral blood of healthy donors as follows: Peripheral blood mononucleocytes (PBMCs) were isolated from human blood by Ficoll-Hypaque density gradient centrifugation. PBMCs were washed three times with D-PBSE/CMF (PBS containing 1mM EDTA, Ca and Mg free), resuspended at 5×10^7 cells in 4 ml of D-PBSE/CMF containing 0.5% of human gamma globulins, and incubated at room temperature for at least 15 minutes. After incubation, CD8+ T cells were purified from the PBMC cell suspension by positive panning. Specifically, the PBMC suspension was loaded into a pre-washed T-25 tissue culture flask coated with an antibody specific for the human CD8 receptor (AIS CD8 selection flask (Applied Immune Sciences, Santa Clara, CA)) at a density of 5×10^7 cells per 4 ml per T-25 flask. Cells were incubated for one hour at room temperature, and the non-adherent cells removed by gentle pipetting and washing the flask three times with the D-PBSE/CMF. The CD8+ T cells were simultaneously released from the flask and activated by adding 10 ml of T cell medium (see below for composition) containing 10 ng/ml OKT3 (Ortho Pharmaceuticals, Raritan, NJ) and 10% IL2 (Pharmacia). Cells were incubated with this media for 48 hours, harvested from the flask, and washed once with T cell medium, and finally resuspended in fresh T cell medium plus 10% IL2 at a density of $0.5-1.0 \times 10^6$ /ml in 24 well plates.

In order to remove residual cells (usually present at 2-3%) which cross-reacted with either the CD4-specific antibody used for detection of F3 surface expression, or the human Fc-specific antibody used to detect F15 surface expression, the enriched CD8+ T cell population was subjected to a further round of purification in which the contaminating cells were removed by negative panning, using AIS selection flasks described above, coated with either the anti-CD4 or anti-human Fc antibody. Specifically, the enriched CD8+ T cell population was incubated in the selection flask for one hour, and then non-adherent (i.e., highly purified CD8+ T

cells) were removed. Cells were subsequently washed, and allowed to recover for 24 hours in the T cell medium plus 10% IL2 for 24 hours. CD8+ T cells prepared in this manner were greater than 95% CD8+ and CD3+, and less than 0.5% CD4+ or 5 FC+, and were subsequently employed as targets for retroviral transduction.

Retroviral transduction of CD8+ T cells by cocultivation or supernatant infection:

10

293 cells were plated at 1×10^6 cells/6 well plate, and then transfected with the appropriate construct after 48 hours as described above. 24 hours post transfection, the transfection media was removed and replaced with T cell 15 growth media (see below for composition).

(a) Cocultivation: 2 to 4 hours later, 0.5×10^6 purified and activated human CD8+ T cells prepared as described above (usually at day 4 or 5 post-
20 purification/activation) were added per well containing the transfected 293 cells, and polybrene added at a final concentration of 2 $\mu\text{g/ml}$. 24 hours after plating the 293 cells for the initial transfection, a second set of 293 cells were plated and transfected as described above. 24 hours
25 after the initial cocultivation, T cells were removed from the first cocultivation and transferred to the second 293 transfection plate for an additional 24 hours of cocultivation employed the same conditions. Similar conditions were employed for transduction of CD8+ T cells by
30 cocultivation with either transiently transfected 3T3 cells, or the stable PA317 producer cell line 40.39 (see below).

(b) Supernatant infection: 0.5×10^6 purified and activated human CD8+ T cells prepared as described above
35 (usually at day 4 or 5 post-purification/activation) were incubated with 1 ml of fresh T cell medium (plus 10% IL2 and 2 $\mu\text{g/ml}$ polybrene) together with 1 ml of viral supernatant

obtained from the 293 transient transfection system described above, or from the stable PA317 producer cell line 40.39 (see below). After an 8 hour incubation period, 1.5 ml of medium was removed from each well, and replaced with 0.5 ml of fresh T cell medium together with 1.0 ml of viral supernatant (polybrene at 2 μ g/ml and IL2 at 10%). After a 12 hour incubation period, the two step supernatant procedure was repeated.

10 For both cocultivation and supernatant infection, CD8+ T cells were allowed to recover for a 24-28 hour period in fresh T cell medium plus 10% IL2. Cells were then analyzed by flow cytometry for surface expression of either CD4 (for the CD4- Δ F3 receptor) or Fc for the F15 antibody- Δ 15 receptor) in order to determine transduction efficiencies. T cells which were under cocultivation with transfected 293 cells were gently removed as a suspension from the 293 monolayer. Both cocultivated and supernatant infected T cells were washed 1x with phosphate buffered saline (PBS) 20 plus 2% (FCS) fetal calf serum (Hyclone). T cells were then incubated with the appropriate FITC-conjugated detection antibody in the presence of PBS plus 2% FCS at a density of 1×10^6 /ml for 30 minutes at 40OC, washed 3x with PBS plus 2% FCS, and finally resuspended in 0.5 ml PBS and analyzed by 25 flow cytometry.

The transduced CD8+ T cell population was subsequently maintained in T cell medium (10% FCS, Hyclone; RPMI1640, CellGro; 10mM Hepes buffer (Gibco); 1% Sodium 30 pyruvate (Gibco); 1% non-essential amino acids (Gibco); 2mM glutamine (Gibco); 25 μ M 2-mercaptoethanol (Sigma) and 1% streptomycin/penicillin). T cells were periodically re-stimulated every 7 to 10 days by the addition of OKT3 at 10ng/ml or by exposing the cells to immobilized OKT3 in a T- 35 25 tissue culture flask at a density of $1-2 \times 10^7$ CD8+ T cells/10 ml T cell medium plus 10% IL2. Cells were incubated

for 48 hours, washed 1x with T cell medium, and resuspended in fresh medium plus 10% IL2 at $0.5-1.0 \times 10^6$ /ml.

Analysis of CD8+ T cell Transduction:

5

Transduction efficiency of primary human CD8+ T cells by retrovirus produced transiently using the KAT system was compared to retrovirus produced from a high-titer, stable producer clone derived from the amphotropic packaging line PA317 (Miller and Buttimore, supra). The stable producer clone 40.39, which transduces the F3 chimeric receptor was isolated by transfection of the ecotropic packaging line gpe (Markowitz et al. supra) with pRTD4.2F3, followed by supernatant harvest 48 hours post transfection and infection PA317 in the presence of 8 micrograms/ml of polybrene (Miller and Buttimore, supra). Individual clones were obtained by limiting dilution and 50 were screened for virus production by isolation of viral mRNA from the media of clones, followed by dot blot hybridization using a 603 base pair zeta chain probe. The clone that gave the strongest hybridization signal, clone 40.39, was assayed by limiting dilution infection of 10^6 NIH 3T3 cells followed by flow cytometry. 50 μ l of supernatant transduced 17% cells, equivalent to 340% or an average of 3.4 proviral copies/cell/ml. The transduction efficiency following a 48 hour cocultivation with primary human CD8+ T cells with 40.39 producer cells was 1%-3% CD4+ (Table 4).

This result was compared to the transduction efficiency following the KAT transient-transfection and cocultivation method of the invention, which was used to transduce the chimeric receptor F3 and F15 into CD8+ T cells (Figure 4). Four experiments were carried out in which CD8+ T cells were cocultivated on transfected 293 cells for 48 hours, followed by harvest and growth of T cells for 14 days and analysis of transduction efficiency by FACS as described above. The transduction efficiency of CD8 cells with both F3

and F15 constructs varies between 8% and 38%, and appears to be highly donor dependent. However, on average, this efficiency is 8 to 12-fold greater than the transduction efficiency obtained by cocultivation with the high-titer 5 stable PA317 clones tested. In addition, the high transduction efficiency is not specific to F3 constructs because F15 constructs are transduced at similar efficiencies (Figure 4). This data demonstrates that CD8 T cells can be transduced at efficiencies that are at least 5 fold greater 10 than or equal to any other published reports, and that generation of stable producers are not required.

Supernatants from transduced T cells, 3 weeks post-transduction, were tested in an extended S+L- assay (Miller 15 et al., Mol. Cell. Biol. 5:431-437 (1985)) and shown to be free of replication-competent retrovirus.

High efficiency transduction is mediated by cell-cell contact

20 In order to explore the mechanism of the high efficiency CD8 T cell transduction following transient transfection of KAT plasmids and cocultivation with CD8+ T cells, the transduction efficiency of CD8+ T cells using the following approaches was compared: (1)infection with 25 supernatants derived from a high titer, stable PA317 producer line, (2)cocultivation with a high titer, stable PA317 producer line(3)infection with supernatants derived from transient transfection of NIH 3T3 cells with pIK6.1MMSVampac and pRTD4.2F3 (4)48 hour cocultivation with NIH 3T3 cells 30 following transient transfection with pIK6.1MMSVampac and pRTD4.2F3 (5)infection with supernatants derived from transient transfection of 293 cells with pIK6.1MCVampac and pRTD2.2F3 and (6) 48 hour cocultivation with 293 cells following transient transfection with pIK6.1MCVampac and 35 pRTD2.2F3 (Table 4). For each transient transfection experiment, duplicate plates of transfected cells were used to harvest media for supernatant infection of 3T3 cells and

duplicate plates were used for cocultivation of CD8 T cells. The same approach was used for stable producers.

Table 4

5	Expt.#	Pkg. Line	Virus Production Method	Infection Method	3T3 titer Supernatant	% T-cell Transduction
	1A	PA317	PA317, Stable	Supernatant	70%	1
	1B	PA317	PA317, Stable	co-cultivation	ND ¹	3
	1C	3T3	KAT, Transient	co-cultivation	26%	3
10	1D	293	KAT, Transient	co-cultivation	14%	10
	2A	PA317	PA317, Stable	Supernatant	30%	1
	2B	PA317	PA317, Stable	co-cultivation	30%	1
	2C	293	KAT, Transient	Supernatant	45%	1
15	2D	293	KAT, Transient	co-cultivation	45%	14

ND¹ = not determined

Supernatant infection of CD8+ T cells was 1%, whether the virus was produced in 293 cells, 3T3 cells or a stable PA317 producer (Table 4, experiments 1A, 2A and 2C). In contrast, cocultivation of CD8 T cells with 293 cells cotransfected with pIK6.1MCVampac and pRTD2.2F3, resulted in 10% to 14% CD8 T cell transduction (Table 4, experiment 1D, 2D), 10 to 14-fold greater than all supernatant infections, including supernatants produced by cotransfection of these plasmids into 293 cells. This demonstrates that cell-cell contact is responsible for high efficiency transduction of CD8+ T cells. In addition, the efficiency of KAT transfection followed by cocultivation is 1 to 3-fold greater than the transduction efficiency of cocultivation with a stable PA317 producer when 3T3 cells are used (compare 1B and 2B with 1C, table 4) and 5-10 fold greater when 293 cells are used. This data confirms that 293 cells have unique properties that support high efficiency transduction of mammalian cells.

35

While not wishing to be limited to any particular theory of the invention, these results suggest that high

titer virus production into the culture media is not sufficient for efficient T cell transduction and that the high efficiency transduction observed is mediated by cell-cell contact of 293 cells and CD8+ T cells, resulting in up to ten-fold greater efficiencies.

The results presented in this example demonstrate that, in the absence of selection, 10-40% of the CD8+ T cells were virally transduced, a significantly greater transduction frequency compared to prior results.

EXAMPLE III

Transduction of Primary Human Hematopoietic Stem Cells

15 This example describes the use of the KAT constructs and method of the invention to transduce primary human CD34+ bone marrow stem cells.

Preparation of Bone Marrow Cells

20

Human bone marrow was obtained from healthy volunteers. It was first fractionated into a mononuclear cell fraction over a Ficoll gradient (Pharmacia, Piscataway, NJ). The CD34+ cells are isolated using positive selection on a CellPro CEPTRATE LC™ affinity column (CellPro, Bothell, WA). Post-purification FACS analysis provided a population of approximately 90% CD34+ cells. This population of cells was then plated in 24 well plates at a density of 5×10^5 cells/ml in Myeloid Long Term Culture Medium supplied as a complete medium from Terry Fox Labs, (Vancouver, Canada) in the presence of 100 ng/ml human Stem Cell Factor (hSCF) (R&D Systems, Minneapolis, MN) 50 ng/ml hIL-3, and 10 ng/ml hIL-6 for 48 hours.

35 Transduction of CD34+ bone marrow stem cells

293 cells were transfected by first plating at a density of 1×10^6 cells/6 well plate 48 hours prior to transfection, followed by transfection with 10 μ g each of pRTD2.2F3 and pIK6.1MCVampac. Twenty-four hours later, 5 transfection media was removed, replaced with T cell growth media, as described in Example II, plus 50 ng/ml hIL-3, 100 ng/ml hSCF, and 10 ng/ml hIL-6. Two to four hours later, the transfected 293 cells were cocultivated with 5×10^5 purified CD34+ cells/well in the presence of 8 μ g/ml polybrene. After 10 48 hours, the cells were collected off of the 293 monolayer, and replated in Myeloid Long Term Media with growth factors as described above. Cultures were replenished with media plus growth factors daily via demi-depopulation. Four days later, the media was replenished and G-CSF was added at 2 ng/ml plus 15 20 ng/ml hSCF to promote differentiation into granulocytes. Four to six days later, cells were analyzed for surface expression of human CD4 from the transduced gene and CD15, a granulocyte marker. In addition, DNA was prepared for Southern blot analysis.

20

Figure 5 shows the FACS analysis of the transduced hematopoietic stem/progenitor cells after 14 days of growth and differentiation into granulocytes. Panel A shows the forward and side scatter gates used in the analysis of all 25 cell populations in the Figure. In panel B are shown the untransduced cells stained with the isotype control antibodies (FITC and PE). In panel C are shown the untransduced cells stained with antibodies for human CD4 (transduced gene, y axis) and CD15 (granulocyte 30 differentiation marker, x axis). In panel D, KAT packaging system was used in conjunction with 293 cell co-cultivation to transduce the hematopoietic stem/progenitor cells. A comparison of the top right quadrant for panels C and D indicate that 5-6% of the transduced cells expressed the CD4 35 protein.

Southern blot analysis of transduction efficiency

Southern blot analysis was carried out to determine whether the hematopoietic stem/progenitor cells were infected by retrovirus produced with the KAT system. Genomic DNA was prepared from differentiated stem cells and digested with Eco RV. 10 µg of DNA from infected (Figure 6, lane 2) and control cells (Figure 6, lane 1), as well as Eco RV-digested plasmid DNA equivalent to 0.12, 0.6, 1.2 and 6.0 copies per diploid genome of pRTD1.2F3 (Figure 6, lanes 4-7) and 5 copies per diploid genome of pRTD2.2F3 (Figure 6, lane 8) were electrophoresed on a 0.8% agarose gel, transferred to Zetabind and probed with a 605 base pair fragment encoding the zeta transmembrane and cytoplasmic domains. Eco RV digestion of the transfected plasmid pRTD2.2 yields a 4.2 kb band (Figure 6, lane 8). Eco RV digestion of pRTD1.2, which contains MMLV 5' and 3' LTRs, yields a 3.6 kb fragment (Figure 6, lanes 4-7). Following virus infection, integration and duplication of the 3' LTR, Eco RV digestion should yield a 3.6 kb fragment. In infected CD34+ cells, the probe hybridized to the appropriate 3.6 kb band, corresponding to integrated provirus (Figure 6, lane 7). Control cells lacked a proviral band, however the probe hybridized to bands that corresponded to the endogenous zeta gene sequences (Figure 6, lane 8). Scanning densitometry was used to quantitate transduction efficiency and demonstrated that the average proviral copy number per cell in infected cells was 0.5 (50% transduction). In addition, densitometry of the endogenous bands confirmed that equal amounts of DNA were loaded in the lanes corresponding to infected and uninfected cells.

In a second experiment, the transduction efficiency of a high titer PA317 producer clone was compared to the transduction efficiency of virus produced by the KAT system. 293 cells were transient cotransfection with pIK6.1MCVampac and pRTD2.2F3, isolation of CD34+ cells, cocultivation, purification of infected cells was carried out as described

above. Clone 40.39, described above in Example II, was plated at 5×10^5 cells/6 well plate 24 hours prior to initiation of cocultivation with CD34+ cells. Isolation of CD34+ cells, cocultivation, purification of infected cells was carried out as described for 293 cells. Transduction efficiency was analyzed by southern blotting of Eco RV digested DNA as described above and is shown in Figure 7. The band present in DNA isolated from CD34+ cells cocultivated with KAT plasmids hybridized to a 3.6 kb band (Figure 7, lane 2), identical in size to Eco RV digested plasmid DNA (Figure 7, lanes 4-7) and corresponding to integrated provirus. Hybridizing bands were absent from DNA isolated from CD34+ cells cocultivated with either mock transfected 293 (Figure 7, lane 1) cells or 40.39 cells (Figure 7, lane 3). The plasmid standards ranged from 0.3 to 10 copies of integrated provirus per cell. Therefore, the absence of a band in the PA317 lane suggests that KAT transduction is at least 10-fold more efficient.

Although FACS analysis of surface expression of the transduced gene indicates only a 5-6% efficiency of transduction, Southern analysis indicates a much higher efficiency of transduction (50-100%). It is possible that the level of expression of the human CD4 protein is below the level of detection of the FACS analysis, alternatively, the gene may be present but not efficiently expressed.

Modifications to the constructs could be made to increase the level of expression. The high efficiency of transduction of human hematopoietic stem/progenitor cells via the KAT packaging system in conjunction with 293 cell co-cultivation is contrasted to the transduction efficiencies obtained using traditional mouse fibroblast packaging systems such as PA317, Figure 7. The data from the PA317 packaging line indicates that although high titer virus can be generated when transducing mouse cells, the transduction efficiency of human bone marrow stem/progenitor cells is poor.

These results demonstrate, that, in addition to rapid production of high titer viral supernatants, the KAT constructs can be used to transduce at high efficiencies target cells, such as human T cells and hematopoietic cells, that are refractory to transduction by conventional methods.

EXAMPLE IV

Production of High Titer Virus in Human Cells with pIKT 10 Retroviral Vectors and High Efficiency Transduction of Human CD34+ Hematopoietic Cells

This example describes the use of the novel retroviral vectors of the invention to obtain high titer virus in a human cell line and the use of that virus to obtain high efficiency transduction in primary human hematopoietic stem cells.

The packaging vector pIK6.1MCVampac UTA described above and the retroviral vector pIKT2.2SVGe-F3 were transiently co-transfected (as described above) into human tsa54 cells as described above. tsa54 cells were derived from 293 cells by the transfection of the Large SV40 T antigen (Heinzel et al., J. Virol. 62(10):3738-3746 (1988)). pIKT2.2SVGe-F3 differs from pRTD2.2F3 in that the plasmid backbond contains the SV40 origin of replication as described above. This results in high copy number plasmid replication in tsa54 cells containing the SV40 t-antigen. tsa54 cells were transfected, viral supernatants were harvested and used to infect 3T3 cells as described above. 38% CD4 positive cells/100 μ l frozen viral supernatant equivalent to 7×10^6 /ml.

The pIKT vectors were used to produce retrovirus in tsa54 cells as described above and used to transduce primary human CD34+ bone marrow stem cells by co-cultivation. The bone marrow stem cells were purified and transduced with the pIKT2.2SVGe-F3 as described above in Example III with the

following changes. tsa54 cells were transfected at a density of 5×10^5 cells/6 well plate. The media used to replace the transfection media was IMDM + 10% FBS (fetal bovine serum). CD34+ cells were removed following two days co-cultivation with virus-producing tsa54 cells and cultured in Meloid Long Term Media with growth factors. Eight to ten days later G-CSF was added at 2 ng/ml plus 10 ng/ml hSCF to promote differentiation. Cells were analyzed for surface expression of human CD4 six to eight days later.

10

Figure 8 shows the results of FACS analysis of the transduced hematopoietic stem/progenitor cells after 18 days of growth and differentiation into granulocytes. Panel A shows the forward and side scatter gates used in the analysis of all cell populations in the Figure. In panel B are shown the untransduced cells stained with the isotype control antibodies (FITC and PE). In panel C are shown the untransduced cells stained with antibodies for human CD4 (transduced gene, y axis) and CD15 (granulocyte differentiation marker, x axis). In panel D a PA317 clone (78.81), with a titer equivalent to 107 near CFU clones/ml on 3T3 plates, was used as a stable viral producer in a co-cultivation with the hematopoietic stem cells. In panel E, KAT packaging retroviral vectors were used in conjunction with 293 cell co-cultivation to transduce the hematopoietic stem/progenitor cells. A comparison of the top right quadrant for panels C and D, and C and E indicates that 1.7% of the PA317 transduced cells expressed the CD4 protein as compared to 24% of those transduced using the KAT constructs.

30

EXAMPLE V

Production of a Single Vector 293 or tsa54 Stable Packaging Clone

35

This example describes the production and use of a single vector packaging clone. Human 293 or tsa54 cells were

plated at 5×10^5 per 10 cm plate in DME (JRH Biosciences, Lenexa, Kansas), 1 g/l glucose, 10% Donor calf serum (Tissue Culture Biologics, Tulare, CA) 48 hours prior to transfection. 10 μ g MCV ampac UTA and 0.1 μ g MC1 neo (Thomas 5 and Capecchi Cell 51:503-512 (1987)) were cotransfected by calcium phosphate precipitation (Wigler et al. Cell 16:777 (1979)). Clones could were also generated as efficiently by co-electroporation of the vectors (Shigekawa and Dower Biotechniques 6(8):742-751 (1988)). 18 hours post-
10 transfection the media was changed. 24 hrs. later the cells were split to duplicate plates of 1:10, 1:20 and 1:50 in media plus 1 mg/ml G418 (Geneticin, GIBCO, Grand Island, NY). Media was changed every 3 days for 14 days. Clones were picked to 24 well plates and grown to confluence. Media was
15 collected from wells, filtered through a .45 μ m filter and flash frozen on dry ice. Cells were resuspended in media plus 10% DMSO (Sigma Chemical Co., St. Louis, MO), frozen on dry ice and stored at -70° C. Supernatants were assayed for production of empty viral particles using an assay for
20 reverse transcriptase which measures the incorporation of radiolabeled thymidine into an RNA template. (Goff et al. J. of Virol. 38:239-248 (1981)).

Clones with the strongest reverse transcriptase
25 signals following autoradiography were thawed, grown up and tested for virus production following transient transfection. Transfections were done as previously described, using 10 μ g 43.3PGKF3. Media was changed 18 hours post-transfection. After 24 hours the viral supernatants were collected,
30 filtered through .45 μ m filters and flash frozen on dry ice. Viral supernatants were assayed on 3T3 cells plated at 5×10^5 per 10 cm plate 24 hours prior to infection. Infections were done as described above. Cells were then harvested, stained with OKT4A anti-CD4 monoclonal antibody (Ortho Diagnostic
35 Systems Inc., Raritan, NJ), and analyzed by flow cytometry as described above. Clones displayed varying amounts of

packaging function. Those clones with the highest transient titer were selected for further characterization (Table 5).

Table 5

CELLS/CLONE #	VECTORS	Transduction Efficiency (% CD4+ 3T3 Cells)
tsa54	none	1.02
tsa54/107.14	ampac + 43.2	2.53
tsa54/107.17	ampac + 43.2	0.67
tsa54/107.18	ampac + 43.2	29.76
tsa54/107.22	ampac + 43.2	1.76
tsa54/107.24	ampac + 43.2	6.47
tsa54/107.25	ampac + 43.2	0.61
tsa54/107.26	ampac + 43.2	1.70
tsa54/107.49	ampac + 43.2	0.89
tsa54/107.57	ampac + 43.2	1.12
tsa54/107.73	ampac + 43.2	1.33
tsa54/107.75	ampac + 43.2	13.18
tsa54/107.142	ampac + 43.2	0.98
293/90.74	ampac + 43.2	24.80
293/90.85	ampac + 43.2	15.06

Clones 90.74, 107.75, and 107.18 were carried for extended time in culture to study ability to maintain the packaging genome over time in the presence or absence of G418. Cells were split 1:10 to 1:20 every 3 to 4 days. At passages 1, 6 and 12 cells were transfected with 10 μ g 43.2 as described above, and transient viral supernatants were analyzed by infection of 3T3 cells as described above. Of the three clones studied, only one (90.74) appeared to have consistent titer over 12 passages (Table 6). It also appeared that titer did not depend on continued G418 selection. Clone 90.74 has a transient titer equivalent to approximately 107/ml. Clone 90.74 has been deposited with

the ATCC, 12301 Parklawn Drive, Rockville, Maryland, under the Budapest Treaty, and has there been identified as follows:

5	<u>Cell Line</u>	<u>ATCC Accession No.</u>	<u>Deposit Date</u>
	90.74	CRL11654	June 10, 1994

Table 6

tsa AMPAC STABILITY TEST
Transduction Efficiency
(% CD4+ 3T3 Cells/0.25 ml supernatant)

	CLONE	VECTORS	p1	p6	p12
	tsa	none	0.10		1.20
	tsa	Ampac	0.10		4.06
15	tsa	Ampac+43.2	63.90	30.97	66.94
	107.18+G418	Ampac+43.2	41.50	22.35	22.68
	107.18	Ampac+43.2	-	23.50	26.22
	107.75+G418	Ampac+43.2	15.42	12.18	4.36
	107.75	Ampac+43.2	-	11.17	2.05
20	90.74+G418	Ampac+43.2	34.83	30.79	27.79
	90.74	Ampac+43.2	-	28.40	31.32

Supernatants were also analyzed for production of RCR by a S+/L-assay on PG4 cells. PG4 cells are Moloney sarcoma virus-transformed cells from cat brain (ATCC CRL2032). When infected with competent murine retrovirus, PG4 cells produce discernable foci which can be distinguished microscopically (Haapala, et al. J. of Virol., 53(3):827-833). PG4 cells were seeded at 5×10^6 on 10 cm plates 24 hours prior to infection. Infections were done with 1 ml of test supernatant and 4 mls of media containing 8 μ g/ml polybrene. Media was changed 24 hours later, and then replaced every 2-3 days until foci developed on positive control plates. All clones studied remained RCR-free through 12 passages.

Unexpectedly, these results demonstrate that using the retroviral vectors of the invention, stably transfected 293-

derived cell lines that produce gag, pol and env proteins were generated. The virus production from these cell lines was equivalent to that produced from transient co-transfection of packaging and retrovirus vectors. Moreover, 5 surprisingly, in the absence of drug selection, these cell lines maintained production of gag, pol and env proteins. Previous attempts to generate 293-based retroviral producers using retrovirus constructs described in the literature have failed (Pear et al. Proc. Nat'l. Acad. Sci. (USA) 90:8392-10 8396 (1993)). After extended passage in culture these packaging cell lines do not spontaneously generate replication competent retrovirus.

EXAMPLE VI

15 Production of Double Genome Stable Packaging Cells

This example describes the construction and use of two genomes in 293 or tsA54 packaging cells. First a gag/pol clone was created in human tsA54 cells. Cells at 1×10^6 per 20 0.8 ml of PBS were co-electroporated with 15 μ g notI linearized gag/pol ATG (described above) and 1 μ g MC1 neo. Electroporation was done at 960 μ F and 260 mV (Shigekawa and Dower, (1988) supra) on a Gene Pulser (Biorad, Richmond, CA). Cells were immediately plated on a 10cm plate in DME, 1 g/l 25 glucose, 10% donor calf serum for 48 hours. Cells were then split 1:5, 1:10, 1:20 and 1:50 in 1.0 mg/ml G418 selection. Media was changed every 3 days, and after 12 days of selection in G418 clones were picked to 24 well plates. Once cells were confluent, media was collected, filtered through a 30 0.45 μ m filter and flash frozen on dry ice. Clones were trypsinized and frozen at -70° C. Supernatant was thawed and analyzed for reverse transcriptase activity (Goff et al., (1981) supra). Those clones displaying the highest RT activity were grown up and evaluated for transient virus 35 production by calcium phosphate transfection of 5 μ g pIK6.1MCMenvATGUTΔ described above and 10 μ g pRT43.2F3 described above. Media was changed after 18 hours, and after

an additional 24 hours the viral supernatants were collected, filtered and frozen for analysis by infection of 3T3s. Transient virus titers were comparable to the transient virus titer of the single genome pack line 90.74 transfected with 5 pRT43.2F3. and approximately 50% of the viral titer following co-transfection of tsa54 cells with pIK6.1MCVampacUTA and pRT43.2F3 (Table 7).

Table 7
GAG/POL TRANSIENT TEST

10

15

20

25

CLONE	VECTORS	Transduction Efficiency (% CD4+ 3T3 Cells)
tsa		0.02
293/90.74	ampac + 43.2	28.50
tsa54/111.3	gag/pol + 43.2	11.72
tsa54/111.8	gag/pol + 43.2	13.30
tsa54/111.44	gag/pol + 43.2	20.29
tsa54/111.89	gag/pol + 43.2	17.93
tsa54/111.4	gag/pol + 43.2	32.95
tsa54/111.47	gag/pol + 43.2	19.53
tsa54/111.25	gag/pol + 43.2	21.18
tsa54/111.45	gag/pol + 43.2	14.22
tsa54/111.43	gag/pol + 43.2	23.75
tsa54/111.22	gag/pol + 43.2	24.27

The four best clones were selected for long term stability studies with and without G418 selection. They were also assayed on PG4 cells for the production of RCR and are 30 RCR negative.

Clone 111.4 is co-transfected with pIK6.1MCVamenvATGUTA and sv2his (Hartman and Mulligan, Proc. Nat'l. Acad. Sci. (USA) 85:8047-8051 (1988)) selected in 35 histinol as described. Clones are picked and characterized for virus production by transient transient transfection as

described above. Several high titer clones are characterized for stability and RCR as described.

Packaging lines can be created by replacing the
5 amphotropic env gene in pIK6.1MCVamenvATGUTΔ with other retroviral envelopes, for example, ecotropic, xenotropic, polytropic, MLV, 10A1, Gibbon ape leukemia virus, feline leukemia virus C, Vesicular stomatitis virus (VSV) G protein, human T cell Leukemia (HTLV) I and II, and combinations
10 thereof, using the methods described above.

EXAMPLE VII

Construction of Packaging Plasmids and Production of Viruses of Different Tropisms

15

a) Xenotropic packaging plasmid constructions

This example describes the construction of packaging plasmids which encode a xenotropic envelope protein to allow
20 the production of retroviruses with expanded host range. The envelope protein for these packaging plasmids was derived from xenotropic NZB virus (O'Neill et al., J. Virol., 53(1):100-106 (1985)).

25 pIK6.1MCVxenopac contains gag/pol from pIK6.1MCVampacUTΔ as well as the xenotropic envelope protein. It was constructed by replacing the 4061 Sall-Nhe fragment of pIK6.1MCVampacUTΔ (described previously in the detailed description) with the 4200 base pair Sall - NheI fragment
30 from the NZB9-1 xenotropic virus (O'Neill et al., supra).

pIK6.1MCVxenopacUTΔ encodes the ecotropic MMLV gag/pol gene and the NZB xenotropic envelope coding region linked to the SV40 polyadenylation sequence. This plasmid was
35 constructed by deleting untranslated sequences 3' from the envelope gene of pIK6.1MCVxenopac by performing a PCR reaction using pIK6.1MCVxenopac as the template with

synthetic oligonucleotides 5' GACCACACTGGCGTAGTAAG 3' (SEQ ID NO 33) and 5' GAATTCGCTAGCTTATTCACGCGATTCTACTTC 3' (SEQ ID NO 34). The resulting 340 base pair fragment was digested with BstB1 and Nhe 1 and the 250 base pair product was isolated 5 and used to replace the 312 base pair BstB1 - Nhe 1 fragment of pIK6.1MCVxenopac.

pIK6.1MCVxenoenvUTA is a packaging plasmid which encodes only the NZB xenotropic envelope as a packaging 10 function. This plasmid was constructed as follows: The ATG at the translational start of the xenotropic env gene was converted to an Nco1 site by PCR using pIK6.1MCVxenopacUTA as the template with synthetic oligonucleotides 5' GAATTCATGGAAGGTTTCAGCGTTCTC 3' (SEQ ID NO 35) and 5' 15 CGTTAGCTGTTTGTCTGTC 3' (SEQ ID NO 36) followed by digestion with Nco1 and Bgl11. The resulting 120 base pair fragment was purified and ligated in a 4 part ligation with a 450 base pair Bgl11 - EcoR1 fragment from pIK6.1MCVxenopacUTA, a 4541 base pair EcoR1 - Hind111 from pIK6.1MCVxenopacUTA and a 916 20 base pair Hind111 - Nco1 fragment from pIK6.1MCVgag/pol ATG to produce pIK6.1MCVxenoenvUTA.

b) Ecotropic packaging plasmid constructions

25 This example describes the construction of packaging plasmids which encode an ecotropic envelope protein from ecotropic MMLV (Shinnick et al., Nature, 293:543-548(1981)).

pIK6.1MCVecopac contains the gag/pol genes as well as 30 the ecotropic envelope protein. It was constructed by replacing the 4175 base pair Sal1-Nhe1 fragment of pIK6.1MCVampac with the 4141 basepair Sal1 - Nhe1 fragment from ecotropic MMLV described previously in the detailed description.

35

pIK6.1MCVecopacUTA contains a deletion in the sequences 3' of the envelope gene. It was constructed by

replacing the 4003 Sall-ClaI fragment of pIK6.1MCVampacUTΔ with the 3969 base pair Sall - ClaI fragment from pIK6.1MCVecopac.

5 pIK6.1ecoenvUTΔ is a packaging plasmid which encodes only the ecotropic envelope as a packaging function. It was constructed by replacing the 1405 base pair HhaI-ClaI fragment of pIK6.1amenvATGUTΔ with the 1912 base pair HhaI - ClaI fragment from pIK6.1MCVecopac in a 3-way ligation with a
10 3368 base pair ClaI-SpeI fragment and a 889 base pair SpeI-HhaI from pIK6.1amenvATGUTΔ.

c) Polytropic packaging plasmid construction

15 This example describes the construction of a packaging plasmid which encodes a polytropic envelope protein. pIK6.1MCVpolypac contains a pol/env fragment from polytropic MCF 247 virus (Holland et al., J. Virol., 47:415-420 (1983)). It was constructed by replacing the 4061 Sall-Nhe I fragment
20 of pIK6.1MCVampacUTΔ with the 4200 base pair Sall - NheI fragment from MCF 247 (Holland et al., supra).

d) 10A1 packaging plasmid construction

25 This example describes the construction of a packaging plasmid which encodes an amphotropic envelope protein. pIK6.1MCV10A1pac contains a pol/env fragment from a recombinant amphotropic MMLV isolate, 10A1 (Ott et al., J. Virol., 64(2):757-766 (1990)). It was constructed by
30 replacing the 4003 Sall-ClaI fragment of pIK6.1MCVampacUTΔ with the 4000 base pair Sall - ClaI fragment from pB6 (Ott et al., supra).

e) Supernatant Transduction of a Wide Variety of Cell Types

This example demonstrates that supernatants from 293 derivatives transiently transfected with pRT43.2F3 and packaging plasmids expressing various viral envelopes of a variety of tropisms can efficiently transduce a variety of other mammalian cells in addition to 3T3 (mouse fibroblasts) and human T cells described in the previous examples. Other cells tested include CF2Th (dog thymus), 143B (human osteocarcinoma), HT1080 (human fibrosarcoma), *M. dunni* (mouse fibroblasts) and 293 (human embryonic kidney).

In Table 8 below 1×10^6 of the indicated cells were transduced with 1 ml (* 10ul) of supernatant from 293 derivatives transiently transfected with pRT43.2F3 and the indicated packaging plasmid.

Table 8

Packaging Plasmid	3T3 (% transduction)	CF2Th (% transduction)	143B (% transduction)	HT1080 (% transduction)	<i>M. dunni</i> (% transduction)	293 titer (%transduction)
pIK6.1MCVampac UTΔ	73	76	76	78	79	58
pIK6.1MCVxenopa c	2.7	36	50	66	67	65
pIK6.1MCV10A1pa c	63	50	71	68	75	60
pIK6.1MCVpolypac	40	1.3	4.0	3.1	64	23
pIK6.1MCVecopac*	10	2.2	nd	nd	7.2	nd

nd = not determined

As shown in examples a) through e), the above described plasmids encoding envelope genes of a variety of tropisms can be substituted for plasmids encoding an

amphitrophic envelope and used in the instant invention to generate retroviral particles capable of infecting a wide variety of mammalian cell types.

5

EXAMPLE VIIIProduction of Stable 293 Viral Producer Clones

This example describes the construction of stable 293 viral producer clones. These stable producer clones can be created from stable packaging clones either by transfection with retroviral vector or by infection with retrovirus.

In the first method (transfection with a retroviral vector), the 293 amphotropic packaging clone 90.74, described in Example V, was plated at 6.5×10^5 per 10 cm plate in DME (JRH Biosciences, Lenexa, Kansas), 1 g/l glucose, 10% Donor calf serum (JRH Biosciences, Lenexa, Kansas) 48 hours prior to transfection. 10 μ g pRT43.2F3 (a retroviral vector containing the CD4/zeta chimeric receptor described supra) and 0.5 μ g SV2 hyg were cotransfected by calcium phosphate precipitation (Wigler *et al.* Cell 16:777(1979)). pSV2hyg was derived from pSV2DHFR (Subramani *et al.* Mol. and Cell. Biol. 1:854-864 (1981)) in the following fashion. First, pSV2DHFR was digested with Hind III and filled in with the Klenow fragment of DNA polymerase I (New England Biolabs, Beverly MA) and a mixture of the four deoxynucleotide triphosphates. Next, BgII 8-mer linkers (New England Biolabs, Beverly MA) were ligated to the blunt ends, followed by Bgl II digestion to remove extra linkers and the DHFR cDNA. The vector backbone was isolated and ligated to the Bgl II/Bam HI fragment encoding the hygromycin phospho-transferase gene from pTG4 (Giordano and McAllister Gene 88:285-288(1990)), resulting in pSV2hyg. Eighteen hours post-transfection the media was changed. Twenty four hours later the cells

were split to duplicate plates of 1:10, 1:20, 1:50 and 1:100 in media plus 200 μ g/ml hygromycin B (Boehringer Mannheim, Indianapolis, IN). Media was changed every 3 to 4 days for 14 days. Clones were picked to 24 well
5 plates and grown to confluence. Media was collected from wells, filtered through a 0.45 μ filter and flash frozen on dry ice. Cells were resuspended in media plus 10% DMSO (Sigma Chemical Co., St. Louis, MO), frozen on dry ice and stored at -70° C.

10

The second method for the creation of stable producer clones is by serial infection of the stable packaging clone with a transient supernatant containing viral particles that display a tropism different from
15 that of the stable packaging clone. Tsa54 cells were plated at 6.5×10^5 per 10 cm plate for 48 hours prior to transfection. 5 μ g of pIK6.1MCVxenopacUT Δ and 10 μ g of pRT43.2F3 were cotransfected by calcium phosphate precipitation as previously described. Eighteen hours
20 post-transfection the media was changed and 24 hours later the media was collected, filtered through 0.45 μ filters and frozen on dry ice. The 90.74 amphotropic packaging clone was plated at 2×10^6 cell per 10 cm plate and 24 hours later infected with 2 ml of the
25 transient supernatant, 3 mls of medium and 8 μ g/ml of polybrene. Twenty four hours later the media was changed and the cells were grown to confluence, at which time the cells were split 1:10 and grown to confluence. The cells were subsequently serially re-infected as above with the
30 xenotypic envelope-containing retrovirus in the transient supernatants, for a total of 8 serial infections. After eight serial infections the population was cloned by limiting dilution in 96 well plates. Clones were transferred to 24 well plates, grown to confluence, and
35 the supernatants filtered with 0.45 μ filters and frozen on dry ice. The cells were resuspended in media plus 10% DMSO, frozen on dry ice and stored at -70° C.

Supernatants from clones produced by either method were assayed for viral particles containing CD4/zeta by viral RNA dot blots as follows. Lysis buffer was added to thawed supernatants for final concentrations of 500 µg/ml. proteinase K (Boehringer Mannheim, Indianapolis, IN), 100 µg/ml. tRNA (Sigma, St. Louis, MO), 2.5 mM EDTA pH 7.5 and 0.5% SDS and incubated for 45 min at 37 °C. The lysate was then extracted with an equal volume of phenol, followed by an equal volume of chloroform. The lysate was split to 2 samples, brought to a final concentration of 375 mM NaCl and vortexed. 1 ml of chilled ethanol was added to each sample and the RNA was precipitated over night at -70°C. The lysate was thawed and spun at full speed in a microfuge for 10 minutes. The supernatant was discarded, the RNA pellet drained, dried, and then resuspended in 20 µl 2 mM EDTA pH 7.5, vortexed and heated for 5 minutes at 65°C. 37.5 µl of formamide (USB, Cleveland, OH) and 12.5 µl of formaldehyde (Mallinckrodt Chemical, Paris, KY) were added and the solution was incubated at 50°C for 20 minutes. Finally 100 µl of filtered 10X SSC (1.5M NaCl, 150mM sodium citrate pH 7.0) was added and the samples were spotted on a nylon filter through the use of a dot blot apparatus. The wells were washed twice with 10X SSC. The blotter was then dismantled, and the RNA was UV crosslinked to the nylon at 1600 µJ (Stratagene, San Diego, CA). The filter was hybridized with a DIG-labelled (digoxigenin, Boehringer Mannheim, Indianapolis, IN) CD4/zeta probe, and hybridization detected using the Boehringer Mannheim Genius System. Clones with the strongest CD4/zeta signals were assayed for viral titer on 3T3 cells and for RCR as previously described.

Supernatants were assayed for RCR by S+/L- PG4 assay as described previously in Example V. At passage 13 the clones were expanded to a 850 cm² roller bottle and supernatant collected at confluence. The entire

supernatant was inoculated on *Mus dunni* cells, which were grown for 2 passages and their final supernatants were then assayed for RCR by PGA S+/L- assay. The clones were all negative for RCR.

5

Clones with the highest titers were chosen and further characterized for stability of virus production by passaging the clones twice weekly for six weeks, plus and minus the appropriate selection drug (hygromycin B for the transfected clones, G418 for the infected clones). Viral titers on 3T3 cells are shown in Table 9 below for passages 1, 6 and 12 (p1, 6 and 12).

10

Table 9: Stability of CD4/Zeta Producer Clones Derived from 90.74

15

(Viral titers = % CD4+ 3T3 cells / 0.01 ml supernatant / 10^6 cells)

clone	method of producing clones	p1 titer x 10^6 /ml	p6 titer x 10^6 /ml	p12 titer x 10^6 /ml
143.15	transfection	1.94	5.05	3.65
143.15 +Hyg	transfection	ND	4.91	4.47
143.43	transfection	0.85	3.38	2.88
143.43+Hyg	transfection	ND	3.94	2.80
143.64	transfection	1.37	4.42	3.26
143.64 +Hyg	transfection	ND	4.27	3.79
143.84	transfection	1.56	3.70	2.65
143.84 +Hyg	transfection	ND	5.50	3.40

20

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5	143.86	transfection	2.59	5.50	4.45
	143.86 +Hyg	transfection	ND	4.02	2.90
10	143.90	transfection	1.53	4.00	2.53
	143.90 +Hyg	transfection	ND	2.73	2.15

15	clone	method of producing clones	p1 titer x 10 ⁶ /ml	p4 titer x 10 ⁶ /ml	p8 titer x 10 ⁶ /ml	p12 titer x 10 ⁶ /ml
20	142H.15	infection	24.30	14.78	11.72	13.57
	142H15+ G418	infection	ND	16.56	15.04	14.04
25	142H.34	infection	9.43	11.17	8.63	8.20
	142H.34+ G418	infection	ND	9.89	12.10	10.00
30	142H.62	infection	19.61	16.63	12.44	13.23
	142H.62+ G418	infection	ND	15.16	14.05	11.62
35	142H.69	infection	7.47	10.44	8.69	7.43
	142H.69+ G418	infection	ND	9.62	11.90	9.48

Table 9 demonstrates that both methods for creating producer clones (via transfection or infection) resulted in clones that had stable virus production for 6 weeks both in the presence or the absence of selection.

Also, the 142H and 143 clones were RCR-free (data not shown). The 3T3 titers of the infection amplified clones (142H) were 2-3 fold greater than the transfected clones (143).

The producer clone supernatants were further characterized by transducing human primary CD8+ T cells that were isolated as previously described. CD8+ cells were plated at 10^6 cell/ml/well of a 24-well plate in AIM V + 100 Cetus units/ml IL-2 (Chiron, Emeryville, CA) 24 hours prior to transduction. For transduction the cell volume was reduced to 0.5 ml and 0.75 ml of appropriate supernatant and 0.75 ml of AIM V + 200 Cetus units/ml IL-2 + 4 μ g/ml polybrene were added for 4-24 hours at 37° C. This was done once a day for three days. After the third day cells were returned to growth media (50% RPMI, 50% AIM V, 5% human serum (Sigma Chemical Co., St. Louis, MO)) for an additional three days and then analyzed for surface CD4 as described previously. The 293 supernatants were compared with supernatant from 78.81, a CD4/zeta producer clone generated by amplification co-cultivation (Bestwick et al., Proc. Natl. Acad. Sci., USA, 85:5404-5408(1988)) from the 3T3-based PA317 packaging line (Miller et al, U.S. Patent #4,861,719). Table 10 below demonstrates transduction efficiency of CD4/zeta producer clones on human CD8+ T lymphocytes.

Table 10: Human Primary CD8+T Cell Transduction by Supernatants of Transfected or Infected Amplified Clones

Clones	packaging clone	method	3T3 titer x 10^6 /ml	T cell transduction (%CD4+)
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	78.81	PA317	Infection	2.86	13.97
5	142H.15	90.74	Infection	13.16	60.55
	142H.34	90.74	Infection	6.14	45.44
10	142H.62	90.74	Infection	11.32	88.63
	142.69	90.74	Infection	5.22	48.99
15	143.15	90.74	Transfection	3.5	42.65
	143.43	90.74	Transfection	1.87	14.26
20	143.64	90.74	Transfection	3.5	51.48
	143.84	90.74	Transfection	2.11	22.66
25	143.86	90.74	Transfection	3.66	58.14
	143.90	90.74	Transfection	3.69	41.87

30

The 293-based clones consistently provides
 higher titer on the 3T3 cells and a higher level of
 transduction of the T cells than the 3T3 based producer,
 78.81. These transduction results are 5 to 10-fold
 35 greater than those previously reported in the literature
 for the cocultivation of T cells with producer clones,
 and 40 to 80-fold greater than the 1-2% T cell

transduction by supernatants previously reported (Fauser, J. Cell. Biochem., 45:353-358 (1991), Hwu et al., J. Immunol., 150:4104-4115 (1993)), Imbert et al., Cancer Gene Therapy, 1:259-265 (1994)), and Mavillo et al., Blood, 83:1988-1997 (1994)).

Table 10 also demonstrates that the 3T3 titer does not predict T cell transduction. For example, although infection- amplified clone 142H.15 has 3-fold greater 3T3 titer than transfected clone 143.64, they have equivalent T cell transductions of 60.55% and 58.14%, respectively.

EXAMPLE IX

Construction of Stable Two Genome 293 Packaging Line

This example describes the creation of stable clones of 293 cells that contain two helper sequences encoding packaging functions. Applicants first constructed a stable gag/pol clone in 293 cells that was then used for the production of a variety of packaging clones with the different envelope encoding plasmids of Example VII.

To construct the gag/pol clone, 293 cells were plated at 6.5×10^5 for 48 hours, then cotransfected with 10 μ g pIK6.1MCVgag/polATG and 1 μ g MC1neo (Stratagene, La Jolla, CA) by calcium phosphate precipitation as previously described. Medium was changed after 18-24 hours, and after an additional 24 hours the cells were diluted 1:10, 1:20, 1:50, 1:100, 1:500 and 1:1000 into 1mg/ml G418 selection. Cells were fed every 3-4 days, and after 12 days clones were picked to 24-well plates. Once the cells were confluent, the medium was collected, 0.45 μ filtered and frozen on dry ice. The cells were frozen in medium plus 10% DMSO and stored at -70° C.

Supernatant was thawed and analyzed for reverse transcriptase activity (Goff et al, supra). Those clones displaying the highest reverse transcriptase activity were grown and evaluated for transient virus production by transfecting with 5 μ g of pIK6.1amenvATGUT Δ and pRT43.2F3 as described above. Supernatants were collected and assayed for CD4 titer by infection of 3T3 cells as previously described. The four clones with the best transient titers were studied for long term stability. Clones were passaged twice a week for 6 weeks, with and without G418 selection. Transient transfections were done at passage (p) 1, 5, 9 and 13 with pIK6.1amenvATGUT Δ and pRT43.2F3, and the supernatants were evaluated for titer on 3T3 cells. (Table 11).

Table 11: Stability of packaging cell clones encoding gag/pol

clone	p1 titer x 10 ⁶	p5 titer x 10 ⁶	p9 titer x 10 ⁶	p13 titer x 10 ⁶
35.32	0.73	2.01	1.29	1.53
35.32+G418	ND	1.95	1.77	1.60
35.35	0.21	0.87	1.23	1.11
35.35+G418	ND	0.74	1.14	0.92
35.74	0.46	0.99	1.11	0.64
35.74+G418	ND	1.07	1.90	1.07
35.88	0.30	0.64	0.47	0.28
35.88+G418	ND	0.59	0.34	0.15

Virus production from the clones was stable over 13 passages in both the presence and absence of G418. Clone supernatants were also evaluated for RCR by S+/L-

PG4 assay as previously described in Example V and found to be negative.

Gag/pol clone 35.32 was selected for further
5 transfection with envelope plasmids since it was
determined to be stable and RCR-free in long term
passage, and had the best growth characteristics. Clone
35.32 was plated at 6.5×10^5 48 hours prior to
transfection. Ten μg of pIK6.1amenvATGUT Δ + 0.5 μg of
10 SV2 hyg were cotransfected, media was changed at 18-24
hours, and after an additional 24 hours cells were split
into 200 $\mu\text{g}/\text{ml}$ hygromycin B (Boehringer Mannheim,
Indianapolis, IN). Medium was changed every 3-4 days and
at day 14 clones were transferred to 24 well plates.
15 Once the cells were confluent, the medium was collected,
0.45 μ filtered and frozen on dry ice. Cells from each
clone were divided in half, and half were frozen down in
medium +10% DMSO and stored at -70°C . The other half of
the cells were then analyzed for envelope protein
20 production using rat anti-gp70 antibody 83A25 (Evans *et*
al., *J. Virol.*, 64:6176-6183 (1990)). Cells were
trypsinized, washed with PBS plus 2% FBS, incubated with
47 μg antibody at 4° for 30 min, washed three times with
PBS/FBS, incubated with goat anti-rat IgG-PE at 0.5
25 $\mu\text{g}/\text{tube}$ (Biosource International, Camarillo, CA), washed
two times and resuspended in 0.1% formaldehyde. Cells
were then analyzed by flow cytometry. Supernatants were
thawed and analyzed for reverse transcriptase activity
(Goff *et al*, *supra*). Clones positive for gp70 and with
30 the highest reverse transcriptase levels were grown up
and transiently transfected with 10 μg of pRT43.2F3 as
described, and assayed for CD4 titer on 3T3 cells. The
clones with the highest titers were passaged twice a week
for six weeks with and without hygromycin B. Transient
35 transfections of 10 μg of pRT43.2F3 were done at p1 and
p5 (Table 12). Clone supernatants were also negative for
RCR by S+/L- PG4 assay.

Table 12: Stability of Two Genome Packaging Clones gag/pol+amenv

clones	p1 titer x 10 ⁶	p5 titer x 10 ⁶
37S2.8	1.01	1.39
37S2.18	0.35	0.60
37P2.4	0.30	0.90
37P2.9	0.19	0.34

Table 12 demonstrates that packaging clones containing the gag/pol and amphotropic envelope genes can be isolated which stably produce high titer retroviral supernatants. Two genome packaging clones containing a xenotropic envelope gene were also constructed as described above. In this case, 10 μ g of pIK6.1CMVxenoenvUT Δ + 0.1 μ g pIKpur was cotransfected. pIKpur was constructed by the insertion of a 600 base pair cDNA encoding Streptomyces alboniger puromycin-N-acetyltransferase (abbreviated pa, GenBank Accession No. M25346 nucleotides 254-853) into pIK 6.1, described previously. Cells were selected in 0.5 μ g/ml puromycin (Sigma, St. Louis, MO). Clones were then isolated, assayed for reverse transcriptase activity and envelope protein as above, and the clones positive for gp70 with the highest reverse transcriptase levels are chosen for long term stability studies. These clones are also evaluated for virus production by transient transfection of 10 μ g of pRT43.2F3. Those clones with the highest transient titers are maintained for six weeks in culture with and without puromycin, and assayed for virus production at passages 1, 5, 9 and 13 after transient infection with pRT43.2F3, as described above, to determine the stability of packaging function.

EXAMPLE X

Construction of a retroviral vector and packaging
plasmids containing the RSV Enhancer and Promoter

5 This example describes the construction of a
retroviral vector wherein the 5' LTR of the retroviral
vector contains the enhancer and promoter from the U3
region of the Rous Sarcoma Virus (RSV) joined to the R
region of MMLV and the construction of packaging plasmids
wherein the packaging functions are encoded by two
10 plasmid based expression vectors in which expression is
under the control of the enhancer and promoter from the
U3 region of the Rous Sarcoma Virus (RSV).

pRT43.RSV.F3 is a retroviral construct in which
15 the enhancer and promoter of RSV is joined to the R
region of MMLV as follows: pIK 6.1RSV was derived from
pIK6.1 by replacing the 679 base pair HindIII - XbaI CMV
IE enh/pro fragment of pIK6.1 with a 235 base pair
HindIII - Xba I RSV enhancer/promoter fragment generated
20 by PCR using pREP4 (Invitrogen Corp. San Diego, CA) as a
template with synthetic oligonucleotides 5' -
GAATTC AAGCTTAATGTAGTCTTATGCAAT 3' (SEQ ID NO. 37) and 5'
GAATTC TCTAGAGTTTATTGTATCGAGCTA 3' (SEQ ID NO. 38),
followed by digestion with HindIII and XbaI.
25 pRT43.RSV.F3 was derived from pRT43.2F3 by replacing the
710 base pair HindIII - Asp718 fragment of pRT43.2F3
with a 219 base pair HindII - TaqI fragment from pIK6.1
RSV and a 46 base pair TaqI - Asp718 synthetic
oligonucleotide (consisting of oligonucleotides 5'
30 CGATACAATAAACGCGCCAGTCCTCCGATTGACTGAGTCCCCGG 3' (SEQ ID
NO. 39) and 5'
GTACCCGGGCGACTCAGTCAATCGGAGGACTGGCGCGTTTATTGTAT 3' (SEQ ID
NO. 40) in a four - part ligation with a 1006 base pair
Asp718 - BglII fragment from pRT43.2F3 and a 6897 base
35 pair Bgl II - HindIII fragment from pRT43.2F3.

pIK6.1RSVgag/polATG is a packaging plasmid encoding the gag/pol genes under the control of the RSV U3 region. It was derived from pIK6.1gagpolATG by replacing the 1258 base pair AflIII - NsiI fragment of pIK6.1gagpolATG with the corresponding 806 base pair fragment from pIK 6.1RSV in a two part ligation.

pIK6.1RSVamenvATGUTΔ is a packaging plasmid encoding the amphotropic envelope gene under the control of the RSV U3 region. It was derived from pIK6.1amenvATGUTΔ by replacing the 1373 base pair AflIII - BglII fragment of pIK6.1amenvATGUTΔ with the 1085 base pair AflIII - BglII fragment of pIK6.1RSV in a three - part ligation with the 2714 base pair BglII - DraIII fragment and the 2033 base pair DraIII - AflIII fragment from pIK6.1amenvATGUTΔ.

The following table (Table 13) compares the titers of retroviral particles when the enhancer/promoter regions of the retroviral vectors and packaging plasmids are derived from RSV, MMSV or CMV. These results demonstrate that the vectors and packaging plasmids containing RSV LTRs are as efficient as those containing MMSV or CMV LTRs.

Table 13: Comparison of Retroviral Production in tsA54 Cells using RSV, MMSV or CMV Enhancer/Promoter Vectors

Retroviral Vector	Enhancer-Promoter/Packaging Protein	Titer/ml supernatant on 3T3 cells (expt.1)
pRT4.3.2F3(CMV)	MCV/gagpol MCV/amenv	3.4 x 10 ⁶
pIKT4.2 F3 (MMSV)	MCV/gagpol MCV/amenv	5.3 x 10 ⁶
pRT43.RSVF3	MCV/gagpol MCV/amenv	6.9 x 10 ⁶
pRT42.2F3 (CMV)	MCV/ampac	6.1x10 ⁶ (expt.2)

	pIKT4.2 F3 (MMSV)	MCV/ampac	1.3 x 10 ⁷
	pRT43.RSVF3	MCV/ampac	1.5 x 10 ⁷
5	pRT43.2F3 (CMV)	MCV/gagpol MCV/amenv	4.6 x 10 ⁶ (expt.3)
	pRT43.2F3 (CMV)	MCV/gagpol MMSV amenv	4.8 x 10 ⁶
	pRT43.2F3 (CMV)	MCV/gagpol RSV/amenv	3.2 x 10 ⁶
10	pRT43.2F3 (CMV)	RSV/gagpol MCV/amenv	1.6 x 10 ⁶
	pRT43.2F3 (CMV)	RSV/gagpol MMSV/amenv	1.7 x 10 ⁶
	pRT43.2F3 (CMV)	RSV/gagpol RSV/amenv	2.3 x 10 ⁶
15	pIKT4.2 F3 (MMSV)	RSV/gagpol RSV/amenv	2.4 x 10 ⁶
	pRT43.2F3 (CMV)	MCV/gagpol MCV/amenv	6.5 x 10 ⁶ (expt.4)
20	pRT43.RSVF3	RSV/gagpol RSV/amenv	5.1 x 10 ⁶

EXAMPLE XIHigh Level Supernatant Transduction of Human CD8+ T Cells

In this example, Applicants demonstrate the high level transduction of CD8+ T-cells from different donors with retroviral supernatants from 293-derived cells which were either stably or transiently transfected with the retroviral vectors and packaging plasmids of the instant invention. Transduction efficiencies of retroviral supernatants from transient and stable 293-derived cells are compared with supernatants from 3T3-derived stable packaging cells.

For transient viral production, tsA54 cells are seeded at 0.6×10^6 /10cm plate 48 hrs prior to CaPO_4 transfection with 5ug of packaging and 10ug of retroviral plasmid. Twenty four hours post transfection, the media is exchanged for fresh media. Forty eight hours post transfection, supernatant was harvested, filtered through 0.45 micron filters, stored at -70°C and thawed immediately before use.

The supernatants were characterized by transducing human primary CD8+ T cells that were isolated as previously described. CD8+ cells were plated at 10^6 cell/ml/well of a 24-well plate in AIM V + 100 Cetus units/ml IL-2 (Chiron, Emeryville, CA) 24 hours prior to transduction. For transduction, the cell volume was reduced to 0.5 ml and 0.75 ml of appropriate supernatant and 0.75 ml of AIM V + 200 Cetus units/ml IL-2 + 4 $\mu\text{g/ml}$ polybrene were added for 4-24 hrs at 38°C . After the transduction cells were returned to growth media for an additional 3-20 days and analyzed for CD4/zeta surface expression as described previously. Table 14 summarizes the results when three independent isolates of CD8 T-cells were transduced with the indicated viral supernatants from transiently transfected tsA54 cells, stably transfected 293 cells (142H.62) or stably transfected 3T3-derived PA317 cells (78.81).

Table 14: High level supernatant transduction of Human CD8+ T cells

Method	retroviral vector	packaging vector	CD8 % transduction (Donor 1)	CD8 %transduction (Donor 2)	CD8 % transduction (Donor 3)
tsA54 Transient	mock	mock	.1	1.8	1.6
tsA54 Transient	pRT43.2F3	pIK6.1MCVampacUTΔ	51	44	46
tsA54 Transient	pRT43.3PGKF3	pIK6.1MCVampacUTΔ	63	53	50
293Stable 142H.62	pRT43.2F3	pIK6.1MCVampacUTΔ	57	53	57
PA317 stable 78.81	pRTD4.2svgF3e		7	9.7	8.8

Table 14 demonstrates that virus-containing supernatants from 293 cells, either stably or transiently transfected according to the methods of the instant invention transduce CD8+T cells at significant higher frequency than supernatants from 3T3 cells.

EXAMPLE XII

High-Level Transduction of Primary Human Cells

This example describes the use of the constructs of the instant invention to efficiently transduce primary human CD34+ bone marrow cells using a protocol involving supernatant infection. To increase the efficiency of viral infection, purified CD34+ cells were placed onto plates which were coated with monoclonal antibodies against the adhesion molecules VLA-4, VLA-5, CD29, CD11a, CD11b, and CD44 prior to infection. This supernatant protocol results in levels of infection which are equivalent to those found with the cocultivation of virus-producing 293 cells. The example also describes

the use of this method for the high level transduction of other primary human cells.

5 The ability to maintain both self-renewing and
differentiating populations of cells derived from stem
cells depends upon cell - cell contact of stem cells and
stromal cells in the bone marrow (Gordon and Greaves,
Bone Marrow Transplantation, 4:335-338 (1989)). The
10 contact of stromal cells and hematopoietic stem cells
involves many molecules including growth factors,
exemplified by the kit ligand on stromal cells and c-kit
receptor found on stem cells (Zsebo et al., Cell, 63:213-
224 (1990)) and adhesion molecules, fibronectin on
15 stromal cells and VLA-4 on hematopoietic stem cells
(Williams et al., Nature 352:438-441 (1991)). These
contact molecules are either transmembrane or, if located
extracellularly, they are proteins which contact
transmembrane proteins and enable signals for either
20 self-renewal or differentiation to be transmitted between
the stromal cells and the stem cells.

 In order to improve the poor retroviral gene
transfer into hematopoietic stem cells by supernatant
infection, recreation of the cell-cell contacts was
25 attempted and resulted in higher efficiency of gene
transfer (Morre et al., Blood 79:1393 (1992)). However,
cocultivation of bone marrow cells on stroma is neither
acceptable by the FDA nor is it economically feasible.
Therefore, attempts at recreation of the cell cell
30 contacts have been undertaken. The interaction of
fibronectin on stromal cells and VLA-4 on hematopoietic
stem cells (Williams et al., supra) has been previously
demonstrated. By isolating the CS-1 domain of
fibronectin responsible for this interaction and coating
35 plates with this protein molecule, Moritz et al.
demonstrated that retroviral gene transfer by supernatant
infection can be significantly enhanced (J. Clin.

Invest., 93:1451-1457 (1994)). This approach necessitates the isolation of significant quantities of proteolytic fragments from natural material. Furthermore, many molecules participate in the cell cell interactions of stroma and stem cells (Liesveld et al. Blood 81:112-121 (1993)).

We have taken the generalizable approach taken of coating cell culture plates with antibodies to adhesion molecules that participate in stromal - hematopoietic stem cell cell-cell contact either singly or in combination, followed by retroviral gene transfer. We have confirmed the observations of Moritz et al., (supra) that the fibronectin / VLA-4 interaction can enhance retroviral transduction and that purified CS-1 fragment can be replaced by anti-VLA4 antibody. We have gone on to show that not only is the recreation of CS1-VLA4 cell-cell contact effective at enhancing retroviral gene transfer but that other cell-cell contacts between stromal cells and hemopoietic cells can be recreated using antibodies to VLA5, CD29, CD11a, CD11b (Liesveld et al. Blood 81:112-121, (1993)) and can also improve retroviral transduction.

Cell-cell contact plays an important role for the activation and growth of many cells of the hemapoietic lineage. For example, many cell-cell contacts have been identified that are essential for T cell activation (Bolhuis et al., Cancer Immunol. Immunother. 34:1-8 (1991)) including the interactions of receptor/coreceptor pairs on T lymphocytes and antigen presenting cells such as LFA-1 and ICAM-1, and CD-2 and LFA-3. In B lymphocytes, the CD40 / gp39 interaction takes place between B lymphocytes and T lymphocytes and is necessary for B lymphocyte activation (Armitage et al., Sem. Immunol., 6:267-278 (1994)). Antibodies to CD2 (Springer et al., Nature 323:262 (1987)) or CD40 can

substitute for the ligands and mediate cell-cell interaction and activation. The transduction of T and B lymphocytes by supernatant infection has been reported to be of low efficiency (Hwu et al., J. Immunol., 9:4104-4115 (1993); Baker et al., Nucleic Acids Res., 20:5234 (1992)). Using an approach similar to that for stem cells, antibodies to the receptor present on the target cells (i.e. anti-CD2 or LFA1 antibody for T lymphocytes and anti-CD40 antibody for B lymphocytes), which have been shown to activate these respective cell types, can also be used to enhance the supernatant transduction efficiency of these cells.

High level transduction of primary human hematopoietic stem cells

CD34+ cells were isolated from the peripheral blood of patients undergoing cyclophosphamide and G-CSF treatment. Mononuclear cells are isolated from leucopheresed blood by fractionation using a standard Ficoll gradient (Pharmacia, Piscataway, NJ). The CD34+ cells are isolated using positive selection on a CellPro CEPRATE LC affinity column (CellPro, Bothell, WA). Post purification analysis via flow cytometry demonstrates that this population is approximately 90% CD34+. This population of cells is then cultured for a period of 48-72 hours at a density of $0.5 - 1 \times 10^6$ cells/ml in "prestimulation medium" which contains Myeloid Long Term Culture Medium supplied as a complete medium from Terry Fox Labs, (Vancouver, Canada) with the addition of 100 ng/ml human Stem Cell Factor (SCF), 50 ng/ml human IL-3, and 10 ng/ml human IL-6 (Genzyme, Cambridge, MA).

Viral supernatant for infection of the CD34+ cells was produced as follows. 293 cells were transfected by first plating at a density of 1.4×10^6 cells/10cm dish 24 hours prior to transfection, followed by co-

transfection with 10 ug pRT43.2F3 vector DNA (encoding CD4/zeta) and 7.5 ug of the packaging plasmid pIK6.1MCVampacUTΔ. Eighteen hours later, transfection media is removed and replaced with 10 mls IMDM (JRH Biosciences, Woodland CA) + 10%FBS. Viral supernatant is then collected 24-36 hours later and 100ng/ml human SCF, 50 ng/ml human IL-3, 10 ng/ml human IL-6, and 8 ug/ml polybrene were added.

To produce antibody-coated plates, 10 ug of antibody or a combination of antibodies (Immunotech, Westbrook ME) is dissolved in 1 ml of PBS and incubated overnight in the tissue culture plates as discussed above. After incubation the plates are washed gently with PBS, and cells and viral supernatant are added immediately. As a comparison, tissue culture plates were also coated with fibronectin or a chymotryptic fragment of fibronectin, CS-1, as reported by Williams et al. (Nature 352: 438-441 (1991)) and Moritz et al. (J. Clin. Invest 93: 1451-1457 (1994)). Fibronectin and CS-1 coated plates are made by adding 30ug/ml PBS of fibronectin, derived from human plasma, or CS-1 (Sigma, St Louis, MO) to tissue culture plates. The plates are then incubated at 37° overnight and washed with PBS ("24 hour method"). Alternatively, the plate is placed under UV light for 1 hour with the lid off and then an additional hour with the lid on, the PBS is removed, one ml of 2% BSA is added for 20 minutes, and the plates are washed with DPBS/0.2% HEPES ("2 hour method") (Williams et al. supra).

As shown below in Table 15 Expt. 1, the use of antibody-coated plates dramatically increased the percentage of hematopoietic stem cells which were transduced by the retroviral supernatants (3.5% without coating compared to from 12.2 to 16.9% with coating with a single antibody). The use of a combination of

antibodies increased the transduction frequency even further (43.2% with anti-VLA-4 and anti-CD44, Expt. 2). Expt. 3 demonstrates that the level of transduction with two antibodies is comparable to that achieved with fibronectin or CS-1 coating (Moritz *et al.*, *supra*). Applicants have also determined that the use of the "24 hour method" of fibronectin coating results in consistently greater transduction frequencies than the "2 hour method").

Table 15. Supernatant transduction of CD4/zeta into CD34+ stem cells

Coating of plates	% CD4+ cells
<u>Expt. 1</u>	
None	3.5
anti-VLA-4	16.2
anti-VLA-5	12.2
anti-CD29	16.6
anti-CD11a + anti-CD11b	15.1
Fibronectin (24 hours)	16.9
<u>Expt. 2</u>	
Fibronectin (24 hours)	31.1
anti-VLA-4 and anti-CD44	43.2
<u>Expt. 3</u>	
Fibronectin (2 hours)	29.6
Fibronectin (24 hours)	53.8
CS-1 (2 hours)	69.5

Table 15 above also demonstrates that the use of fibronectin plates, in combination with the viruses of the instant invention, results in a higher efficiency of transduction of stem cells than that previously reported for other retroviral systems (Moritz et al. supra).

Supernatants from the stable CD4/zeta virus producer cells described above in Example VIII are also efficient transducers of hematopoietic stem cells. In this example, the CD34+ cells are harvested after pre-stimulation, washed, and plated at a density of 7.5×10^5 cells/well in a 6-well tissue culture dish coated with CS-1 as described above (10 ug CS-1). For undiluted supernatants, cells are resuspended in 2 mls of viral supernatant with the addition of cytokines and polybrene, as described above. Viral supernatants were then diluted 1:2, 1:10 and 1:50 in medium containing cytokines and polybrene. Four hours after infection, the cells were collected, washed, and resuspended in viral supernatant for additional exposure to the virus overnight. Fresh "pre-stimulation" media was then added to the cells after washing them free of viral supernatant. The cells were then cultured and analyzed via flow cytometry for CD4 expression.

25

As shown in Table 16 below, viral supernatants from stable producers can also be used to efficiently transduce CD34+ cells. Applicants have also found that the level of transduction of CD34+ cells by the viral supernatants from the various producer clones is correlated with their ability to transduce T cells (Table 10), and not their viral titer as determined by infection of 3T3 cells.

30

35

Table 16. Transduction of hematopoietic stem cells using supernatants from stable producers

Viral Supernatant	Dilution	% transduction of CD34+ cells	Viral titer on 3T3 cells
293 142H.15	1:1	38.1	1.0×10^7
	1:2	35.6	
	1:10	24.8	
	1:50	6.8	
293 142H.62	1:1	62.7	1.1×10^7
	1:2	54.9	
	1:10	29.2	
	1:50	8.2	
293 142.69	1:1	30.4	1.3×10^7
	1:2	26.6	
	1:10	18.1	
	1:50	4.8	

Viral supernatants from stable producer clones can also be used to transduce CD34+ cells incubated on antibody-coated plates. As described above, plates are coated with antibodies to anti-adhesion molecules and the CD34+ cells are purified and added to the plates. Viral supernatants are then added and the percentage of transduced cells is determined.

EXAMPLE XIIIEpisomal Replication of Retroviral Plasmids

5 In another embodiment of the invention, we
obtain high level transient retroviral production using
plasmids containing the Epstein-Barr Virus (EBV) EBNA1
and oriP gene sequences. These sequences have been shown
to direct multi-copy episomal replication of plasmid
10 sequences for many cell generations (Yates et al., Nature
313:812-815 (1985); Margolskee et al., Mol. Cell. Biol.
8:2837-2847 (1988)). Plasmids containing the EBNA1 and
oriP sequences along with the retroviral genome may allow
for maintenance of multiple copies of these retrovirus-
15 containing plasmids in the absence of plasmid
integration. This invention will alleviate the need for
multiple cross infections of retroviral producer cells
with pseudotyped retroviral particles to obtain high
titer stable producer cell lines (Bestwick et al.,
20 supra). The use of this plasmid will also eliminate the
time required to screen multiple clones to isolate high
titer producer clones. These sequences have the
additional benefit of enabling the generation of high
titer producer cell populations containing vectors that
25 have internal promoters and deletions in the enhancer or
enhancer/promoter regions of the 3' LTRs, and therefore
can not be amplified by amplification cocultivation or
serial infection. Due to the enhancer deletion in the 3'
LTR in internal promoter vectors, such constructs without
30 the EBV sequence would need to be transfected into
packaging cell lines followed by screening 50 -100
clones in order to produce high titer retrovirus producer
clones. Insertion into the EBV/oriP replicating vectors
eliminates the need to screen large numbers of clones and
35 enables rapid isolation of producer populations for
internal promoter vectors.

Vectors containing internal promoters are of particular interest for the following reasons. Upon transduction of some primary cells with retroviral vectors in which the transcription of the gene of interest is driven from the viral long terminal repeat (LTR), gene expression is eliminated over time in vivo due to methylation of the viral LTR. One example of this behavior has been observed following transduction of hematopoietic stem cells (Challita and Kohn Proc. Natl. Acad. Sci., USA 91:2567-2571 (1994)). In order to overcome this problem, transcriptional control elements (enhancers, promoters, dominant control elements) can be introduced internal to the vector. These internal promoters, which are resistant to inactivation (Lim et al., Mol. Cell. Biol. 7:3459-3465 (1987); Wilson et al. Proc. Natl. Acad. Sci., USA 87:439-443 (1990); Correll et al., Blood 84:1812-1822 (1994)), include cellular promoters (human or mouse phosphoglycerate kinase, chicken beta actin) as well as viral promoters (SV40 early region, herpes simplex virus thymidine kinase). These vectors can be constructed with either an intact 3' LTR (Correll et al., Blood 84:1812-1822 (1994)) or with a 3' LTR containing an enhancer deletion (for example, Wilson et al. Proc. Natl. Acad. Sci., USA 87:439-443 (1990)). The internal promoters enable expression in all of the differentiated cell types derived from a pluripotent hematopoietic stem cell. Other internal promoters can be used to regulate expression specific for a single cell type. For example, the human beta globin promoter directs specific expression in murine erythrocytes following stem cell gene transfer (Dzierzak et al., Nature 331:35-41(1988)) and the creatine kinase promoter is specific for expression in myoblasts (Dai et al., Proc. Natl. Acad. Sci., USA 89:10892-10895 (1992)). This example describes the construction of retroviral plasmids containing EBNA1 and oriP sequences. pRT43.3PGKF3CEP4ro is a retroviral vector plasmid

containing all necessary elements for high level production of full length packageable retroviral transcripts (a 5' LTR, a psi site, an internal PGK promoter, a 3' LTR with an enhancer deletion and the 3' flanking regions including SV40 poly A site and origin of replication (ori)) on a plasmid backbone containing EBV EBNA1 and oriP sequences. The use of the SV40 origin also enables virus titer to be transiently increased by transfection of plasmids encoding SV40 T antigen, which induces replication via the SV40 origin and increases plasmid and gene expression (Heinzel *et al.* J. Virol., 62(10):3738-3746 (1988)). This vector plasmid was generated in the following manner:

pUC.CEP4 was created to enhance bacterial plasmid production by replacing the 3086 base pair Sall fragment of pCEP4 (Invitrogen city, state) with a 2691 base pair Sall fragment comprised of a 1371 base pair Sal 1 - filled Afl111 fragment from pUC19 (New England Biolabs, Beverly, MA) and a 1316 base pair blunted Bsm1 - Sca1 from pHEBO (Sugden *et al.* Mol. and Cell Biol. 5:410-413 (1985)). (pRT43.3PGK3 contains a deletion of sequences in the 3' LTR which results in the loss of enhancer function. One skilled in the art can produce other 3' LTR sequences lacking enhancer function for use in the instant invention using conventional techniques).

The pRT43.3PGKF3CEP4do vector was created by inserting a 5327 base pair SnaB1 - Avr11 fragment from pRT43.3PGKF3 (described above in the detailed description) into a 9695 base pair SnaB1 - Nhe1 fragment from pUC.CEP4. The pRT43.3PGKF3CEP4ro vector was generated by inserting an 6313 basepair Sal 1-Sal1(partial) fragment from pRT43.3PGKF3CEP4do into an 8672 base pair Sall-Sall backbone fragment from pUC.CEP4. pRT43.3PGK3 contains a deletion of sequences in the 3'LTR

which results in the loss of enhancer function, but still allows virus polyadenylation and transmission. One skilled in the art can produce other 3'LTR sequence lacking enhancer function for use in the instant invention using conventional techniques.

Retroviral supernatants were produced by transient transfection of these pUC.CEP4 based plasmids into tsA54 cells along with the pMCVampacUT Δ packaging plasmid described previously. The titer of these supernatants was determined by infection of 3T3 cells as shown in Table 19.

Table 19

Retroviral vector	3T3 titer (expt. 1)	3T3 titer (expt. 2)
43.3PGKF3	2.4×10^6	ND
43.3PGKF3CEP4do	2.1×10^6	6×10^6
43.3PGKF3CEP4ro	ND	6×10^6

Table 19 shows that the plasmids containing the EBNA1 and oriP sequences along with the retroviral genome (pRT43.3PGKF3CEP4do and pRT43.3PGKF3CEP4ro) produce high titer supernatants comparable to those produced with a vector without the EBV sequences (pRT43.3PGKF3).

The EBNA1 and oriP containing vector plasmids can also be packaged in long-term, stable cell lines, as shown below. To produce these cell lines, the 90.74 amphotropic packaging cells were plated 48 hours prior to transfection and 10 ug of 43.3PGKF3CEP4ro was transfected as previously described and cells resistant to 200 ug/ml hygromycin B were selected. Virus-containing supernatants from independent bulk populations of hygromycin-resistant cells were collected and the viral titers were determined by infecting 3T3 cells. Viral supernatants were also

used to infect CD8+ T cells and the percent of vector-containing cells was determined by analysing the production of the CD4 antigen encoded by the 43.3PGKF3CEP4ro vector. These results are shown below in Table 20.

Table 20: TR157 CEP4 Population Titers

Bulk Population Number	3T3 Titerx10 ⁶
2-10A	0.92
2-10B	0.76
2-20A	1.55
2-20B	1.47
2-50A	0.63
2-50B	1.16
2-100A	0.68
2-100B	1.21
3-10A	1.1
3-10B	1.08
3-20A	0.65
3-20B	1.52

The above table demonstrates that the EBNA1 and oriP containing plasmids can be used to rapidly produce virus which can efficiently infect 3T3 cells and T cells without the need to isolate stable packaging clones.

Virus-producing clones are produced from the bulk population of hygromycin-resistant cells by standard procedures. The clones are screened for production of viral RNA by hybridization analysis using dot blots, and the clones with highest production are selected for

further growth and analysis. After a further 6 weeks of selection in hygromycin, viral supernatants are analyzed for the infection of 3T3 cells and T cells.

5

EXAMPLE XIV

GALV-based vectors and packaging plasmids

Many human cells are not efficiently infected using retroviral vectors and packaging systems based on MMLV. To aid in circumventing this problem, vectors and packaging plasmids can be prepared which are based on other retroviruses, for example the primate retrovirus GALV. Many MMLV sequences (LTRs, psi packaging sites, splice/donor and acceptor sites, and/or primer binding sites) may be substituted using conventional methods in the instant retroviral vectors by the analogous regions from GALV viruses to produce viruses capable of infecting a wide variety of mammalian cells, in particular human cells, when used in the present invention. GALV gag and pol genes can also be used in retroviral packaging plasmids. The production of pseudotyped virions having GALV envelope proteins has been demonstrated. (Wilson et al., J.Virol. 63:2374-2378 (1989)). In addition, Miller et. al., (J. Virol. 65:2220-2224 (1991)), describe construction of hybrid packaging cell lines that express GALV envelope and MMLV gag-pol proteins.

The construction of retroviral packaging plasmids which contain genes encoding GALV gag/pol or envelope proteins is described below.

pIK6.1GALVSEenv contains the gene encoding the GALV envelope protein. It was constructed by replacing the 1980 base pair Bgl11 - Nhe1 amphotropic envelope region of pIK6.1amenvATGUTΔ with the corresponding GALV envelope encoding region from GALV Seato strain (Kawakami, et al., Transplant Proc., 6:193-198 (1974)). A

PCR reaction was performed with synthetic oligonucleotides 5' AATTCGAGATCTGCCGCCATGGTATTGCTGCCTGGGTC 3' (SEQ ID NO. 41) and 5' TGAGGGTCATGGGCTGGTGG 3' (SEQ ID NO. 42) using pGalV-I1 (Eglitis et al., J. Virol., 67:5472-5477 (1993)) as the template. The 180 base pair PCR product was cut with Bgl11 and Afl11 and the resulting 110 base pair fragment was isolated. This fragment was ligated in a four-part ligation with a 1.95 kb Afl11 - BstE11 fragment from pGalV-I1, a 4.2 kb Nhe1 - Bgl11 fragment from pIK6.1amenvATGUTΔ and a DNA fragment composed of synthetic oligonucleotides 5' - GTAACCTTTAAG 3' (SEQ ID NO. 43) and 5' - CTAGCTTAAAG - 3' (SEQ ID NO. 44) to give pIK6.1 GALVSEenv.

pIK6.1MCVGALVgagpol contains the genes encoding the gag and pol proteins of GALV. It is constructed by replacing the 1.98 kb Bgl11 - Nhe MMLV amphotropic envelope encoding fragment of pIK6.1MCVamenvATGUTΔ with the gag/pol sequences from GALV. It is constructed as follows. pGalV-I1 is first digested with Tsp509-1, and then ligated to a DNA fragment composed of synthetic oligonucleotides 5' - GATCTGCCGCCGCCATGGGACAAGAT 3' (SEQ ID NO. 45) and 5' - AATTATCTTGTCCCATGGCGGCGGCA - 3' (SEQ ID NO. 46). This ligation mixture is digested with Rsr11 and the resulting 475 base pair fragment is isolated. pGalV-I1 is also digested with Afl11 and then ligated to a DNA fragment composed of synthetic oligonucleotides 5' TTAAGCTGCGTATTCGGCGGCGGCGGGACGAGTCTGCAAAATAAG 3' (SEQ ID NO. 47) and 5' CTAGCTTATTTTGCAGACTCGTCCCGCCGCCGCGCAATACGCAGC 3' (SEQ ID NO. 48). This ligation mixture is digested with Rsr11 and the resulting 4.59 kb fragment is isolated. These 475 base pair and 4.59kb fragments, described above, are then ligated in a three - part ligation with a 4.2 kb Nhe1 - Bgl11 fragment from pIK6.1MCVgagpolATG to produce pIK6.1MCVGaLVgagpol.

5 All publications and patent applications cited in this specification are herein incorporated by reference in their entirety as if each individual publication or patent application were specifically and individually indicated to be incorporated by reference.

10 As will be apparent to those skilled in the art to which the invention pertains, the present invention may be embodied in forms other than those specifically disclosed above, for example to transfect and transduce other mammalian cell types, without departing from the spirit or essential characteristics of the invention. The particular embodiments of the invention described above, are, therefore, to be considered as illustrative and not restrictive. The scope of the present invention is as set forth in the appended claims rather than being limited to the examples contained in the foregoing description.

20 SEQUENCE LISTING

(1) GENERAL INFORMATION:

25 (i) APPLICANT: FINER, MITCHELL H.
DULL, THOMAS J.
ZSEBO, KRISZTINA M.
COOKE, KEEGAN
FARSON, DEBORAH A.

30 (ii) TITLE OF INVENTION: METHOD FOR PRODUCTION OF HIGH TITER VIRUS AND HIGH EFFICIENCY RETROVIRAL MEDIATED TRANSDUCTION OF MAMMALIAN CELLS

(iii) NUMBER OF SEQUENCES: 48

35 (iv) CORRESPONDENCE ADDRESS:

(A) ADDRESSEE: CELL GENESYS, INC.

(B) STREET: 322 LAKESIDE DRIVE
(C) CITY: FOSTER CITY
(D) STATE: CALIFORNIA
(E) COUNTRY: USA
(F) ZIP: 94404

5

(v) COMPUTER READABLE FORM:

(A) MEDIUM TYPE: Floppy disk
(B) COMPUTER: IBM PC compatible
(C) OPERATING SYSTEM: PC-DOS/MS-DOS
(D) SOFTWARE: PatentIn Release #1.0, Version

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(viii) ATTORNEY/AGENT INFORMATION:

(A) NAME: KRUPEN, KAREN I.
(B) REGISTRATION NUMBER: 34,647
(C) REFERENCE/DOCKET NUMBER: CELL 13.3

30

(ix) TELECOMMUNICATION INFORMATION:

(A) TELEPHONE: 415-358-9600 X131
(B) TELEFAX: 415-349-7392

35

(2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:

- 5 (A) LENGTH: 33 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

10

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

15 GGTCGACCTG GATCCGCCAT ACCACATTTG TAG
33

(2) INFORMATION FOR SEQ ID NO:2:

20 (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 33 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

25

(ii) MOLECULE TYPE: DNA (genomic)

30 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

GCCGCGGCTC TAGAGCCAGA CATGATAAGA TAC
33

35

(2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:

- 5 (A) LENGTH: 33 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

10

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

15 AAGCTTGTGC TAGCTATCCC GCCCCTAACT CCG
33

(2) INFORMATION FOR SEQ ID NO:4:

20 (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 33 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

25

(ii) MOLECULE TYPE: DNA (genomic)

30 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

CGAAATCGGT CGACCGCAAA AGCCTAGGCC TCC
33

35

(2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:

- 5 (A) LENGTH: 30 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

10

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

15 GTCTATAGCA TGCTCCCCTG CTCCGACCCG
30

(2) INFORMATION FOR SEQ ID NO:6:

20 (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 30 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

25

(ii) MOLECULE TYPE: DNA (genomic)

30 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

GGTACCGAAT TCTCCTGCGG GGAGAAGCAG
30

35

(2) INFORMATION FOR SEQ ID NO:7:

(i) SEQUENCE CHARACTERISTICS:

- 5 (A) LENGTH: 26 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

10

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

15 CGCCAAGCTT GGCCATTGCA TACGGT

26

(2) INFORMATION FOR SEQ ID NO:8:

20 (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 30 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

25

(ii) MOLECULE TYPE: DNA (genomic)

30 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

GAGGTCTAGA CGGTTCACTA AACGAGCTCT

30

35

(2) INFORMATION FOR SEQ ID NO:9:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 13 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

Gly Ser Thr Ser Gly Ser Gly Ser Ser Glu Gly Lys Gly
1 5 10

(2) INFORMATION FOR SEQ ID NO:10:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 27 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

CGGAGATCTC GTGCGACCGC GAGAGCC

27

(2) INFORMATION FOR SEQ ID NO:11:

(i) SEQUENCE CHARACTERISTICS:

- 5 (A) LENGTH: 38 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

10 (ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

15 GGAATTCGCT AGCTTTCAG GAGCGCAAAT GTTGTGTC
38

(2) INFORMATION FOR SEQ ID NO:12:

20 (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 27 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

25 (ii) MOLECULE TYPE: DNA (genomic)

30 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

CGGAGATCTC RCGCGACCCC GAGAGCC
27

35

(2) INFORMATION FOR SEQ ID NO:13:

(i) SEQUENCE CHARACTERISTICS:

- 5 (A) LENGTH: 24 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

10

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

15 CGGGATCCAG AGCTGCAACT GGAG
24

(2) INFORMATION FOR SEQ ID NO:14:

20 (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 26 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

25

(ii) MOLECULE TYPE: DNA (genomic)

30 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

GAAGATCTGA CCTTGAAGAA GGTGAC
26

35

(2) INFORMATION FOR SEQ ID NO:15:

(i) SEQUENCE CHARACTERISTICS:

- 5 (A) LENGTH: 36 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

10

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

15 TCTCCTCCAG TTGCAGCTCC GGAGACAGGG AGAGGC
36

(2) INFORMATION FOR SEQ ID NO:16:

20 (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 16 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

25

(ii) MOLECULE TYPE: DNA (genomic)

30 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

TTGCAGCTCC GGAGAC
16

35

(2) INFORMATION FOR SEQ ID NO:17:

(i) SEQUENCE CHARACTERISTICS:

- 5 (A) LENGTH: 36 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

10

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

15 CAGCACAATC AGGGCCATGT CCAGCTCCCC GTCCTG
36

(2) INFORMATION FOR SEQ ID NO:18:

20 (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 16 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

25

(ii) MOLECULE TYPE: DNA (genomic)

30 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

AGGGCCATGT CCAGCT
16

35

(2) INFORMATION FOR SEQ ID NO:19:

(i) SEQUENCE CHARACTERISTICS:

- 5 (A) LENGTH: 28 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

10

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:

15 CGGAATTCGG TACCTCCTGT GCAAGAAC
28

(2) INFORMATION FOR SEQ ID NO:20:

(i) SEQUENCE CHARACTERISTICS:

- 20 (A) LENGTH: 26 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

25

(ii) MOLECULE TYPE: DNA (genomic)

30 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:

CGGAATTCGC CTCCACCAAG GGCCCA
26

35

(2) INFORMATION FOR SEQ ID NO:21:

(i) SEQUENCE CHARACTERISTICS:

- 5 (A) LENGTH: 31 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

10

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:

15 CGGAATTCAC GCGTCCCACT CAGGACACAG C
31

(2) INFORMATION FOR SEQ ID NO:22:

20 (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 35 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

25

(ii) MOLECULE TYPE: DNA (genomic)

30 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:

GAGAGAGATC TGCTAGCGGT CAGGCTGGAA CTGAG
35

35

(2) INFORMATION FOR SEQ ID NO:23:

(i) SEQUENCE CHARACTERISTICS:

- 5 (A) LENGTH: 36 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

10

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:

15 GCATGTGTGA GTTTTGTCTG AGGAGACGGT GACCAG
36

(2) INFORMATION FOR SEQ ID NO:24:

(i) SEQUENCE CHARACTERISTICS:

- 20 (A) LENGTH: 16 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

25

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:

30 GTTTTGTCTG AGGAGA
16

35

(2) INFORMATION FOR SEQ ID NO:25:

(i) SEQUENCE CHARACTERISTICS:

- 5 (A) LENGTH: 32 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

10

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:

15 GTGACAGTCG ACCCCTTGAA GTCCACTTTG GT
32

(2) INFORMATION FOR SEQ ID NO:26:

20 (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 21 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

25

(ii) MOLECULE TYPE: DNA (genomic)

30 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:

CCACCCCTCA CTCTGCTTCT C
21

35

(2) INFORMATION FOR SEQ ID NO:27:

(i) SEQUENCE CHARACTERISTICS:

- 5 (A) LENGTH: 43 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

10

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:

15 TCGACCAGCG GCAGCGGCAA GAGCAGCGAG GGTAAGGGTA CCA
43

(2) INFORMATION FOR SEQ ID NO:28:

(i) SEQUENCE CHARACTERISTICS:

- 20 (A) LENGTH: 43 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

25

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:

30 GATCTGGTAC CCTTACCCTC GCTGCTCTTG CCGCTGCCGC TGG
43

35

(2) INFORMATION FOR SEQ ID NO:29:

(i) SEQUENCE CHARACTERISTICS:

- 5 (A) LENGTH: 36 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

10

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:29:

15 CTCCTGTAGT AGCACCTGAC CCTTACCCTC GCTGCT
36

(2) INFORMATION FOR SEQ ID NO:30:

20 (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 16 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

25

(ii) MOLECULE TYPE: DNA (genomic)

30 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:30:

AGCACCTGAC CCTTAC
16

35

(2) INFORMATION FOR SEQ ID NO:31:

(i) SEQUENCE CHARACTERISTICS:

- 5 (A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

10

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:31:

15 CTGATCTTAC TCTTTGGACC
20

(2) INFORMATION FOR SEQ ID NO:32:

(i) SEQUENCE CHARACTERISTICS:

- 20 (A) LENGTH: 32 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

25

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:32:

30 GAATTCGCTA GCCTATGGCT CGTACTCTAT AG
32

35

(2) INFORMATION FOR SEQ ID NO:33:

(i) SEQUENCE CHARACTERISTICS:

- 5 (A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

10

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:33:

15 GACCACACTG GCGTAGTAAG
20

(2) INFORMATION FOR SEQ ID NO:34:

20 (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 33 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

25

(ii) MOLECULE TYPE: DNA (genomic)

30 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:34:

GAATTCGCTA GCTTATTCAC GCGATTCTAC TTC
33

35

(2) INFORMATION FOR SEQ ID NO:35:

(i) SEQUENCE CHARACTERISTICS:

- 5 (A) LENGTH: 27 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

10

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:35:

15 GAATTCATG GAAGGTTTCAG CGTTCTC
27

(2) INFORMATION FOR SEQ ID NO:36:

20 (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

25

(ii) MOLECULE TYPE: DNA (genomic)

30 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:36:

CGTTAGCTGT TTGTCCTGTC
20

35

(2) INFORMATION FOR SEQ ID NO:37:

(i) SEQUENCE CHARACTERISTICS:

- 5 (A) LENGTH: 30 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

10

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:37:

15 GAATTCAAGC TTAATGTAGT CTTATGCAAT
30

(2) INFORMATION FOR SEQ ID NO:38:

20 (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 30 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

25

(ii) MOLECULE TYPE: DNA (genomic)

30 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:38:

GAATTCTCTA GAGTTTATTG TATCGAGCTA
30

35

(2) INFORMATION FOR SEQ ID NO:39:

(i) SEQUENCE CHARACTERISTICS:

- 5 (A) LENGTH: 44 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

10

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:39:

15 CGATAACAATA AACGCGCCAG TCCTCCGATT GACTGAGTCC CCGG
44

(2) INFORMATION FOR SEQ ID NO:40:

20 (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 47 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

25

(ii) MOLECULE TYPE: DNA (genomic)

30 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:40:

GTACCCGGGC GACTCAGTCA ATCGGAGGAC TGGCGCGTTT ATTGTAT
47

35

(2) INFORMATION FOR SEQ ID NO:41:

(i) SEQUENCE CHARACTERISTICS:

- 5 (A) LENGTH: 38 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

10 (ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:41:

15 AATTCGAGAT CTGCCGCCAT GGTATTGCTG CCTGGGTC
38

(2) INFORMATION FOR SEQ ID NO:42:

- 20 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

25 (ii) MOLECULE TYPE: DNA (genomic)

30 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:42:

TGAGGGTCAT GGGCTGGTGG
20

35

(2) INFORMATION FOR SEQ ID NO:43:

(i) SEQUENCE CHARACTERISTICS:

- 5 (A) LENGTH: 12 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

10 (ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:43:

15 GTAACCTTTA AG
12

(2) INFORMATION FOR SEQ ID NO:44:

20 (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 11 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

25 (ii) MOLECULE TYPE: DNA (genomic)

30 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:44:

CTAGCTTAAA G
11

35

(2) INFORMATION FOR SEQ ID NO:45:

(i) SEQUENCE CHARACTERISTICS:

- 5 (A) LENGTH: 26 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

10

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:45:

15 GATCTGCCGC CGCCATGGGA CAAGAT
26

(2) INFORMATION FOR SEQ ID NO:46:

20 (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 26 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

25

(ii) MOLECULE TYPE: DNA (genomic)

30 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:46:

AATTATCTTG TCCCATGGCG GCGGCA
26

35

(2) INFORMATION FOR SEQ ID NO:47:

(i) SEQUENCE CHARACTERISTICS:

- 5 (A) LENGTH: 45 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

10 (ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:47:

15 TTAAGCTGCG TATTCGGCGG CGGCGGGACG AGTCTGCAAA ATAAG
45

(2) INFORMATION FOR SEQ ID NO:48:

20 (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 45 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

25 (ii) MOLECULE TYPE: DNA (genomic)

30 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:48:

CTAGCTTATT TTGCAGACTC GTCCCGCCGC CGCCGAATAC GCAGC
45

35

International Application No: PCT/ /

MICROORGANISMS	
Optional Sheet in connection with the microorganism referred to on page __, lines __ of the description *	
A. IDENTIFICATION OF DEPOSIT * Further deposits are identified on an additional sheet *	
Name of depositary institution * American Type Culture Collection	
Address of depositary institution (including postal code and country) * 12301 Parklawn Drive Rockville, MD 20852 US	
Date of deposit * <u>August 22, 1995</u> Accession Number * <u>CRL 11971</u>	
B. ADDITIONAL INDICATIONS * (leave blank if not applicable). This information is continued on a separate attached sheet	
C. DESIGNATED STATES FOR WHICH INDICATIONS ARE MADE * (if the indications are not all designated States)	
D. SEPARATE FURNISHING OF INDICATIONS * (leave blank if not applicable)	
The indications listed below will be submitted to the International Bureau later * (Specify the general nature of the indications e.g., "Accession Number of Deposit")	
E. <input checked="" type="checkbox"/> This sheet was received with the International application when filed (to be checked by the receiving Office)	
<div style="text-align: right;"><u>Mindy Walker RO/LIS</u> (Authorized Officer)</div>	
<input type="checkbox"/> The date of receipt (from the applicant) by the International Bureau *	
was _____ (Authorized Officer)	

Form PCT/RO/134 (January 1981)

International Application No: PCT/ /

Form PCT/RO/134 (cont.)

American Type Culture Collection

12301 Parklawn Drive
Rockville, MD 20852
US

<u>Accession No.</u>	<u>Date of Deposit</u>
CRL 11972	August 25, 1995
CRL 11654	June 10, 1994
75483	June 11, 1993
75484	June 11, 1993
75485	June 11, 1993
75486	June 11, 1993
CRL 1658	
CRL 2017	
CRL 2032	

Claims

1. A retroviral packaging plasmid for the production of high titers of recombinant retrovirus in human cells comprising one retroviral helper DNA sequence derived from a replication-incompetent retroviral genome encoding in trans all virion proteins required for packaging a replication-incompetent retroviral vector and for producing virion proteins capable of packaging said replication-incompetent retroviral vector at high titer, without the production of replication-competent helper virus, said retroviral DNA sequence lacking the region encoding the native enhancer and/or promoter of the viral 5' LTR of said virus and lacking both the psi function sequence responsible for packaging helper genome and the 3'LTR, and encoding a foreign enhancer and/or promoter functional in a selected mammalian cell, and a foreign polyadenylation site,

wherein said helper DNA sequence codes for ecotropic MMLV or GALV gag and pol and an envelope protein selected from a xenotropic, amphotropic, ecotropic or polytropic envelope protein.

2. A retroviral packaging plasmid for the production of high titers of recombinant retrovirus in human cells comprising one retroviral helper DNA sequence derived from a replication-incompetent retroviral genome encoding in trans all virion proteins required for packaging a replication-incompetent retroviral vector and for producing virion proteins capable of packaging said replication-incompetent retroviral vector at high titer, without the production of replication-competent helper virus, said retroviral DNA sequence lacking the region encoding the native enhancer and/or promoter of the viral 5' LTR of said virus and lacking the both the psi function sequence responsible for packaging helper genome and the 3'LTR, and encoding a foreign enhancer and/or promoter functional in

a selected mammalian cell, and a foreign polyadenylation site,

and wherein said helper DNA sequence codes for ecotropic MMLV or GALV gag and pol proteins, and envelope protein from the group consisting of 10A1 murine leukemia virus envelope protein, gibbon ape leukemia virus envelope protein, HIV envelope protein, vesicular stomatitis virus G protein, human T cell leukemia type I envelope protein and human T cell leukemia type II envelope protein.

3. A retroviral packaging plasmid for the production of high titers of recombinant retrovirus in human cells comprising two retroviral helper DNA sequences derived from a replication-incompetent retroviral genome encoding in trans all virion proteins required for packaging a replication-incompetent retroviral vector and for producing virion proteins capable of packaging said replication-incompetent retroviral vector at high titer, without the production of replication-competent helper virus, said retroviral DNA sequences lacking the region encoding the native enhancer and/or promoter of the viral 5' LTR of said virus and lacking both the psi function sequence responsible for packaging helper genome and the 3'LTR, and encoding a foreign enhancer and/or promoter functional in a selected mammalian cell, and a foreign polyadenylation site,

wherein a first retroviral helper sequence comprises a cDNA sequence encoding the gag and pol proteins of ecotropic MMLV or GALV and a second retroviral helper sequence comprises a cDNA encoding the envelope protein, and

wherein said second retroviral helper DNA sequence codes for an envelope protein selected from a xenotropic, amphotropic, ecotropic or polytropic envelope protein.

4. A retroviral packaging plasmid for the production of high titers of recombinant retrovirus in human cells

comprising two retroviral helper DNA sequences derived from a replication-incompetent retroviral genome encoding in trans all virion proteins required for packaging a replication-incompetent retroviral vector and for producing
5 virion proteins capable of packaging said replication-incompetent retroviral vector at high titer, without the production of replication-competent helper virus, said retroviral DNA sequence lacking the region encoding the native enhancer and/or promoter of the viral 5' LTR of said
10 virus and lacking both the psi function sequence responsible for packaging helper genome and the 3'LTR, and encoding a foreign enhancer and/or promoter functional in a selected mammalian cell, and a foreign polyadenylation site,

15 wherein a first retroviral helper sequence comprises a cDNA sequence encoding the gag and pol proteins of ectropic MMLV or GALV and a second retroviral helper sequence comprises a cDNA encoding the envelope protein, and

20 wherein said second retroviral helper DNA sequence codes for an envelope protein from the group consisting of 10A1 murine leukemia virus envelope protein, gibbon ape leukemia virus envelope protein, HIV envelope protein, vesicular stomatitis virus G protein, human T cell leukemia
25 type I envelope protein and human T cell leukemia type II envelope protein.

5. The stable packaging cell line comprising helper sequences encoding gag and pol proteins designated 35.32,
30 ATCC Designation CRL 11972.

6. A human embryonic kidney cell stably transfected with an expression vector encoding the gag and pol protein and a selectable marker wherein the expression of gag and pol
35 proteins are stable in the absence of a selective agent.

7. The human embryonic kidney cell of claim 6 wherein the gag and pol proteins are derived from MMLV or GALV
8. The human embryonic kidney cell of claim 6 wherein
5 said cell is either 293 or tsa54.
9. A human embryonic kidney cell stably transfected with two expression vectors wherein the first expression vector encodes the gag and pol proteins and the second expression
10 vector encodes the envelope protein.
10. The human embryonic kidney cell of claim 9 wherein
15 said cell is either 293 or tsa54 and said gag and pol proteins are derived from MMLV or GALV.
11. The human embryonic kidney cell of claim 9 or 10 wherein said envelope protein is derived from amphotropic, xenotropic, ecotropic and polytropic envelope proteins.
12. The human embryonic kidney cell of claim 9 or 10
20 wherein said envelope protein is derived from 10A1 murine leukemia virus envelope protein, gibbon ape leukemia envelope protein, HIV envelope protein, vesicular stomatitis virus G protein, human T cell leukemia type I
25 envelope protein and human T cell leukemia type II envelope protein.
13. A stable packaging cell line comprising helper sequences encoding gag, pol and envelope proteins
30 designated 37S2.8, ATCC Designation CRL 11971.
14. A retroviral packaging plasmid for the production of high titers of recombinant retrovirus in human cells comprising one retroviral helper DNA sequence derived from
35 a replication-incompetent retroviral genome encoding in trans all virion proteins required for packaging a replication-incompetent retroviral vector and for producing

virion proteins capable of packaging said replication-incompetent retroviral vector at high titer, without the production of replication-competent helper virus, said retroviral DNA sequence lacking the region encoding the native enhancer and/or promoter of the viral 5' LTR of said virus and lacking the both the psi function sequence responsible for packaging helper genome and the 3'LTR, and encoding a foreign enhancer and/or promoter functional in a selected mammalian cell, and a foreign polyadenylation site,

wherein said foreign enhancer is the RSV enhancer and promoter.

15. A retroviral packaging plasmid for the production of high titers of recombinant retrovirus in human cells comprising two retroviral helper DNA sequences derived from a replication-incompetent retroviral genome encoding in trans all virion proteins required for packaging a replication-incompetent retroviral vector and for producing virion proteins capable of packaging said replication-incompetent retroviral vector at high titer, without the production of replication-competent helper virus, said retroviral DNA sequences lacking the region encoding the native enhancer and/or promoter of the viral 5' LTR of said virus and lacking both the psi function sequence responsible for packaging helper genome and the 3'LTR, and encoding a foreign enhancer and/or promoter functional in a selected mammalian cell, and a foreign polyadenylation site,

wherein a first retroviral helper sequence comprises a cDNA sequence encoding the gag and pol proteins of ectropic MMLV or GALV and a second retroviral helper sequence comprises a cDNA encoding the envelope protein,

wherein said second retroviral helper DNA sequence codes for an envelope protein selected from a xenotropic, amphotropic, ecotropic or polytropic envelope protein; and

wherein said foreign enhancer is the RSV enhancer and promoter.

5 16. A retroviral packaging plasmid for the production of
high titers of recombinant retrovirus in human cells
comprising two retroviral helper DNA sequences derived from
a replication-incompetent retroviral genome encoding in
trans all virion proteins required for packaging a
10 replication-incompetent retroviral vector and for producing
virion proteins capable of packaging said replication-
incompetent retroviral vector at high titer, without the
production of replication-competent helper virus, said
retroviral DNA sequence lacking the region encoding the
15 native enhancer and/or promoter of the viral 5' LTR of said
virus and lacking both the psi function sequence
responsible for packaging helper genome and the 3'LTR, and
encoding a foreign enhancer and/or promoter functional in
a selected mammalian cell, and a foreign polyadenylation
site,

20 wherein a first retroviral helper sequence comprises
a cDNA sequence encoding the gag and pol proteins of
ectropic MMLV or GALV and a second retroviral helper
sequence comprises a cDNA encoding the envelope protein,

25 wherein said second retroviral helper DNA sequence
codes for an envelope protein from the group consisting of
10A1 murine leukemia virus envelope protein, gibbon ape
leukemia virus envelope protein, HIV envelope protein,
vesicular stomatitis virus G protein, human T cell leukemia
type I envelope protein and human T cell leukemia type II
30 envelope protein, and

wherein said foreign enhancer is the RSV enhancer and
promoter.

35 17. A high efficiency method to transduce mammalian
hematopoietic stem cells with retroviral supernatants
produced by transient transfection comprising the steps
of

A) transient cotransfection of a first population of mammalian cells that can produce virus with:

5 (i) one retroviral helper DNA sequence derived from a replication-incompetent retroviral genome encoding in trans all virion proteins required for packaging a replication-incompetent retroviral vector and for producing virion proteins capable of packaging said replication-incompetent retroviral vector at high titer, without the production of replication-competent helper virus, said
10 retroviral DNA sequence lacking the region encoding the native enhancer and/or promoter of the viral 5' LTR of said virus and lacking both the psi function sequence responsible for packaging helper genome and the 3'LTR, and encoding a foreign enhancer and/or promoter functional in
15 a selected mammalian cell, and a foreign polyadenylation site; and

(ii) a retroviral vector encoding a foreign gene to produce replication-defective recombinant retroviral vectors carrying said foreign gene in said first population
20 of mammalian cells;

B) separation of said first population of mammalian cells from cell supernatant;

C) adding adhesion molecules or antibodies to adhesion molecules to culture plates;

25 D) growing a second population of mammalian hematopoietic stem cells on said culture plates; and

E) incubating said supernatant containing replication-defective recombinant retroviral vectors carrying said foreign gene with said second population of
30 mammalian hematopoietic stem cells, to transduce said second population of cells with said foreign gene,

whereby target cells efficiently transduced with said foreign gene are obtained.

35 18. A high efficiency method to transduce mammalian hematopoietic stem cells with retroviral supernatants produced by transient transfection comprising the steps of

A) transient cotransfection of a first population of mammalian cells that can produce virus with:

5 (i) two retroviral helper DNA sequences derived from a replication-incompetent retroviral genome encoding in trans all virion proteins required for packaging a replication-incompetent retroviral vector and for producing virion proteins capable of packaging said replication-incompetent retroviral vector at high titer, without the production of replication-competent helper virus, said
10 retroviral DNA sequences lacking the region encoding the native enhancer and/or promoter of the viral 5' LTR of said virus and lacking both the psi function sequence responsible for packaging helper genome and the 3' LTR, and encoding a foreign enhancer and/or promoter functional in
15 a selected mammalian cell, and a foreign polyadenylation site,

wherein a first retroviral helper sequence comprises a cDNA sequence encoding the gag and pol proteins of ectropic MMLV or GALV and a second retroviral helper
20 sequence comprises a cDNA encoding the envelope protein, and

wherein said second retroviral helper DNA sequence codes for an envelope protein; and

25 (ii) a retroviral vector encoding a foreign gene to produce replication-defective recombinant retroviral vectors carrying said foreign gene in said first population of mammalian cells;

30 B) separation of said first population of mammalian cells from cell supernatant;

C) adding adhesion molecules or antibodies to adhesion molecules to culture plates;

D) growing a second population of mammalian hematopoietic stem cells on said culture plates; and

35 E) incubating said supernatant containing replication-defective recombinant retroviral vectors carrying said foreign gene with said second population of

mammalian hematopoietic stem cells, to transduce said second population of cells with said foreign gene,

whereby target cells efficiently transduced with said foreign gene are obtained.

5

19. A high efficiency method to transduce mammalian hematopoietic stem cells with retroviral supernatants produced by transient transfection comprising the steps of:

10

A) transient cotransfection of a first population of mammalian cells stably transfected with an expression vector encoding the gag and pol proteins and a selectable marker wherein the expression of the gag and pol proteins are stable in the absence of a selective agent with:

15

(i) one retroviral helper DNA sequence derived from a replication-incompetent retroviral genome, said retroviral DNA sequence lacking the region encoding the native enhancer and/or promoter of the viral 5' LTR of said virus and lacking both the psi function sequence responsible for packaging helper genome and the 3'LTR, and encoding a foreign enhancer and/or promoter functional in a selected mammalian cell, and a foreign polyadenylation site, and encoding an envelope protein; and

20

25

(ii) a retroviral vector encoding a foreign gene to produce replication-defective recombinant retroviral vectors carrying said foreign gene in said first population of mammalian cells;

30

B) separation of said first population of mammalian cells from cell supernatant;

C) adding adhesion molecules or antibodies to adhesion molecules to culture plates;

D) growing a second population of mammalian hematopoietic stem cells on said culture plates; and

35

E) incubating said supernatant containing replication-defective recombinant retroviral vectors carrying said foreign gene with said second population of

mammalian hematopoietic stem cells, to transduce said second population of cells with said foreign gene,

whereby target cells efficiently transduced with said foreign gene are obtained.

5

20. A high efficiency method to transduce mammalian hematopoietic stem cells with retroviral supernatants produced by transient transfection comprising the steps of:

10

A) transient transfection of a first population of mammalian cells stably transfected with at least one expression vector encoding the gag, pol and env proteins and a selectable marker wherein the expression of the gag, pol and env proteins are stable in the absence of a selective agent with a retroviral vector encoding a foreign gene to produce replication-defective recombinant retroviral vectors carrying said foreign gene in said first population of mammalian cells;

15

B) separation of said first population of mammalian cells from cell supernatant;

20

C) adding adhesion molecules or antibodies to adhesion molecules to culture plates;

D) growing a second population of mammalian hematopoietic stem cells on said culture plates; and

25

E) incubating said supernatant containing replication-defective recombinant retroviral vectors carrying said foreign gene with said second population of mammalian hematopoietic stem cells, to transduce said second population of cells with said foreign gene,

30

whereby target cells efficiently transduced with said foreign gene are obtained.

35

21. A high efficiency method to transduce mammalian hematopoietic stem cells with retroviral supernatants produced by stable mammalian viral producer cells comprising the steps of:

- A) separation of said first population of stable mammalian viral producer cells from cell supernatant;
- B) adding adhesion molecules or antibodies to adhesion molecules to culture plates;
- 5 C) growing a second population of mammalian hematopoietic stem cells on said culture plates; and
- D) incubating said supernatant containing replication-defective recombinant retroviral vectors carrying said foreign gene with said second population of
- 10 mammalian hematopoietic stem cells, to transduce said second population of cells with said foreign gene,
- whereby target cells efficiently transduced with said foreign gene are obtained.
- 15 22. The method of claims 17 - 21 in the alternative wherein said adhesion molecules is selected from the group consisting of fibronectin and CS-1.
- 20 23. The method of claims 17 - 21 in the alternative wherein said antibodies to adhesion molecules is selected from the group consisting of antibodies to VLA-4, VLA-5, CD29, CD11a, CD11b and CD44.
- 25 24. A retroviral vector plasmid for maintenance as a multicopy episome in human cells comprising a selectable marker and a retroviral genome encoding a foreign gene and the Epstein-Barr Virus EBNA1 and oriP sequences.
- 30 25. The retroviral vector plasmid of claim 24 wherein the retroviral genome comprises in the 5' to 3' direction a 5' LTR, a splice donor site, a psi site, a splice acceptor site, a foreign gene and a 3' LTR.
- 35 26. The retroviral vector plasmid of claim 24 wherein the retroviral genome comprises in the 5' to 3' direction a 5' LTR, a psi site, a foreign promoter, a foreign gene and a 3' LTR.

27. The retroviral vector plasmid of claim 24 wherein the retroviral genome comprises in the 5' to 3' direction a 5' LTR, a psi site, a foreign promoter, a foreign gene and a 3' LTR having a deletion in the enhancer region, wherein
5 enhancer activity is eliminated.

28. A method for the production of human stable retroviral producer cells comprising the steps of:

transfecting a human cell stably transfected with at
10 least one expression vector encoding the gag, pol and envelope proteins with the vector plasmid of claims 24 - 27 in the alternative, selecting with a selectable agent and growing the cells in the presence of said selectable agent.

29. A high efficiency method to transduce mammalian T and B lymphocytes with retroviral supernatants produced by transient transfection comprising the steps of :

A) transient cotransfection of a first population of mammalian cells that can produce virus with:

(i) one retroviral helper DNA sequence derived from
20 a replication-incompetent retroviral genome encoding in trans all virion proteins required for packaging a replication-incompetent retroviral vector and for producing virion proteins capable of packaging said replication-incompetent retroviral vector at high titer, without the
25 production of replication-competent helper virus, said retroviral DNA sequence lacking the region encoding the native enhancer and/or promoter of the viral 5' LTR of said virus and lacking both the psi function sequence
30 responsible for packaging helper genome and the 3' LTR, and encoding a foreign enhancer and/or promoter functional in a selected mammalian cell, and a foreign polyadenylation site; and

(ii) a retroviral vector encoding a foreign gene to
35 produce replication-defective recombinant retroviral vectors carrying said foreign gene in said first population of mammalian cells;

B) separation of said first population of mammalian cells from cell supernatant;

C) adding antibodies to adhesion molecules to culture plates;

5 D) growing a second population of mammalian T or B lymphocytes on said culture plates; and

E) incubating said supernatant containing replication-defective recombinant retroviral vectors carrying said foreign gene with said second population of mammalian T or B lymphocytes, to transduce said second
10 population of cells with said foreign gene,

whereby target cells efficiently transduced with said foreign gene are obtained.

15 30. A high efficiency method to transduce mammalian T or B lymphocytes with retroviral supernatants produced by transient transfection comprising the steps of :

A) transient cotransfection of a first population of mammalian cells that can produce virus with:

20 (i) two retroviral helper DNA sequences derived from a replication-incompetent retroviral genome encoding in trans all virion proteins required for packaging a replication-incompetent retroviral vector and for producing virion proteins capable of packaging said replication-
25 incompetent retroviral vector at high titer, without the production of replication-competent helper virus, said retroviral DNA sequences lacking the region encoding the native enhancer and/or promoter of the viral 5' LTR of said virus and lacking both the psi function sequence
30 responsible for packaging helper genome and the 3' LTR, and encoding a foreign enhancer and/or promoter functional in a selected mammalian cell, and a foreign polyadenylation site,

wherein a first retroviral helper sequence comprises
35 a cDNA sequence encoding the gag and pol proteins of ectropic MMLV or GALV and a second retroviral helper

sequence comprises a cDNA encoding the envelope protein,
and

wherein said second retroviral helper DNA sequence
codes for an envelope protein; and

5 (ii) a retroviral vector encoding a foreign gene to
produce replication-defective recombinant retroviral
vectors carrying said foreign gene in said first population
of mammalian cells;

10 B) separation of said first population of mammalian
cells from cell supernatant;

C) adding antibodies to adhesion molecules to culture
plates;

D) growing a second population of mammalian T or B
lymphocytes on said culture plates; and

15 E) incubating said supernatant containing
replication-defective recombinant retroviral vectors
carrying said foreign gene with said second population of
mammalian T or B lymphocytes, to transduce said second
population of cells with said foreign gene,

20 whereby target cells efficiently transduced with said
foreign gene are obtained.

31. A high efficiency method to transduce mammalian T or
B lymphocytes with retroviral supernatants produced by
25 transient transfection comprising the steps of :

A) transient cotransfection of a first population of
mammalian cells stably transfected with an expression
vector encoding the gag and pol proteins and a selectable
marker wherein the expression of the gag and pol proteins
30 are stable in the absence of a selective agent with:

(i) one retroviral helper DNA sequence derived from
a replication-incompetent retroviral genome, said
retroviral DNA sequence lacking the region encoding the
native enhancer and/or promoter of the viral 5' LTR of said
35 virus and lacking both the psi function sequence
responsible for packaging helper genome and the 3'LTR, and
encoding a foreign enhancer and/or promoter functional in

a selected mammalian cell, and a foreign polyadenylation site, and encoding an envelope protein; and

(ii) a retroviral vector encoding a foreign gene to produce replication-defective recombinant retroviral vectors carrying said foreign gene in said first population of mammalian cells;

B) separation of said first population of mammalian cells from cell supernatant;

C) adding antibodies to adhesion molecules to culture plates;

D) growing a second population of mammalian T or B lymphocytes on said culture plates; and

E) incubating said supernatant containing replication-defective recombinant retroviral vectors carrying said foreign gene with said second population of mammalian T or B lymphocytes, to transduce said second population of cells with said foreign gene,

whereby target cells efficiently transduced with said foreign gene are obtained.

32. A high efficiency method to transduce mammalian T or B lymphocytes with retroviral supernatants produced by transient transfection comprising the steps of :

A) transient transfection of a first population of mammalian cells stably transfected with at least one expression vector encoding the gag, pol and env proteins and a selectable marker wherein the expression of the gag, pol and env proteins are stable in the absence of a selective agent with a retroviral vector encoding a foreign gene to produce replication-defective recombinant retroviral vectors carrying said foreign gene in said first population of mammalian cells;

B) separation of said first population of mammalian cells from cell supernatant;

C) adding antibodies to adhesion molecules to culture plates;

D) growing a second population of mammalian T or B lymphocytes on said culture plates; and

E) incubating said supernatant containing replication-defective recombinant retroviral vectors carrying said foreign gene with said second population of mammalian T or B lymphocytes, to transduce said second population of cells with said foreign gene,

whereby target cells efficiently transduced with said foreign gene are obtained.

33. A high efficiency method to transduce mammalian T or B lymphocytes with retroviral supernatants produced by stable mammalian viral producer cells comprising the steps of:

A) separation of said first population of stable mammalian viral producer cells from cell supernatant;

B) adding antibodies to adhesion molecules to culture plates;

C) growing a second population of mammalian T or B lymphocytes on said culture plates; and

D) incubating said supernatant containing replication-defective recombinant retroviral vectors carrying said foreign gene with said second population of mammalian T or B lymphocytes, to transduce said second population of cells with said foreign gene,

whereby target cells efficiently transduced with said foreign gene are obtained.

34. The method of claims 29 - 33 in the alternative wherein said antibodies to adhesion molecules is selected from the group consisting of antibodies to LFA-1, CD-2, CD40 and gp39.

35. A improved method to efficiently transduce mammalian cells with a retroviral supernatant, comprising the steps of:

i) growing said population of mammalian cells on culture plates; and

ii) incubating said supernatant containing replication-defective recombinant retroviral vectors carrying a foreign gene with said population of mammalian cells, to transduce said population of mammalian cells with said foreign gene,

whereby target cells efficiently transduced with said foreign gene are obtained, wherein the improvement comprises adding antibodies to adhesion molecules present on said population of mammalian cells to culture plates.

36. Mammalian target cells transduced with a foreign gene by the method of claim 1.

37. The target cells of claim 36, wherein said cells are human cells.

38. The target cells of claim 37, wherein said cells are selecting from the group consisting of lymphocytes, hematopoietic stem cells, fibroblasts, epithelial cells, endothelial cells, myoblasts, retinal epithelial cells, islets of Langerhans, adrenal medulla cells, osteoblasts, osteoclasts, neurons, glial cells, ganglion cells, embryonic stem cells, and hepatocytes.

39. The method of claim 1, wherein said foreign gene is selected from the group consisting of growth factors, lymphokines, hormones and coagulation factors.

40. The method of claim 1, wherein said foreign gene encodes a chimeric T cell receptor.

41. The method of claim 40, wherein said chimeric T cell receptor is a receptor encoded by a DNA sequence comprising in reading frame:

a sequence encoding a signal sequence;

a sequence encoding a non-MHC restricted extracellular surface membrane protein domain binding specifically to at least one ligand;

- 5 a sequence encoding a transmembrane domain; and
a signal sequence encoding a cytoplasmic signal-transducing domain of a protein that activates an intracellular messenger system.

10 42. Mammalian target cells transduced with a foreign gene by the method of claim 17.

43. The target cells of claim 42, wherein said cells are human cells.

15 44. The target cells of claim 43, wherein said cells are selecting from the group consisting of lymphocytes, hematopoietic stem cells, fibroblasts, epithelial cells, endothelial cells, myoblasts, retinal epithelial cells, islets of Langerhans, adrenal medulla cells, osteoblasts,
20 osteoclasts, neurons, glial cells, ganglion cells, embryonic stem cells, and hepatocytes.

45. The method of claim 17, wherein said foreign gene is selected from the group consisting of growth factors,
25 lymphokines, hormones and coagulation factors.

46. The method of claim 17, wherein said foreign gene encodes a chimeric T cell receptor.

30 47. The method of claim 46, wherein said chimeric T cell receptor is a receptor encoded by a DNA sequence comprising in reading frame:

- a sequence encoding a signal sequence;
a sequence encoding a non-MHC restricted
35 extracellular surface membrane protein domain binding specifically to at least one ligand;
a sequence encoding a transmembrane domain; and

a signal sequence encoding a cytoplasmic signal-transducing domain of a protein that activates an intracellular messenger system.

5 48. Mammalian target cells transduced with a foreign gene by the method of claim 18.

49. The target cells of claim 48, wherein said cells are human cells.

10

50. The target cells of claim 49, wherein said cells are selecting from the group consisting of lymphocytes, hematopoietic stem cells, fibroblasts, epithelial cells, endothelial cells, myoblasts, retinal epithelial cells, 15 islets of Langerhans, adrenal medulla cells, osteoblasts, osteoclasts, neurons, glial cells, ganglion cells, embryonic stem cells, and hepatocytes.

20

51. The method of claim 18, wherein said foreign gene is selected from the group consisting of growth factors, lymphokines, hormones and coagulation factors.

25

52. The method of claim 18, wherein said foreign gene encodes a chimeric T cell receptor.

53. The method of claim 52, wherein said chimeric T cell receptor is a receptor encoded by a DNA sequence comprising in reading frame:

30

a sequence encoding a signal sequence;
a sequence encoding a non-MHC restricted extracellular surface membrane protein domain binding specifically to at least one ligand;

35

a sequence encoding a transmembrane domain; and
a signal sequence encoding a cytoplasmic signal-transducing domain of a protein that activates an intracellular messenger system.

54. Mammalian target cells transduced with a foreign gene by the method of claim 19.

5 55. The target cells of claim 54, wherein said cells are human cells.

56. The target cells of claim 55, wherein said cells are selecting from the group consisting of lymphocytes, hematopoietic stem cells, fibroblasts, epithelial cells, 10 endothelial cells, myoblasts, retinal epithelial cells, islets of Langerhans, adrenal medulla cells, osteoblasts, osteoclasts, neurons, glial cells, ganglion cells, embryonic stem cells, and hepatocytes.

15 57. The method of claim 19, wherein said foreign gene is selected from the group consisting of growth factors, lymphokines, hormones and coagulation factors.

20 58. The method of claim 19, wherein said foreign gene encodes a chimeric T cell receptor.

59. The method of claim 58, wherein said chimeric T cell receptor is a receptor encoded by a DNA sequence comprising in reading frame:

25 a sequence encoding a signal sequence;
a sequence encoding a non-MHC restricted extracellular surface membrane protein domain binding specifically to at least one ligand;
a sequence encoding a transmembrane domain; and
30 a signal sequence encoding a cytoplasmic signal-transducing domain of a protein that activates an intracellular messenger system.

35 60. Mammalian target cells transduced with a foreign gene by the method of claim 20.

61. The target cells of claim 60, wherein said cells are human cells.

62. The target cells of claim 61, wherein said cells are selecting from the group consisting of of lymphocytes, hematopoietic stem cells, fibroblasts, epithelial cells, endothelial cells, myoblasts, retinal epithelial cells, islets of Langerhans, adrenal medulla cells, osteoblasts, osteoclasts, neurons, glial cells, ganglion cells, embryonic stem cells, and hepatocytes.

63. The method of claim 20, wherein said foreign gene is selected from the group consisting of growth factors, lymphokines, hormones and coagulation factors.

64. The method of claim 20, wherein said foreign gene encodes a chimeric T cell receptor.

65. The method of claim 64, wherein said chimeric T cell receptor is a receptor encoded by a DNA sequence comprising in reading frame:

a sequence encoding a signal sequence;
a sequence encoding a non-MHC restricted extracellular surface membrane protein domain binding specifically to at least one ligand;
a sequence encoding a transmembrane domain; and
a signal sequence encoding a cytoplasmic signal-transducing domain of a protein that activates an intracellular messenger system.

66. Mammalian target cells transduced with a foreign gene by the method of claim 21.

67. The target cells of claim 66, wherein said cells are human cells.

5 68. The target cells of claim 67, wherein said cells are selecting from the group consisting of of lymphocytes, hematopoietic stem cells, fibroblasts, epithelial cells, endothelial cells, myoblasts, retinal epithelial cells, islets of Langerhans, adrenal medulla cells, osteoblasts, osteoclasts, neurons, glial cells, ganglion cells, embryonic stem cells, and hepatocytes.

10 69. The method of claim 21, wherein said foreign gene is selected from the group consisting of growth factors, lymphokines, hormones and coagulation factors.

15 70. The method of claim 21, wherein said foreign gene encodes a chimeric T cell receptor.

71. The method of claim 70, wherein said chimeric T cell receptor is a receptor encoded by a DNA sequence comprising in reading frame:

20 a sequence encoding a signal sequence;
a sequence encoding a non-MHC restricted extracellular surface membrane protein domain binding specifically to at least one ligand;
a sequence encoding a transmembrane domain; and
25 a signal sequence encoding a cytoplasmic signal-transducing domain of a protein that activates an intracellular messenger system.

30 72. Mammalian target cells transduced with a foreign gene by the method of claim 29.

73. The target cells of claim 72, wherein said cells are human cells.

35 74. The target cells of claim 73, wherein said cells are selecting from the group consisting of lymphocytes, hematopoietic stem cells, fibroblasts, epithelial cells, endothelial cells, myoblasts, retinal epithelial cells,

islets of Langerhans, adrenal medulla cells, osteoblasts, osteoclasts, neurons, glial cells, ganglion cells, embryonic stem cells, and hepatocytes.

5 75. The method of claim 29, wherein said foreign gene is selected from the group consisting of growth factors, lymphokines, hormones and coagulation factors.

10 76. The method of claim 29, wherein said foreign gene encodes a chimeric T cell receptor.

77. The method of claim 76, wherein said chimeric T cell receptor is a receptor encoded by a DNA sequence comprising in reading frame:

15 a sequence encoding a signal sequence;
 a sequence encoding a non-MHC restricted extracellular surface membrane protein domain binding specifically to at least one ligand;
 a sequence encoding a transmembrane domain; and
20 a signal sequence encoding a cytoplasmic signal-transducing domain of a protein that activates an intracellular messenger system.

25 78. Mammalian target cells transduced with a foreign gene by the method of claim 30.

79. The target cells of claim 78, wherein said cells are human cells.

30 80. The target cells of claim 79, wherein said cells are selecting from the group consisting of lymphocytes, hematopoietic stem cells, fibroblasts, epithelial cells, endothelial cells, myoblasts, retinal epithelial cells, islets of Langerhans, adrenal medulla cells, osteoblasts,
35 osteoclasts, neurons, glial cells, ganglion cells, embryonic stem cells, and hepatocytes.

81. The method of claim 30, wherein said foreign gene is selected from the group consisting of growth factors, lymphokines, hormones and coagulation factors.

5 82. The method of claim 30, wherein said foreign gene encodes a chimeric T cell receptor.

83. The method of claim 82, wherein said chimeric T cell receptor is a receptor encoded by a DNA sequence comprising
10 in reading frame:

a sequence encoding a signal sequence;

a sequence encoding a non-MHC restricted extracellular surface membrane protein domain binding specifically to at least one ligand;

15 a sequence encoding a transmembrane domain; and
a signal sequence encoding a cytoplasmic signal-transducing domain of a protein that activates an intracellular messenger system.

20 84. Mammalian target cells transduced with a foreign gene by the method of claim 31.

85. The target cells of claim 84, wherein said cells are human cells.

25

86. The target cells of claim 85, wherein said cells are selecting from the group consisting of lymphocytes, hematopoietic stem cells, fibroblasts, epithelial cells, endothelial cells, myoblasts, retinal epithelial cells,
30 islets of Langerhans, adrenal medulla cells, osteoblasts, osteoclasts, neurons, glial cells, ganglion cells, embryonic stem cells, and hepatocytes.

87. The method of claim 31, wherein said foreign gene is
35 selected from the group consisting of growth factors, lymphokines, hormones and coagulation factors.

88. The method of claim 31, wherein said foreign gene encodes a chimeric T cell receptor.

89. The method of claim 88, wherein said chimeric T cell receptor is a receptor encoded by a DNA sequence comprising in reading frame:

a sequence encoding a signal sequence;
a sequence encoding a non-MHC restricted extracellular surface membrane protein domain binding specifically to at least one ligand;
a sequence encoding a transmembrane domain; and
a signal sequence encoding a cytoplasmic signal-transducing domain of a protein that activates an intracellular messenger system.

90. Mammalian target cells transduced with a foreign gene by the method of claim 32.

91. The target cells of claim 90, wherein said cells are human cells.

92. The target cells of claim 91, wherein said cells are selecting from the group consisting of lymphocytes, hematopoietic stem cells, fibroblasts, epithelial cells, endothelial cells, myoblasts, retinal epithelial cells, islets of Langerhans, adrenal medulla cells, osteoblasts, osteoclasts, neurons, glial cells, ganglion cells, embryonic stem cells, and hepatocytes.

93. The method of claim 32, wherein said foreign gene is selected from the group consisting of growth factors, lymphokines, hormones and coagulation factors.

94. The method of claim 32, wherein said foreign gene encodes a chimeric T cell receptor.

95. The method of claim 94, wherein said chimeric T cell receptor is a receptor encoded by a DNA sequence comprising in reading frame:

a sequence encoding a signal sequence;

5 a sequence encoding a non-MHC restricted extracellular surface membrane protein domain binding specifically to at least one ligand;

a sequence encoding a transmembrane domain; and

10 a signal sequence encoding a cytoplasmic signal-transducing domain of a protein that activates an intracellular messenger system.

96. Mammalian target cells transduced with a foreign gene by the method of claim 33.

15

97. The target cells of claim 96, wherein said cells are human cells.

98. The target cells of claim 97, wherein said cells are
20 selecting from the group consisting of lymphocytes, hematopoietic stem cells, fibroblasts, epithelial cells, endothelial cells, myoblasts, retinal epithelial cells, islets of Langerhans, adrenal medulla cells, osteoblasts, osteoclasts, neurons, glial cells, ganglion cells,
25 embryonic stem cells, and hepatocytes.

99. The method of claim 33, wherein said foreign gene is selected from the group consisting of growth factors, lymphokines, hormones and coagulation factors.

30

100. The method of claim 33, wherein said foreign gene encodes a chimeric T cell receptor.

101. The method of claim 100, wherein said chimeric T cell
35 receptor is a receptor encoded by a DNA sequence comprising in reading frame:

a sequence encoding a signal sequence;

a sequence encoding a non-MHC restricted extracellular surface membrane protein domain binding specifically to at least one ligand;

a sequence encoding a transmembrane domain; and

5 a signal sequence encoding a cytoplasmic signal-transducing domain of a protein that activates an intracellular messenger system.

10 102. A mammalian cell producing recombinant retrovirus produced by the method of claim 29.

103. The mammalian cell of claim 102, wherein the mammalian cell is a human cell.

15 104. The mammalian cell of claim 103, wherein said human cell is a 293 cell.

20 105. The method of claim 29, further comprising infecting a second population of mammalian target cells with the supernatant from said mammalian cells of claim 29 to transduce said target cells with a foreign gene.

25 106. The method of claim 105, wherein said target cells are lymphocytes or hematopoietic stem cells.

30

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35

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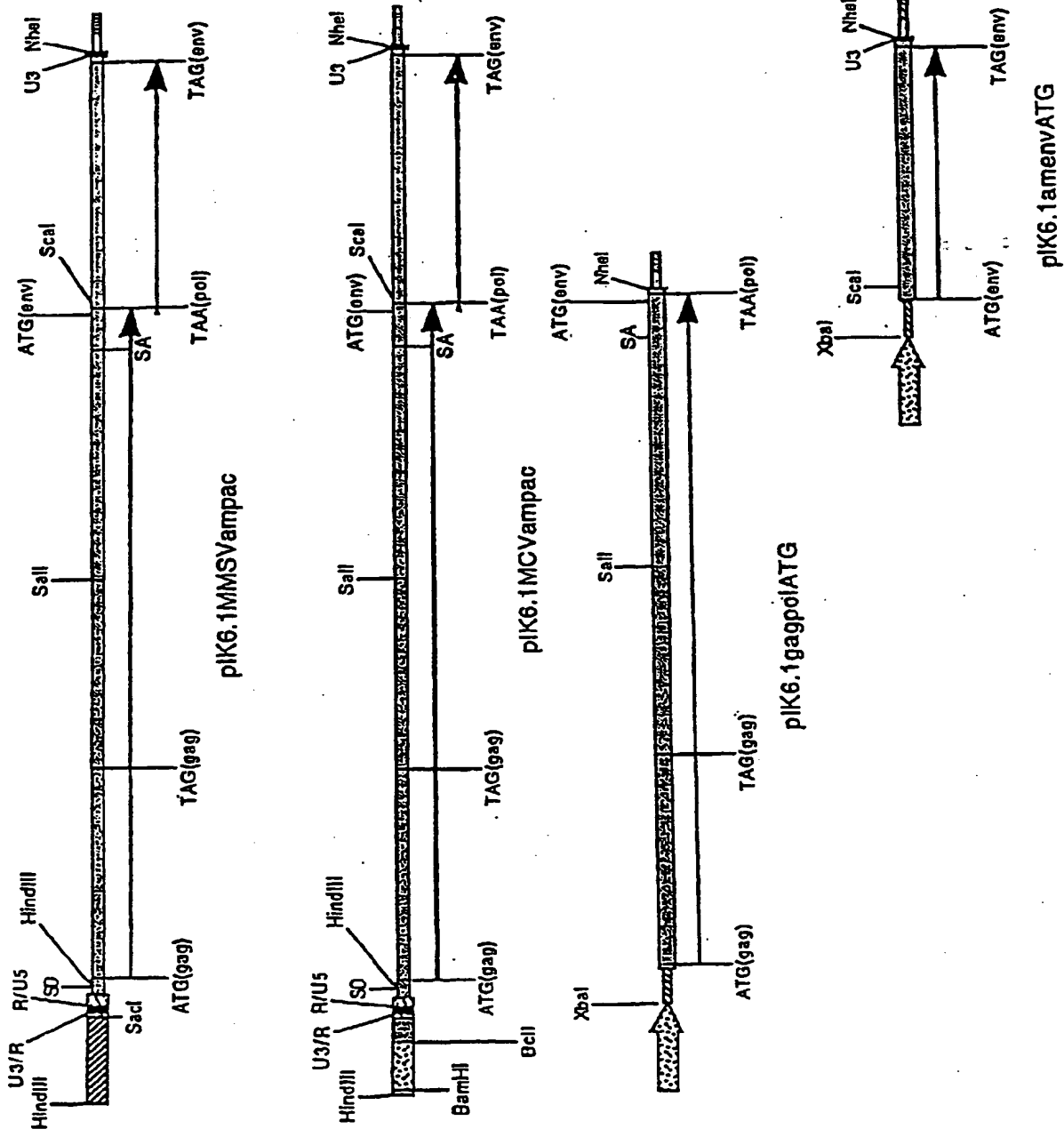


FIGURE 1

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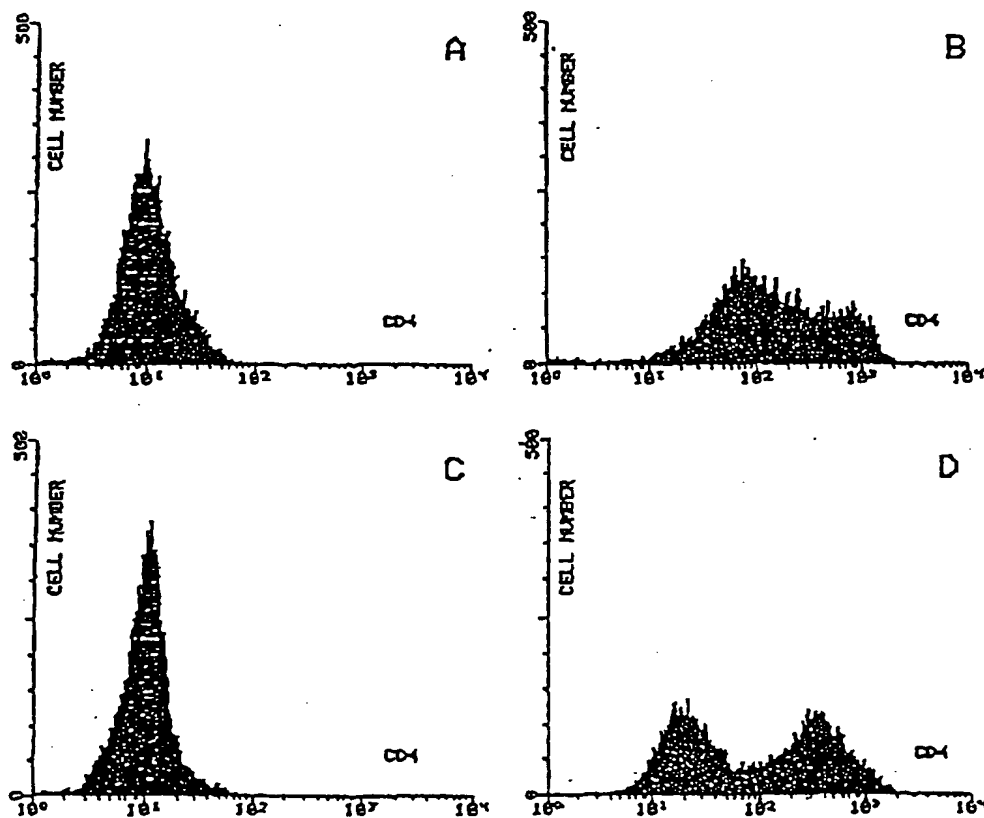


FIGURE 2

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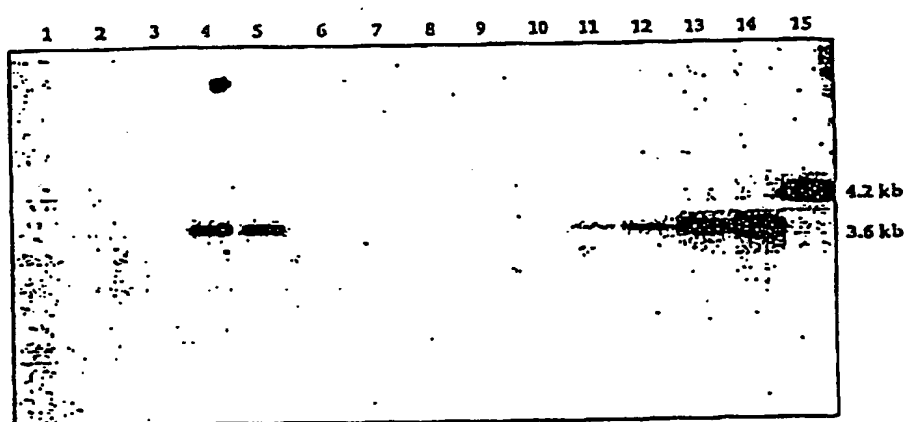


FIGURE 3A

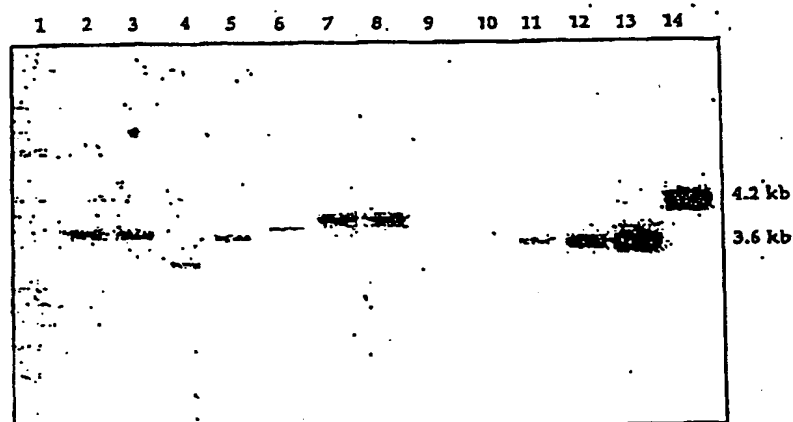


FIGURE 3 B

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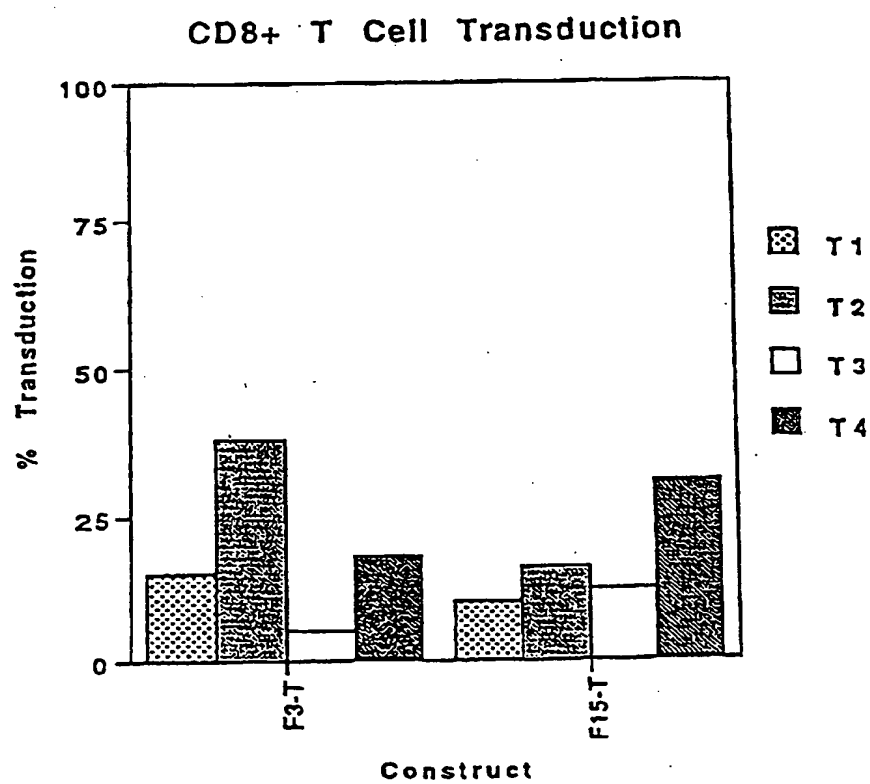


FIGURE 4

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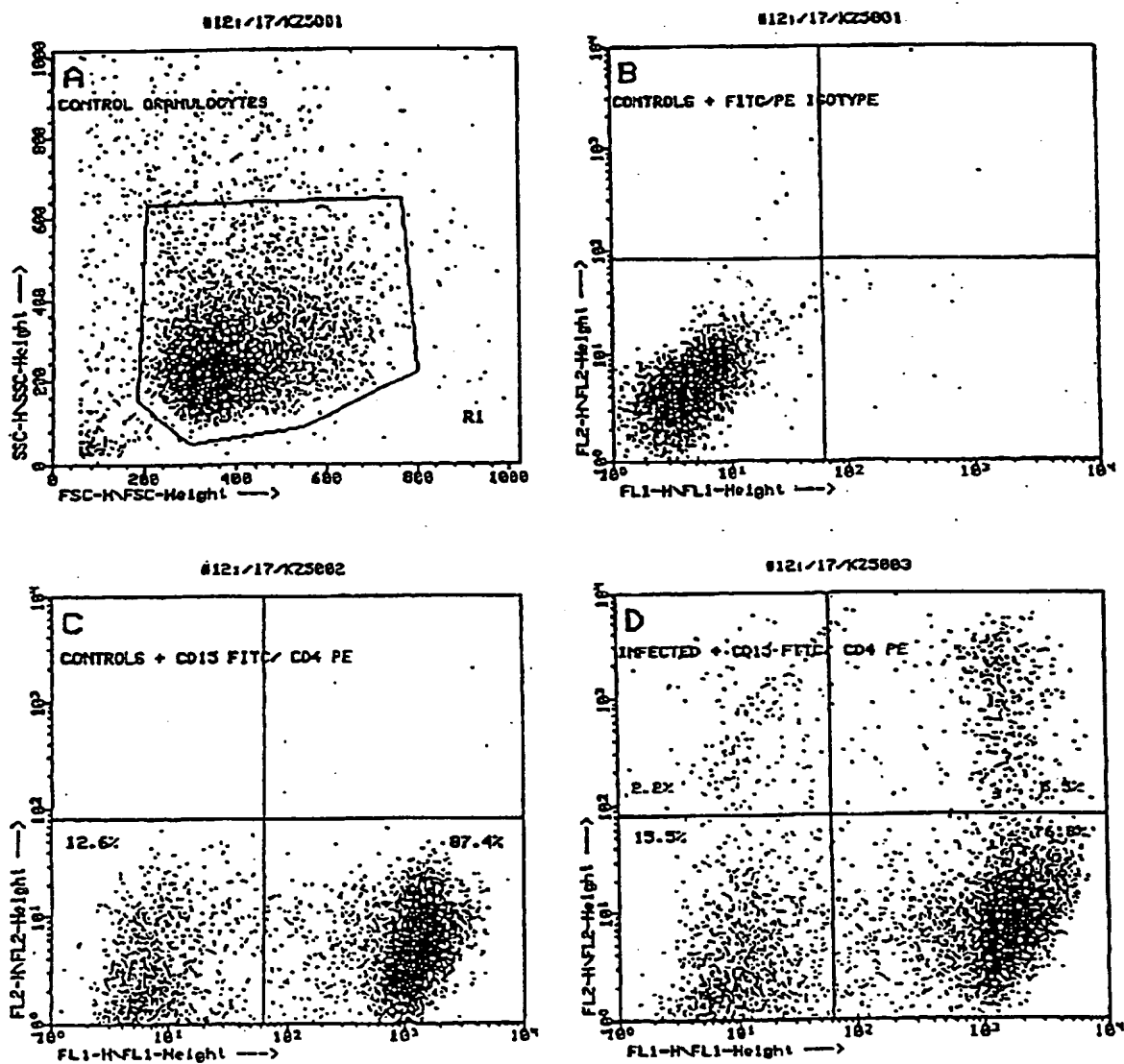


FIGURE 5

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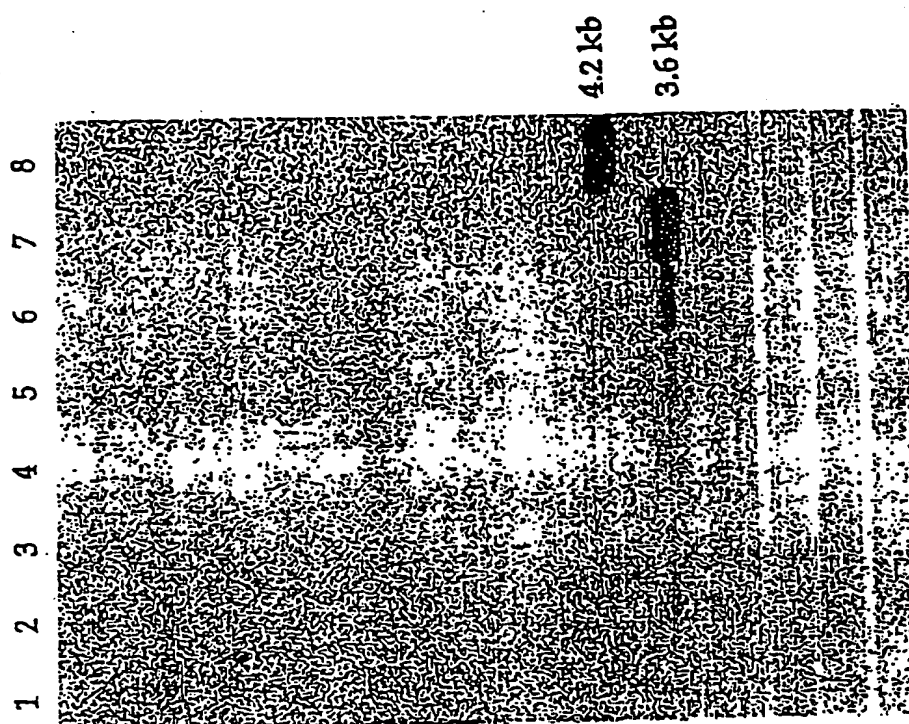


FIGURE 6

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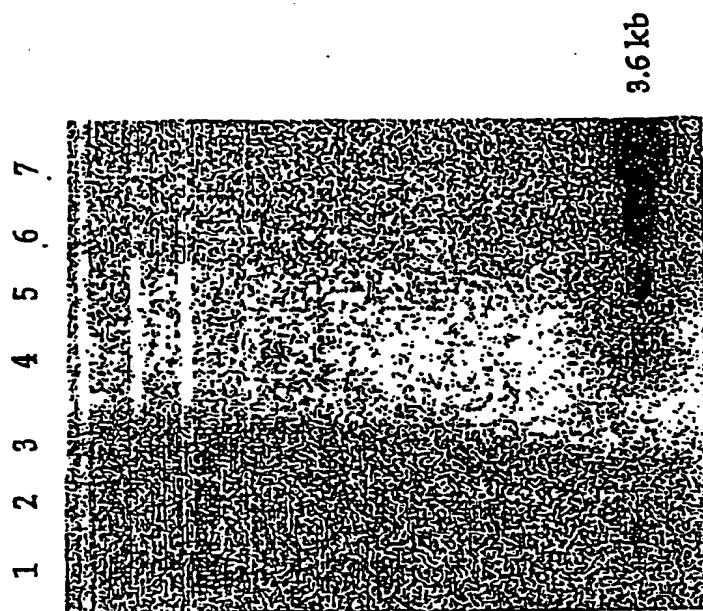


FIGURE 7

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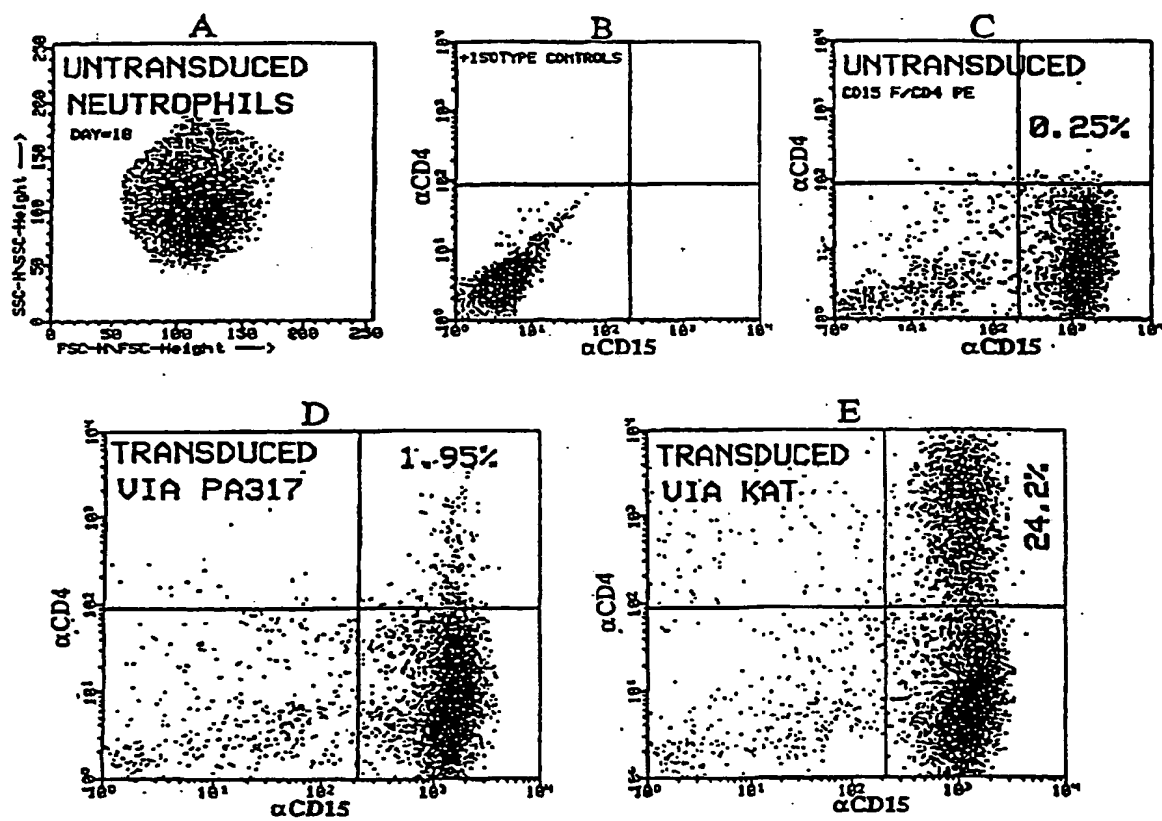


FIGURE 8

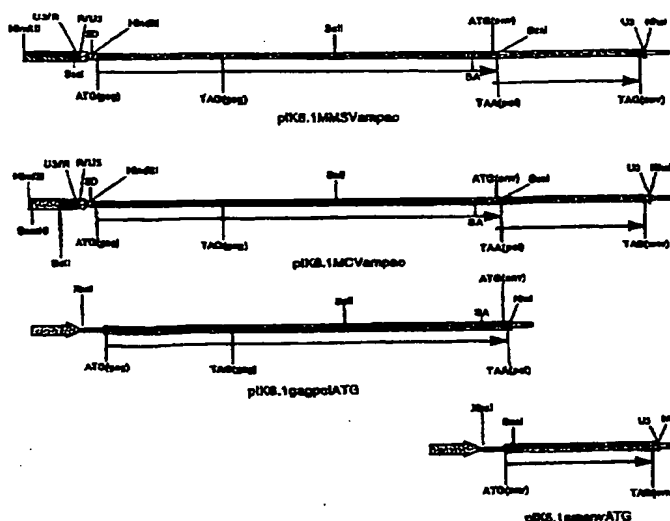
11



INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

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(71) Applicant: CELL GENESYS, INC. [US/US]; 322 Lakeside Drive, Foster City, CA 94404 (US).			
(72) Inventors: FINER, Mitchell, H.; 54 Madera, San Carlos, CA 94070 (US). DULL, Thomas, J.; 1850 Great Highway, San Francisco, CA 94122 (US). ZSEBO, Krisztina, M.; 200 Allen Road, Woodside, CA 94062 (US). COOKE, Keegan; 430 Olive Avenue, Palo Alto, CA 94306 (US). FARSON, Deborah, A.; 521 62nd Street, Oakland, CA 94609 (US).			
(74) Agents: HALLUIN, Albert, P. et al.; Pennic & Edmonds, 1155 Avenue of the Americas, New York, NY 10036 (US).		(88) Date of publication of the international search report: 26 June 1997 (26.06.97)	

(54) Title: HIGH EFFICIENCY RETROVIRAL PACKAGING SYSTEM



(57) Abstract

The invention provides a novel retroviral packaging system, in which retroviral packaging plasmids and packageable vector transcripts are produced from high expression plasmids after stable or transient transfection in mammalian cells. High titers of recombinant retrovirus are produced in these transfected mammalian cells and can then transduce a mammalian target cell by cocultivation or supernatant infection. The methods of the invention include the use of the novel retroviral packaging plasmids and vectors to transduce primary human cells, including T cells and human hematopoietic stem cells, with foreign genes by cocultivation or supernatant infection at high efficiencies. The invention is useful for the rapid production of high titer viral supernatants, and to transduce with high efficiency cells that are refractory to transduction by conventional means.

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INTERNATIONAL SEARCH REPORT

International Application No
PC1/US 96/13737

A. CLASSIFICATION OF SUBJECT MATTER

IPC 6 C12N15/86 C12N5/10 C07K14/15 C12N15/87 C07K14/73
C07K14/725 C07K16/10

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 C12N C07K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 94 29438 A (CELL GENESYS INC) 22 December 1994	1-5
Y	see the whole document	17-23, 29-31

X	BLOOD, vol. 83, no. 1, 1 January 1994, pages 43-50, XP000614890 FINER, M.H. ET AL.: "kat: a high efficient retroviral transduction system for human T lymphocytes"	1-5
Y	see the whole document	22,23, 29-31

	-/-	

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

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Date of the actual completion of the international search

14 February 1997

Date of mailing of the international search report

16. 05. 97

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CHAM BONNET, F

INTERNATIONAL SEARCH REPORT

International Application No.

PC1/US 96/13737

C(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	JOURNAL OF CLINICAL INVESTIGATION, vol. 93, no. 4, April 1994, pages 1451-1457, XP000617796 MORITZ, T. ET AL.: "Bone marrow extracellular matrix molecules improve gene transfer into human hematopoietic cells via retroviral vectors" cited in the application see the whole document ---	17-23, 29-31
A	WO 93 07281 A (NEDERLANDSE ORG TOEGEPAST) 15 April 1993 see claims ---	1,17,18
A	BLOOD, vol. 81, no. 1, 1 January 1993, pages 112-121, XP000617795 LIESVELD, J.L. ET AL.: "Expression of integrins and examination of their adhesive function in normal and leukemic hematopoietic cells" cited in the application see the whole document -----	17,18, 23,29-31

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US 96/ 13737

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:
2. ☒ Claims Nos.: 36-41
because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
- see continuation-sheet PCT/ISA/210
3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

5 inventions * see continuation-sheet PCT/ISA/210 *

1. ☐ As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
4. ☒ No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

1-5, 14-18 and, partially, 17-19, 22, 23, 29-31, 42-59, 72-89, 102-106

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
- ☐ No protest accompanied the payment of additional search fees.

INTERNATIONAL SEARCH REPORT

International Application No. PCT/US 96/ 13737

FURTHER INFORMATION CONTINUED FROM PCT/ISA/210

1. claims 1-5, 14-18 and, partially, 17-19, 22, 23, 29-31, 42-59, 72-89, 102-106:

A retroviral packaging system for the production of high titers of recombinant retrovirus in human cells comprising at least one retroviral helper DNA sequence derived from a replication-incompetent retroviral genome encoding in trans all virion proteins required for packaging a replication-incompetent retroviral vector and for producing virion proteins capable of packaging said replication-incompetent retroviral vector at high titer, without the production of replication-competent helper virus, said retroviral DNA sequence lacking the region encoding the native enhancer and/or promoter of the viral 5' LTR of said virus and lacking both the psi function sequence responsible for packaging helper genome and the 3'LTR, and encoding a foreign enhancer and/or promoter functional in a selected mammalian cell, and a foreign polyadenylation site, wherein said helper DNA sequences codes for ecotropic MMLV or GALV gag and pol and an envelope protein selected from a xenotropic, amphotropic, ecotropic or polytropic envelope protein; plasmids encoding these sequences, cells transduced with them and methods using these.

2. claims 6-8 and, partially, 19, 20, 22, 23, 32, 34, 57-65, 90-95

A human embryonic kidney cell stably transfected with an expression vector encoding the gag and pol protein and a selectable marker wherein the expression of gag and pol proteins are stable in the absence of a selective agent.

3. claims 9-13:

A human embryonic kidney cell stably transfected with two expression vectors wherein the first expression vector encodes the gag and pol proteins and the second expression vector encodes the envelope protein.

4. claims 21, 33, 35-41, 66-71, 96-101 and, partially, 17-20, 22, 23, 29-32, 34, 42-65, 72-95, 102-106:

An improved method to efficiently transduce mammalian cells with a retroviral supernatant, comprising the steps of:

- 1) growing said population of mammalian cells on culture plates, and

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FURTHER INFORMATION CONTINUED FROM PCT/ISA/210

ii) incubating said supernatant containing replication-defective recombinant retroviral vectors carrying a foreign gene with said population of mammalian cells, to transduce said population of mammalian cells with said foreign gene,

whereby target cells efficiently transduced with said foreign gene are obtained, wherein the improvement comprises adding adhesion molecules or antibodies to adhesion molecules, to culture plates.

5. claims 24-28:

A retroviral vector plasmid for maintenance as a multicopy episome in human cells comprising a selectable marker and a retroviral genome encoding a foreign gene, the Epstein-Barr Virus EBNA1 and oriP sequences.

*) Remark: Incomplete search

Claims 36-41 not searched. These claims refer to a "the method of claim 1", and claim 1 concerns not a method, but plasmids.

We supposed that it must be replaced by "method of claim 35". In this case, the set of claims 36-41 has to be bound to the subject no. 4

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/US 96/13737

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